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Synthesis, biological activity and molecular modelling studies of tricyclic alkylimidazo-, pyrimido- and diazepinopurinediones

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Received: 12 October 2012 / Accepted: 28 February 2013 / Published online: 2 April 2013
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Abstract Syntheses and biological activities of imidazo-, pyrimido- and diazepino[2,1-*f*]purinediones containing *N*-alkyl substituents (with straight, branched or unsaturated chains) are described. Tricyclic derivatives were synthesized by the cyclization of 8-bromo-substituted 7-(2-bromoethyl)-, 7-(3-chloropropyl)- or 7-(4-bromobutyl)-theophylline with primary amines under various conditions. Compound **22** with an ethenyl substituent was synthesized by dehydrohalogenation of 9-(2-bromoethyl)-1,3-dimethyltetrahydropyrimido[2,1-*f*]purinedione. The obtained derivatives (**5–35**) were initially evaluated for their affinity at rat A₁ and A_{2A} adenosine

receptors (AR), showing moderate affinity for both adenosine receptor subtypes. The best ligands were diazepinopurinedione **28** ($K_i=0.28 \mu\text{M}$) with fivefold A_{2A} selectivity and the non-selective A₁/A_{2A} AR ligand pyrimidopurinedione **35** ($K_i A_1=0.28 \mu\text{M}$ and $K_i A_{2A}=0.30 \mu\text{M}$). The compounds were also evaluated for their affinity at human A₁, A_{2A}, A_{2B} and A₃ ARs. All of the obtained compounds were docked to the A_{2A} AR X-ray structure in complex with the xanthine-based, potent adenosine receptor antagonist—XAC. The likely interactions of imidazo-, pyrimido- and diazepino[2,1-*f*]purinediones with the residues forming the A_{2A} binding pocket were discussed. Furthermore, the new compounds were tested *in vivo* as anticonvulsants in maximal electroshock, subcutaneous pentylenetetrazole (ScMet) and TOX tests in mice (*i.p.*). Pyrimidopurinediones showed anticonvulsant activity mainly in the ScMet test. The best derivative was compound **11**, showing 100 % protection at a dose of 100 mg/kg without symptoms of neurotoxicity. Compounds **6**, **7**, **8** and **14** with short substituents showed neurotoxicity and caused death. In rat tests (*p.o.*), **9** was characterized by a high protection index (>13.3). AR affinity did not apparently correlate with the antiepileptic potency of the compounds.

Electronic supplementary material The online version of this article (doi:10.1007/s11302-013-9358-3) contains supplementary material, which is available to authorized users.

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Keywords Tricyclic xanthine derivatives · Adenosine A₁, A_{2A}, A_{2B} and A₃ receptor affinity · Anticonvulsant activity · Molecular modelling studies

Abbreviations

ADP	Antiepileptic Drug Development Program
ARs	Adenosine receptors
CHO	Chinese hamster ovary
CNS	Central nervous system
DMF	Dimethylformamide
DMSO	Dimethylsulfoxide

(^3H)CCPA)	[^3H]2-chloro- N^6 -cyclopentyladenosine
(^3H)PSB-603)	[^3H]8-(4-(4-(4-chlorophenyl)piperazine-1-sulfonyl)phenyl)-1-propylxanthine
(^3H)PSB-11)	[^3H]2-phenyl-8-ethyl-4-methyl-(8 <i>R</i>)-4,5,7,8-tetrahydro-1 <i>H</i> -imidazo[2,1- <i>i</i>]purine-5-one
(^3H)MSX-2)	[^3H]1-propargyl-3-(3-hydroxypropyl)-7-methyl-8-(<i>m</i> -methoxystyryl)xanthine
IR	Infrared
MES	Maximal electroshock
NINDS	National Institute of Neurological Disorders and Stroke
NMR	Nuclear magnetic resonance
PI	Protection index
PDB	Protein data bank
PSB-36	1-Butyl-3-hydroxypropyl-8-noradamantylxanthine
RMSD	Root mean square deviation
ScMet	Subcutaneous pentylenetetrazol
TLC	Thin layer chromatography
UV	Ultra-violet
XAC	Xanthine amine congener

Introduction

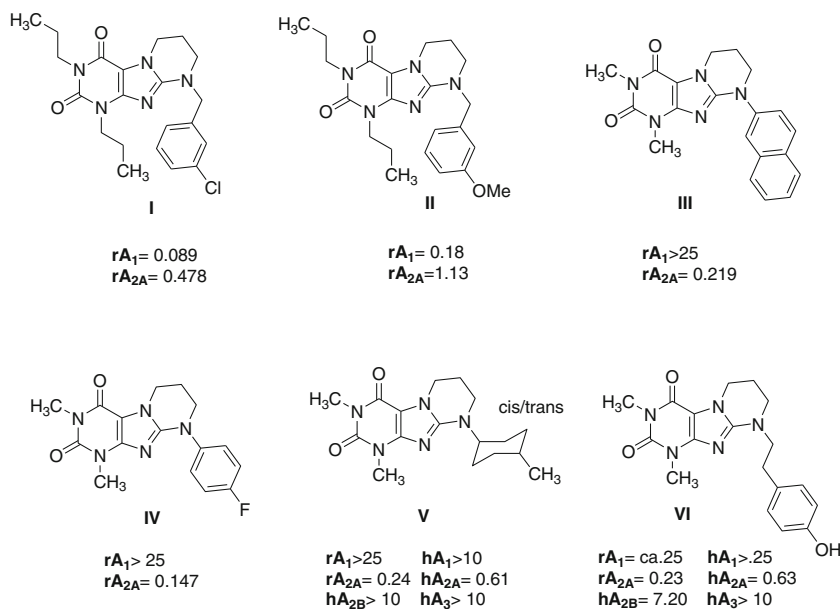
Adenosine, a major constituent of nucleic acids, which consists of the purine base adenine linked to the ribose moiety, has important and diverse effects on many biological processes. Some of the physiological actions of adenosine include effects on heart rate and atrial contractility, vascular smooth muscle tone, release of neurotransmitters, lipolysis, renal function and white blood cell functions [1, 2].

Four adenosine receptor (AR) subtypes are known, A_1 , $\text{A}_{2\text{A}}$, $\text{A}_{2\text{B}}$ and A_3 , all of which were cloned and pharmacologically characterized. The $\text{A}_{2\text{A}}$ and $\text{A}_{2\text{B}}$ receptors are positively coupled to adenylyl cyclase, while A_1 and A_3 adenosine receptors cause inhibition of cAMP formation. Adenosine acts via these different receptor subtypes, the affinity to which ranges from nanomolar (“high affinity” A_1 , 3–30 nM; $\text{A}_{2\text{A}}$ 1–20 nM) to micromolar (“low affinity” $\text{A}_{2\text{B}}$, 5–20 μM ; $\text{A}_3 > 1 \mu\text{M}$) [3, 4]. These receptors belong to the large superfamily of G protein-coupled receptors [2].

Adenosine A_1 receptors are ubiquitously expressed, e.g. in the central nervous system (CNS), especially in the brain, with high levels being expressed in many regions. The distribution of adenosine $\text{A}_{2\text{A}}$ receptors is wide ranging but restricted, including lymphocytes, platelets, brain striatum, vascular smooth muscle and endothelium [2].

The prototypical antagonists of the A_1 adenosine receptor are the xanthines: theophylline and caffeine. Natural xanthines are non-specific adenosine antagonists. They are not selective for any of the adenosine receptor subtypes and have low affinity for the A_1 receptor. Due to their CNS-stimulating effects, A_1 adenosine receptor selective antagonists have been proposed as cognition enhancers for the treatment of dementias, such as Alzheimer’s disease. These receptors have been shown to be involved in sedative, antiseizure and anxiolytic effects. New potential indications are being discovered and investigated: in heart (for the treatment of cardiac arrhythmias and oedemas and as positive inotropic and cardiac protectants), in kidney (for oedemas and nephritis treatment), in lung (for asthma, oedema and lung protection) and in CNS (for depression, stress and coma) diseases. A_1 AR antagonists are being investigated as antihypertensives and potassium-saving diuretics with kidney protective effects, for the treatment of

Fig. 1 Structures and adenosine receptor binding affinities of tricyclic xanthine derivatives (I–VI); K_i values are given in micromolar; *h* human, *r* rat



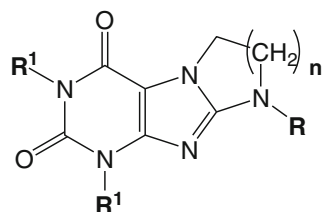
depression and asthma and for the prevention of ischemia-induced injuries [5–7].

In the last years, numerous studies have confirmed the ability of A_{2A} adenosine receptor antagonists to prevent neurodegenerative diseases such as Parkinson's and Alzheimer's diseases, ischemic brain damage and, recently, epilepsy and sensorimotor disorders (restless legs syndrome—RLS) [8–14]. Methylxanthines such as theophylline and caffeine have been known to enhance locomotor activity; however,

these compounds are non-selective antagonists and have weak affinity for A_{2A} AR.

Our efforts were directed towards the development of new selective xanthine adenosine receptor ligands. Our main interest has focussed on the investigation of tricyclic xanthine derivatives [15–19]. The so far most active compounds are shown in Fig. 1. The most potent A_1 AR ligands were found among 1,3-dipropyl-substituted benzylpyrimidopurinediones (**I**, **II**) [17], while A_{2A} adenosine receptor ligands were 1,3-

Table 1 Structures of the tested imidazo-, pyrimido- and diazepino[2,1-*f*]purinediones (5–35)



Cpd	R	R ¹	n	Cpd	R	R ¹	n
5	H	CH ₃	2				
6	CH ₃	CH ₃	2	22		CH ₃	2
7		CH ₃	2	23		CH ₃	2
8		CH ₃	2	24		CH ₃	2
9		CH ₃	2	25	H	CH ₃	3
10		CH ₃	2	26		CH ₃	3
11		CH ₃	2	27		CH ₃	3
12		CH ₃	2	28		CH ₃	3
13		CH ₃	2	29		CH ₃	3
14		CH ₃	2	30		CH ₃	3
15		CH ₃	2	31		CH ₃	3
16		CH ₃	2	32		CH ₃	3
17		CH ₃	2	33	H	CH ₃	1
18		CH ₃	2	34		CH ₃	1
19		CH ₃	2	35		C ₃ H ₇	2

dimethyl-substituted aryl- (**III**, **IV**) [16] cycloalkyl- (**V**) [19] and phenalkylpyrimidopurinediones (**VI**) [18] (Fig. 1). Several of the most active ligands at adenosine A_{2A} AR (e.g. **III**, **IV** and **VI**) [20] were demonstrated to exhibit antiparkinsonian effects. Among pyrimidopurinediones, compounds were found which displayed anticonvulsant properties—protective activity in subcutaneous pentylenetetrazole (ScMet) or in maximal electroshock (MES) and ScMet test; however, the mechanism of this action was not clear [16, 19].

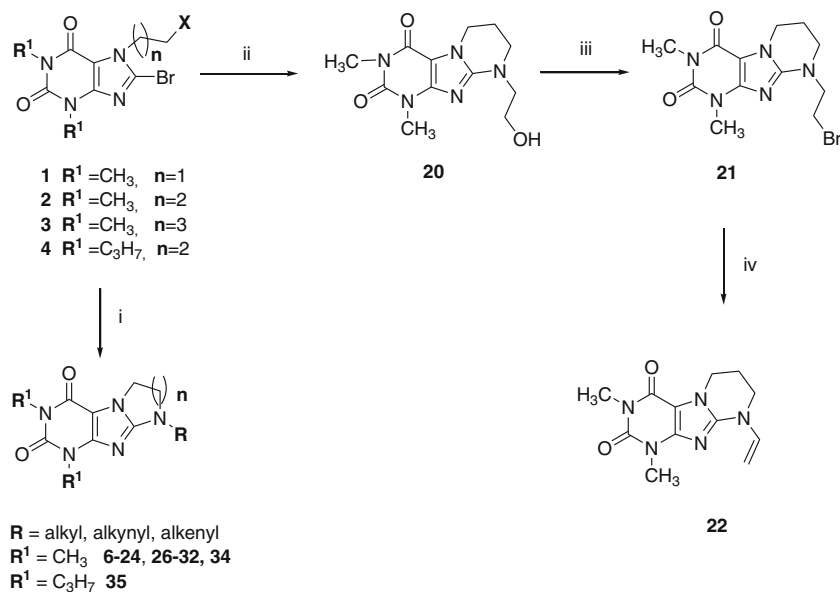
As a continuation of our search for potent adenosine A_1 and especially A_{2A} receptor ligands among cycloalkyl annelated xanthines, we have developed a new series of imidazo-, pyrimido- and diazepino[2,1-*f*]purinedione derivatives possessing aliphatic substituents (open congeners of cycloalkyl derivatives) in the annelated ring, e.g. alkyl, alkynyl and alkenyl chains. The compounds turned out to be selective A_1 or A_{2A} AR antagonists with moderate, submicromolar affinity as shown by radioligand binding studies at native rat receptors. Investigated compounds were examined also for A_{2B} and A_3 AR affinity at human recombinant AR. Additionally, the most active compounds were evaluated for affinity at human recombinant A_1 and A_{2A} AR affinity. Molecular modelling studies were performed to discuss affinity of the compounds through the docking onto the active sites of the A_{2A} adenosine receptor model. Investigated compounds were examined for their anticonvulsant activity as well.

Results and discussion

Chemistry

The synthesis of tricyclic imidazo-, pyrimido- and diazepinopurinediones (Table 1) was accomplished as

Fig. 2 Synthesis of substituted imidazo-, pyrimido- and diazepino[2,1-*f*]purinediones. Reagents and conditions: *i* appropriate primary amines, solvent (EtOH, *n*-PrOH, *n*-BuOH, DMF or without solvent), reflux; *ii* aminoethanol, reflux; *iii* PBr_3 , $CHCl_3$, reflux; *iv* KOH, EtOH, reflux



shown in Fig. 2. As starting material for 1,3-dimethyl-imidazo[2,1-*f*]purinediones (**33**, **34**), 7-(2-bromoethyl)-8-bromotheophylline (**1**) was used, the preparation of which had been described by Caccacae [21]. In our laboratory, a modified procedure was developed using a two-phase catalysis method [16]. The other starting compounds, 7-(3-chloropropyl)-8-bromotheophylline (**2**) for 1,3-dimethyl-pyrimido[2,1-*f*]purinediones (**5–24**) and 7-(4-bromobutyl)-8-bromo-theophylline (**3**) for 1,3-dimethyl-diazepino[2,1-*f*]purinediones (**25–32**), were obtained as previously described [22, 23]. 1,3-Dipropyl-7-(3-chloropropyl)-8-bromoxanthine (**4**) [16, 17] was used as starting material for the synthesis of pyrimido[2,1-*f*]purinedione (**35**). The cyclization reaction with amines possessing straight, branched or unsaturated chains was carried out under various conditions (excess of amine, solvent and different reaction time). The synthesis of compounds **8**, **9** and **23** was described previously [24], but their structures had been confirmed only by UV spectra, and pharmacological tests had not been performed. Unsubstituted compounds **5**, **25** and **33** were previously synthesized in our group [25, 26] and were now subjected to pharmacological tests to compare them with substituted derivatives. Compound **22** with an ethenyl moiety was obtained by dehydrohalogenation of 9-(2-bromoethyl)-1,3-dimethyl-6,7,8,9-tetrahydropyrimido[2,1-*f*]purinedione (**21**) [27] (formed from the appropriate hydroxy ethyl derivative **20** [28]) with ethanolic potassium hydroxide (Fig. 2). The structures of the synthesized compounds were confirmed by UV, IR and 1H NMR spectra: UV spectra showed a bathochromic shift typical for 8-aminoxanthine derivatives with λ_{max} of about 300 nm [29]. The IR absorption bands were typical of xanthine derivatives [30], and in the 1H NMR spectra, the expected chemical shifts were observed. All compounds were purified by recrystallization.

X-ray structure analysis

Among derivatives with various alkyl substituents at N(9) or (10) (Table 1), monocystals of three of them—**10**, **11** and **30**—suitable for X-ray structure analysis could be selected. The structures represent pyrimido- (**10** and **11**) and diazepino- (**30**) purinediones with N1, N3-dimethyl substituents at the xanthine nitrogen atoms. They feature linear (pentyl and hexyl) or branched (isopentyl) substituents at N(9) or (10). The description and discussion of **10**, **11** and **30** spatial properties are attached in the [supplementary section](#).

Pharmacology

All compounds were tested in vitro in radioligand binding assays for affinity to A₁ and A_{2A} adenosine receptors in rat cortical membrane and rat striatal membrane preparations, respectively. The results are presented in Table 2. The non-selective AR ligand caffeine, selective A_{2A} AR antagonist Preladenant (SCH420814) [31] and selective A₁ AR antagonist PSB-36 [31, 32] were included for comparison. Examined compounds were additionally tested for affinity at human recombinant A₁, A_{2A}, A_{2B} and A₃ receptors stably expressed in Chinese hamster ovary (CHO) cells (Table 3).

Table 2 Affinities of imidazo-, pyrimido- and diazepino[2,1-*f*]purinediones at rat A₁ and A_{2A} adenosine receptors

Compound	Adenosine A ₁ receptor (rat brain cortical membranes) vs. [³ H]CCPA <i>K_i</i> ± SEM [μM] (% inhibition ± SEM at 25 μM) (<i>n</i> =3)	Adenosine A _{2A} receptor (rat brain striatal membranes) vs. [³ H]MSX-2 <i>K_i</i> ± SEM [μM] (% inhibition ± SEM at 25 μM) (<i>n</i> =3)	A _{2A} AR selectivity A ₁ /A _{2A}
5	>25 (30±3 %)	5.22±0.90	>5
6	>25 (28±3 %)	4.30±0.67	>6
7	≥25 (42±1 %)	2.65±0.65	>9
8	3.87±0.56	1.29±0.18	3
9	4.32±0.40	1.75±0.55	3
10	3.16±0.33	3.06±1.00	1
11	2.87±0.39	0.82±0.18	4
12	>25 (38±1 %)	4.95±1.32	>5
13	2.69±0.38	0.87±0.03	3
14	≥25 (40±1 %)	4.64±0.85	>5
15	>25 (19±3 %)	8.87±2.04	>3
16	>25 (30±2 %)	5.37±0.32	>5
17	>25 (21±0.3 %)	>25 (37±9 %)	–
18	>25 (30±2 %)	2.38±0.81	>11
19	7.69±0.57	2.03±0.14	>4
22	>25 (30±5 %)	>25 (47±3 %)	–
23	4.40±0.42	2.62±0.72	2
24	≥25 (46±7 %)	7.40±2.52	>3
25	>25 (33±3 %)	>25 (38±9 %)	–
26	2.24±0.90 (71±2 %)	1.73±0.41	1
27	1.24±0.18	0.85±0.08	2
28	1.53±0.08	0.28±0.02	6
29	≥25 (49±0 %)	3.82±0.23	–
30	>25 (30±1 %)	3.51±1.19	>7
31	1.33±0.22	2.27±0.43	0.6
32	1.88±0.40	2.26±0.79	1
33	>25 (12±4 %)	>25 (45±2 %)	–
34	1.12±0.11 (50.77±2.59 %)	3.80±1.30	0.3
35	0.28±0.02	0.30±0.09	1
Caffeine ^[19]	18.8±5.6	32.8±0.09	
Preladenant (SCH420814)	0.0687±0.0087	0.000661±0.000126	104
PSB-36	0.000368±0.000021	0.371±0.049	0.001

Table 3 Affinities of imidazo-, pyrimido- and diazepino[2,1-*f*]purinediones at human recombinant A₁, A_{2A}, A_{2B} and A₃ adenosine receptors

Compound	Adenosine A ₁ receptor (human recombinant) vs. [³ H]CCPA <i>K_i</i> ± SEM [μM] (% inhibition ± SEM at 10 μM) (<i>n</i> =3)	Adenosine A _{2A} receptor (human recombinant) vs. [³ H]MSX-2 <i>K_i</i> ± SEM [μM] (% inhibition ± SEM at 10 μM) (<i>n</i> =3)	Adenosine A _{2B} receptor (human recombinant) vs. [³ H]PSB-603 <i>K_i</i> ± SEM [μM] (% inhibition ± SEM at 10 μM) (<i>n</i> =2)	Adenosine A ₃ receptor (human recombinant) vs. [³ H]PSB-11 <i>K_i</i> ± SEM [μM] (% inhibition ± SEM at 10 μM) (<i>n</i> =2)
5	n.d.	n.d.	>10 (49±1 %)	>10 (10±7 %)
6	n.d.	n.d.	>10 (6±2 %)	>10 (3±3 %)
7	n.d.	n.d.	>10 (12±8 %)	>10 (5±4 %)
8	n.d.	n.d.	>10 (1±2 %)	>10 (10±3 %)
9	n.d.	n.d.	>10 (3±6 %)	>10 (12±2 %)
10	n.d.	n.d.	>10 (−3±2 %)	>10 (21±5 %)
11	3.80±0.91	2.07±0.47	>10 (9±5 %)	>10 (39±1 %)
12	n.d.	n.d.	>10 (−4±3 %)	>10 (8±1 %)
13	1.91±0.27	0.93±0.11	>10 (5±7 %)	>10 (18±2 %)
14	n.d.	n.d.	>10 (−9±4 %)	>10 (14±2 %)
15	n.d.	n.d.	>10 (0±3 %)	>10 (17±1 %)
16	n.d.	n.d.	>10 (1±6 %)	>10 (22±2 %)
17	n.d.	n.d.	>10 (3±7 %)	≥10 (43±4 %)
18	n.d.	n.d.	>10 (−3±5 %)	>10 (32±2 %)
19	n.d.	n.d.	>10 (0±4 %)	2.57±0.53
22	>10 (21±7 %)	>10 (29±8 %)	>10 (1±3 %)	>10 (11±2 %)
23	n.d.	n.d.	>10 (6±4 %)	>10 (17±6 %)
24	n.d.	n.d.	>10 (29±5 %)	>10 (11±2 %)
25	n.d.	n.d.	>10 (9±5 %)	>10 (14±5 %)
26	n.d.	n.d.	>10 (6±5 %)	>10 (12±3 %)
27	2.47±0.10	1.69±0.25	>10 (12±4 %)	>10 (8±3 %)
28	4.13±0.58	1.10±0.16	>10 (10±7 %)	>10 (23±3 %)
29	n.d.	n.d.	>10 (−4±6 %)	>10 (2±4 %)
30	n.d.	n.d.	>10 (−1±3 %)	>10 (27±0 %)
31	n.d.	n.d.	>10 (−1±6 %)	>10 (22±3 %)
32	n.d.	n.d.	>10 (7±3 %)	>10 (7±4 %)
33	n.d.	n.d.	>10 (17±8 %)	>10 (9±3 %)
34	n.d.	n.d.	>10 (16±9 %)	>10 (9±1 %)
35	0.60±0.03	1.39±0.21	1.32±0.23	0.66±0.02
Preladenant (SCH420814)	0.295±0.010	0.000884±0.000232	>1 (25±1 %)	>1 (35±1 %)
PSB-36	0.00397±0.00048	0.332±0.034	0.0486±0.0037	>1 (39±5 %)

n.d. not detected

The following radioligands were used: [³H]2-chloro-N⁶-cyclopentyladenosine ([³H]CCPA) for A₁, [³H]1-propargyl-3-(3-hydroxypropyl)-7-methyl-8-(*m*-methoxystyryl)xanthine ([³H]MSX-2) [33, 34] for A_{2A}, [³H]PSB-603 [³H]8-(4-(4-(4-chlorophenyl)piperazine-1-sulfonyl)phenyl)-1-propylxanthine [35] for A_{2B} and [³H]2-phenyl-8-ethyl-4-methyl-(8*R*)-4,5,7,8-tetrahydro-1*H*-imidazo[2,1-*i*]purine-5-one ([³H]PSB-11) [36] for A₃ binding studies.

Tricyclic xanthine derivatives, previously obtained in our laboratory, showed anticonvulsant activity [15–18]. Therefore, compounds 5–35 were evaluated in vivo as potential anticonvulsants by the ADP (Antiepileptic Drug Development Program of the National Institute of Neurological Disorders and

Stroke NINDS) according to the Antiepileptic Screening Project. Compounds were injected intraperitoneally as a suspension in 0.5 % methylcellulose into the mice and evaluated in the preliminary screenings with at least three dose levels (30, 100 and 300 mg/kg at 0.5- and 4-h time periods). Phase I of the evaluation included three tests: MES, ScMet seizure tests and the rotarod test for neurological toxicity (TOX). The tests were described in detail by Stables and Kupferberg [37–39].

The MES test is a model for generalized tonic-clonic seizures and identifies compounds which prevent seizure spread. The ScMet is a model to test compounds that raise seizure threshold. The minimal motor impairment was measured by the rotarod test. The results are given in Table 4.

Table 4 Anticonvulsant activity and neurotoxicity of imidazo-, pyrimido- and diazepino[2,1-f]purinediones in mice (i.p.)

Compound ^a	MES ^b			ScMet ^{b,c}				Toxicity ^{b,c}				ASP class ^d
	0.25 h	0.5 h	1 h 4 h	0.25 h	0.5 h	1 h	4 h	0.25 h	0.5 h	1 h 4 h		
5												3
6									30 (1/4)	300 ^e		4
7						30 (1/5)			300 ^f	(4/4)		1
8						30 (1/5)			300 ^f	(2/4)		1
9	100 (1/3)				30 (1/4);	100 (4/5)			30 (1/4);	300 ^g	(4/4)	4
10	100 (1/3)				300 (1/1)				100 (3/8);	300 ^g	(4/4)	2
11					100 (5/5)							1
12					30 (1/5);	300 ^h	(1/5)		30 (1/4)			4
13					100 (3/5)				300 (2/4)			1
14					100 (4/5)				300 ^{f,g}	(4/4)		1
15		100 (1/3)			100 (5/5)				300 ^g	(4/4)		1
16	100 (3/3)				100 (5/5)			100 (1/3)				1
17					300 (4/5)		300 ⁱ	(1/1)	100 (1/8)			2
18				100 (1/5)	300 (5/5)		300 (1/1)					1
19		300 (1/1)			30 (1/5)							1
22					100 (3/5)				300 (1/4)			1
23									300 ^g	(1/4)		3
24					j, k				300 ^g	(4/4)		3
25												3
26									300 (4/4)			3
27									300 (3/4)			3
28									300 ^{f,g}	(3/4)		3
29												
30									300 (1/4)			3
31		300 (1/1)			300 (1/1)		1		100 (5/8)			2
32		300 (1/1)							300 (2/4)			2
33									300 (3/4) ^f		300 (1/1) ^g	3
34									100 (6/8) ^{g,m} ;		100 (1/4)	3
									300 (4/4) ^k			
35									100 (2/8) ⁿ ;		300 (2/2) ^f	
									300 (3/4) ⁿ			

^aSuspension in 0.5 % methylcellulose

^bDoses of 30, 100 and 300 mg/kg. The figures in the table indicate the minimum dose whereby activity was demonstrated. The animals were examined 0.5 and 4.0 h after injections were made. For compounds **9**, **10** and **16**, the response was measured after 0.25 h

^cIn anticonvulsant tests, figures, for example, 1/5 mean number of animals protected/number of animals tested, in toxicity test—number of animals exhibiting toxicity/number of animals tested

^dClassification is as follows: 1 anticonvulsant activity at 100 mg/kg or less, 2 anticonvulsant activity at 300 mg/kg, 3 lack of anticonvulsant activity at 300 mg/kg, 4 neurotoxicity at dose 30 mg/kg

^eClonic seizures

^fDeath

^gUnable to grasp rotorod

^hTonic extension

ⁱMyoclonic jerks

^jDeath following tonic extension

^kDeath following clonic seizure

^lDeath following continuous seizure

^mGrooming effect

ⁿPopcorn effect

Table 5 Anticonvulsant activity and neurotoxicity of selected compounds after oral administration (30 or 50 mg/kg) to rats

Compound ^a	MES ^{b,c}					ScMet ^{b,c}					Toxicity ^d
	0.25 h	0.5 h	1 h	2 h	4 h	0.25 h	0.5 h	1 h	2 h	4 h	
9	50 (4/4)					50 (4/4)	50 (4/4)	10 (2/4)	10 (2/4)	10 (3/4)	–
11											–
13	30 (1/4)					30 (1/4)					–
15						50 (1/4)	50 (1/4)	50 (2/4)	50 (2/4)		–
16	30 (1/4)		30 (1/4)		30 (1/4)	50 (1/4)	50 (1/4)	50 (1/4)	50 (1/4)	50 (2/4)	–
22											–
30	30 (2/4)					30 (1/4)					–

^a Form—suspension in 0.5 % methylcellulose

^b Doses of 10, 30 and 50 mg/kg

^c Figures under doses indicate number of animals protected/number of animals tested

^d The dash (–) indicates an absence of toxicity at maximum dose administration (50 mg/kg)

Some compounds (**9**, **11**, **13**, **15**, **16**, **22** and **30**) were also administered orally to rats and examined in the MES, ScMet screen and TOX test (Table 5). Compound **9** was tested also in the hippocampal binding model in rats to evaluate its ability to prevent or modify both the expression and acquisition of focal seizures [39] (Table 6). For two compounds (**9** and **13**), a quantitative test in mice (i.p.) was made (ED₅₀ and TD₅₀ determination). The results of these experiments compared with literature data for valproate [40] are collected in Table 7.

In vitro tests

Synthesized tricyclic xanthine derivatives displayed affinity towards both A₁ and A_{2A} ARs in radioligand binding studies performed at rat brain membranes (Table 2). The most active but not selective A₁ AR ligand was 1,3-dipropylpyrimidopurinedione (**35**). In the group of diazepinopurinediones were found compounds which displayed submicromolar affinity towards adenosine A_{2A} receptors. Diazepinopurinedione **28** (K_i = 0.28 μM) showed moderate A_{2A} AR selectivity (about sixfold). A_{2A} AR ligands with submicromolar affinity were also found among pyrimidopurinediones: compounds **11** and **13** displayed K_i values of 0.82 and 0.87 μM, respectively. N-substituted derivatives displayed higher affinity than unsubstituted analogues (**5**, **25** and **33**). Elongation of the straight *N*-alkyl substituents led to an increase in both A₁ and A_{2A} AR affinities in the group of diazepinopurinediones.

For pyrimidopurinediones, this tendency was not so obvious; it was only observed at A₁ AR. In the case of A_{2A} AR, the most active ones were compounds with 6 (comp. **11**) and 3 (comp. **8**) atom chains. Branched *N*-alkyl, especially α-branched substituents, generally were not favourable for AR affinity. In case of A₁ AR ligands, such modifications led to a loss of affinity (compounds **12**, **14**, **15**, **16** and **18**) or its decrease (compound **19**). A_{2A} AR ligands were also sensitive to these modifications; however, compounds with branched *N*-alkyl substituents still showed micromolar affinity, and β-branched derivative **13** was the most active one with submicromolar affinity. Enlargement of the annelated ring was favourable for both A₁ and A_{2A} AR activities. Diazepinopurinediones with the same *N*-alkyl substituents as pyrimidopurinediones were up to sixfold more potent (compounds **9** and **28**, and **8** and **27**). An exception here was the *N*-isopropyl derivative **34** which possesses a five-membered annelated ring; however, there are only two members of this group for comparison. Introduction of N1,N3-dipropyl substituents into the xanthine core definitely has favourable influence on A₁ AR affinity of *N*-alkylpyrimidopurinediones: compound **35** was 15-fold more potent than its N1,N3-dimethyl analogue **9**. Ligands with unsaturated *N*-substituents (**22**–**24**) displayed only moderate A₁ and A_{2A} AR affinity.

Selected compounds are mainly those that were the most active ones at the A_{2A} AR (**11**, **13**, **22**, **27**, **28** and **35**) and were also tested for affinity to human A₁ and A_{2A} receptors

Table 6 Test results of compound **9**—preliminary hippocampal binding screen in rats (i.p)

Rat ^{a,b}	Seizure pre-drug		Score comp. 9		After discharge pre-drug		Duration (SCCS)	
	Low	High	Low	High	Low	High	Low	High
1	5	–	0	–	47	56	21	–
2	4	5	0	–	32	114	0	–

^a Dose 100 mg/kg

^b Minimal motor impairment

Table 7 Quantitative anticonvulsant activity and neurotoxicity of **9** and **13** and valproate in mice i.p

Compound	TD ₅₀ ^a	ED ₅₀ MES ^b	ED ₅₀ ScMet ^c	PI ^d	
				MES	ScMet
9	137.34 (184.27–222.9) [3.34]	115.63 (100.34–136.05) [8.77]	45.19 (36.48–51.69) [9.35]	1.19	3.04
13	350.18 (258.31–511.58) [5.66]	>500; 0; 0	94.29 (67.32–126.43) [4.64]	<0.7	3.71
Valproate ^e	483	287	209	1.7	2.3

^a Dose (mg/kg) eliciting evidence of minimal neurological toxicity in 50 % of animals; 95 % confidence interval is shown in parentheses; the slope regression line is shown in brackets

^b Dose (mg/kg) eliciting the MES protection in 50 % animals

^c Dose (mg/kg) eliciting the ScMet protection in 50 % animals

^d Protective index—neurotoxic dose (median effective dose)

^e Data from [40, 47]

(Table 3). Generally, the affinity at human recombinant A₁ as well as A_{2A} receptors was worse than that for native rat receptors (in the range of 1.5—threefold lower affinity). Only in the case of isobutylpyrimidopyridone **13**, the affinity for the human A₁ AR was better than that for the rat receptor and almost equal for A_{2A} AR.

Affinity to the human A_{2B} and A₃ receptors of all of the investigated compounds was very weak; only the dipropyl derivative **35** showed micromolar affinity at A_{2B} AR and submicromolar affinity to the human A₃ AR. Only one compound with a double-branched, long chain (**19**) from the group of dimethyl derivatives showed affinity for A₃ AR in the micromolar range.

Two of the most potent rat and human A_{2A} receptor ligands (compounds **28** and **35**) were investigated for their functional properties using cAMP accumulation assay. They were investigated for their potency to inhibit NECA-induced cAMP accumulation in CHO cells expressing the human A_{2A} receptor (Fig. 3). The compounds clearly behaved as competitive antagonists as the concentration–response curve of NECA was shifted to the right in a parallel fashion in their presence. *K_b* values determined in living CHO cells expressing the human adenosine A_{2A} receptor were well in accordance with *K_i* values determined in radioligand binding studies at membrane preparations of the same cell line. Owing to the structural similarity of all compounds in this series, we suppose that they are all antagonists.

Molecular modelling studies

In our previous molecular modelling studies on A₁ and A_{2A} adenosine receptors, the comparison of rhodopsin- and β₂-adrenergic-based homology models through the docking studies was performed [41]. Since that time, a few X-ray structures of A_{2A} AR in complex with various ligands have been reported in the Protein Data Bank [42, 43], among which

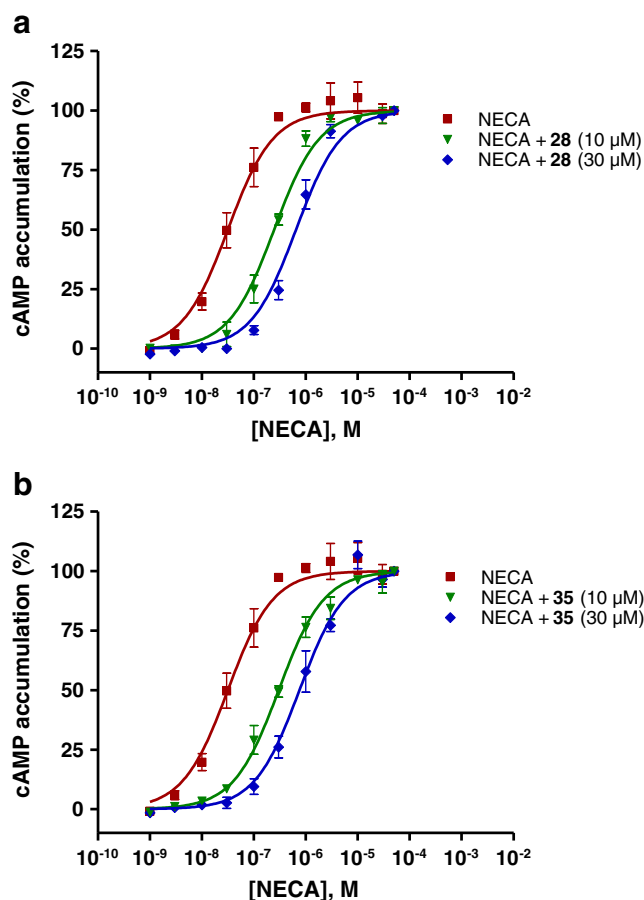


Fig. 3 cAMP accumulation studies in CHO cells expressing the human adenosine A_{2A} receptor. The dose–response curves for the NECA-induced stimulation of cAMP accumulation were generated with NECA in the absence or in the presence of two different concentrations of **28** (a) or **35** (b). Graphs from two independent experiments performed in duplicates with mean values ± SEM are shown. Both investigated compounds shifted the concentration–response curve for NECA in a parallel manner to the right, indicating competitive antagonism. Apparent *K_b* values were as follows: 1,510±20 nM (**28**) and 1,210±130 nM (**35**)

those co-crystallized with xanthines: XAC (PDB code: 3REY) and caffeine (PDB code: 3RFM) are of great importance for our research. Analysis of the ligand binding mode, observed in the crystals 3REY and 3RFM, compared to the binding mode of an inverse agonist ZM241385 bound to A_{2A} AR (PDB codes: 3PWH [42] and 3EML [43]) indicates flexibility of some amino acid side chains within the receptor binding cleft. In particular, the side chain of Asn253 (6.55), described as a crucial residue for ligand binding [44–46], in 3REY is rotated relative to other structures, e.g. 3RFM with caffeine as a ligand. Nevertheless, in both crystal structures of A_{2A} AR co-crystallized with xanthines, the terminal amino group of Asn253 (6.55) forms the hydrogen bond with the same carbonyl group present in purinedione core of ligands.

Due to the structure and the size of synthesized molecules, A_{2A} AR-XAC (3REY) crystal seems to be the best choice as a template for docking. To validate the utilized molecular docking methods, XAC ligand was redocked to its X-ray structure of A_{2A} receptor. In case of simulation without any constraints, the obtained highest ranked pose was distinct from the one in the crystal; superposition of both conformations gave a high RMSD value of 9.68 Å. In this pose, an imidazol nitrogen atom of the ligand plays a role of an H-bond acceptor involved in the contact with the distal amino group of Asn253 (6.55), while a phenyl ring is situated in the proximity of TM6 and ECL2. The docking was then repeated, setting the H-bond interaction between Asn253 (6.55) and one out of carbonyl groups of the ligand as a constraint. This time, the overlay with the experimental binding mode was better (Fig. 4, RMSD=4.53 Å). The superposition of phenylpurinedione cores was almost perfect, giving an RMSD value of 0.42 Å, but calculated and experimental poses differ with orientation of a polar tail of XAC. However, in the crystal structure, the position of this flexible chain is not strictly fixed, as its electron density is not complete [42]. The protocol including constrained H-bond between a ligand and Asn253 (6.55) was chosen for further docking simulations.

As an additional validation to the tested series of xanthines, the structure of caffeine was added and docked together with the rest of the compounds. Docking simulations of the set of all 30 compounds to both templates gave results that can be grouped in three clusters according to the calculated docking score values.

In the two first, highest ranked clusters, the obtained poses adopt a reflected or rotated orientation of the heterocyclic core compared to the XAC conformation from the crystal. In this position, in both cases, the carbonyl group C2=O2 of the purinedione interacts with a side chain amino group of Asn253 (6.55), while C4=O4 either corresponds to the 2-oxo fragment of XAC or points to the top of the receptor, between side chains of Glu169 and Leu267 (7.32).

However, it can be noticed that in both cases of published X-ray structures co-crystallized with xanthines, purinedione

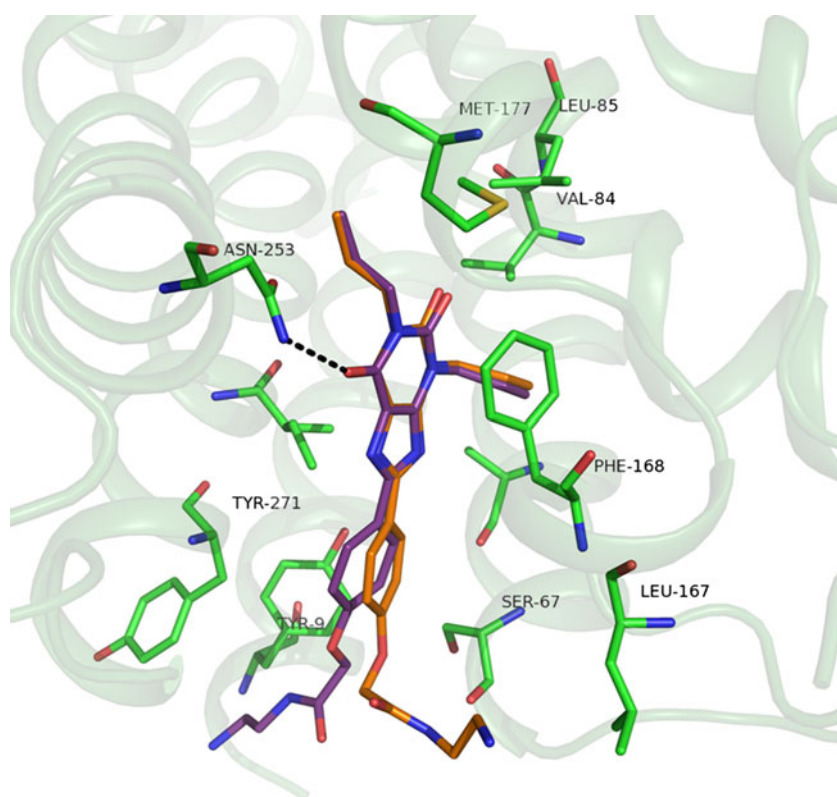
core interacts with Asn253 (6.55) in a similar way, with the same carbonyl oxygen atom (C4=O4) of the ligand, even if Asn253 in this crystal acid adopts two different conformations of the side chain. Taking into account positions of both XAC and caffeine in the binding site, we can presume that the synthesized tricyclic derivatives of xanthine would bind into the A_{2A} receptor in a similar mode (shown in Fig. 5a, b) despite lower docking scores. For this reason, the conformations belonging to the first two clusters were rejected.

The third cluster is created by poses predicted for most of the compounds including caffeine (Fig. 5a, b). Although docking scores are lower than those for the two first sets, this pose is in agreement with the position of the ligand XAC in the crystal structure. The carbonyl group C4=O4 interacts here with the terminal amino group of Asn253 (6.55), and the whole purinedione core of the modelled xanthines overlies the heterocyclic part of XAC with very good RMSD values (from 0.39 to 0.93 Å, for caffeine 0.58 Å), making π - π stacking with the aromatic part of Phe168. Two *N*-methyl (or *N*-propyl) substituents point towards the bottom of the binding site, likewise in case of XAC. The third annelated ring is situated between Phe168 and Glu169 of ECL2 from one side and Ile274 (7.39) and Met270 (7.35) from the other. The alkyl chain is buried in a similar pocket of the binding site to the phenoxy tail of XAC, in the long narrow cleft limited by residues from top fragments of TM2 (Ala63 (2.61), Ile66 (2.64) and Ser67 (2.65)) and TM7 (Leu267 (7.32) and Tyr271 (7.36)) as well as Tyr9 (1.35) (Fig. 5c).

The compounds **25** and **33**, unsubstituted in the annelated ring, were not successfully docked in this position, while long alkyl chains in this position, branched or not, are easily adapted in the groove created by TM2 and TM7. Similarly, longer propyl fragments at 1- and 3-positions of **35** make hydrophobic contacts with residues Leu85 (3.33), Leu249 (6.51) and Met177 (5.38) from one side and Ile66 (2.64), Ala81 (3.29) and Val84 (3.32) from the other, increasing affinity to the receptor relative to short methyl substituents (**35** vs. **9**). Compounds **24** and **30** were not found among poses from this cluster either—the most probable explanation is that the linear structure of the triple bond in **24** or a double-branched chain of **30** causes a steric clash with residues forming the cleft.

The performed docking experiments did not explain in detail the correlation between structure and A_{2A} AR affinity of the obtained xanthines. Nevertheless, it can be stated that both the third annelated ring and alkyl chains as *N*-substituents fit well to the A_{2A} binding pocket, forming additional interactions, and their presence has a big influence on the affinity to the adenosine receptors compared to caffeine—non-selective weak A_{2A} AR antagonist. The similar effect is observed for two *N*-propyl substituents and the phenyl ring of XAC, very potent adenosine receptor antagonist (**35**, Fig. 5b).

Fig. 4 Superposition of the XAC ligand from 3REY crystal structure (purple) and redocked pose of XAC (orange)



In vivo tests

Unsubstituted pyrimido- (**5**) and diazepinopurinedione (**25**) did not show protective activity in both electric and chemical seizures (Table 4). Introduction of *N*-alkyl substituents resulted in anticonvulsant activity in the pyrimidopurinediones, whereas these modifications did not affect in the same way the group of diazepinopurinediones as only compounds **31** and **32** showed protective activity in a dose of 300 mg/kg, but after 4 h caused death of tested animals. Probably, enlargement of the annelated ring in the xanthine derivatives is not favourable for anticonvulsant activity. Unsaturated substituents introduced into the tricyclic xanthine derivatives (**23** and **24**) were also disadvantageous. Generally, *N*-alkyl derivatives showed good protective activity in mice (i.p.), especially in ScMet test in short time (0.5 h). The best compound was *N*-hexylpyrimidopurinedione **11**, with 100 % protection at a dose of 100 mg/kg with no symptoms of neurotoxicity. The other two compounds with long lipophilic chains **18** and **19** also showed good protection: derivative **19** was active in both tests, and ligand **18** displayed protection in ScMet test. Both compounds did not show neurotoxicity. The *N*-butyl and *N*-pentyl derivatives **9** and **10** displayed protective properties in MES and ScMet tests but at the same time showed neurotoxicity. Compounds with short chains (**6**, **7**, **8** and **14**) caused death of test animals. The most toxic substance was *N*-

propargylpyrimidopurinedione **24**, causing death following clonic seizures and tonic extension.

The most active compounds in mice were also examined for activity in rats after oral administration, showing higher seizure protection (Table 5). The best compound was the butyl derivative **9**, showing 100 % protection in ScMet test in the dose of 50 mg/kg in short time (0.25 and 0.5 h) and 50 % protection in dose of 10 mg/kg after 1 2 and 4 h. Ligand **16** was active in MES test at 30 mg/kg and in ScMet test at 50 mg/kg. No symptoms of neurotoxicity were observed in both tested compounds. Ligand **9** was also tested in a hippocampal kindled seizure screen (Table 6). Results suggest the ability of **9** to prevent or modify fully kindled seizures. The *N*-butyl and *N*-isobutyl derivatives **9** and **13** were advanced for phase II of evaluation for quantification of activities (ED_{50} and TD_{50}) against MES- and ScMet-induced seizures in mice (i.p.) (Table 7). These pharmacological parameters were compared with data for valproate [40, 47]. Both tested substances were characterized by a PI_{ScMet} higher than that of valproate, but a lower PI_{MES} .

The anticonvulsant activity of the examined compounds was analysed for correlation with AR affinity. Some coincidence of adenosine A_{2A} affinity and anticonvulsant activity was observed. Compounds showing anticonvulsant activity in ScMet test (**9**, **11**, **13**, **14**, **15**, **16** and **18**) were among the

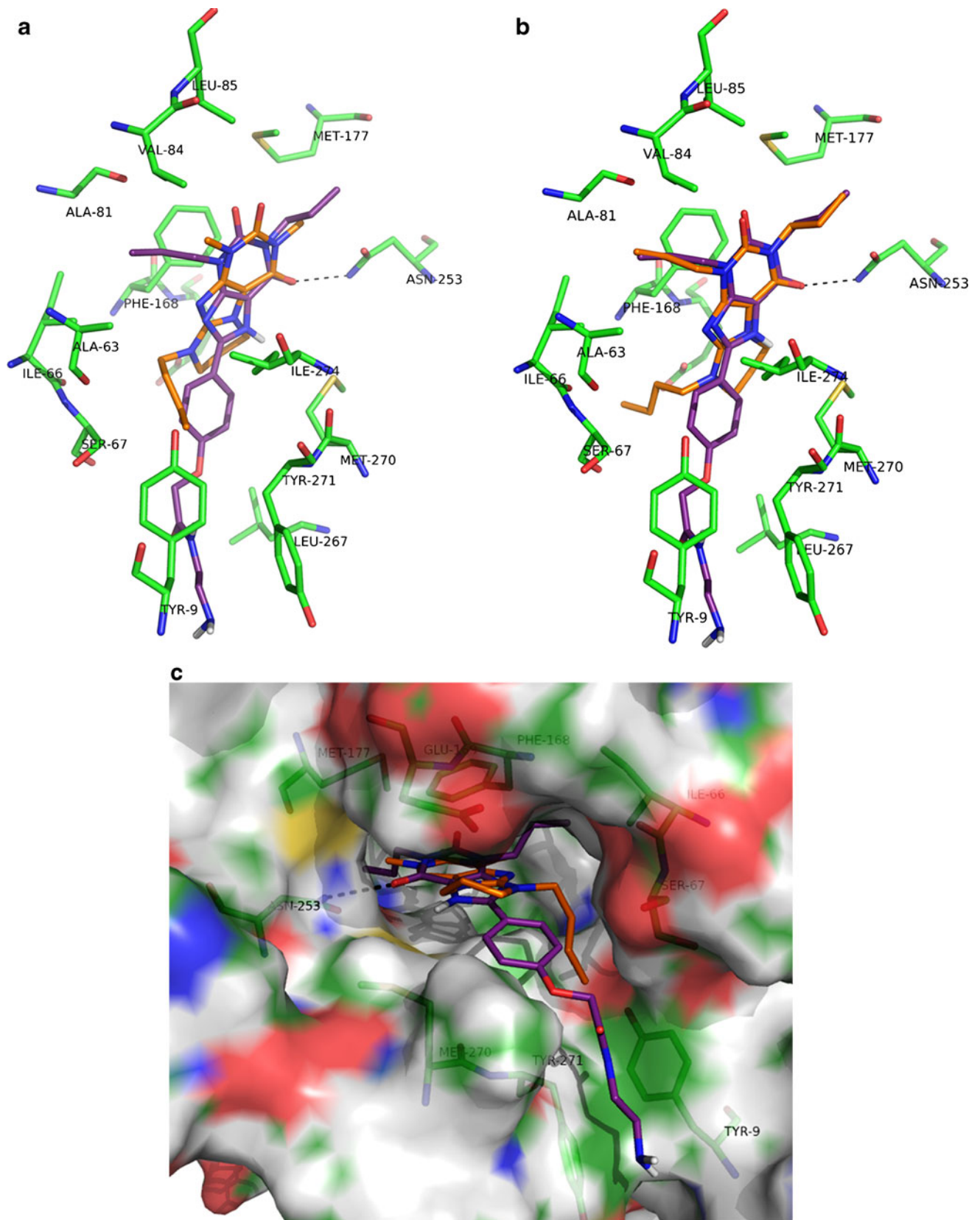


Fig. 5 **a** The model of compound **9** (*orange*) docked to 3REY crystal structure (chosen residues in *green* and XAC ligand in *purple*). **b** The model of compound **35** (*orange*) docked to 3REY crystal structure

(chosen residues in *green* and XAC ligand in *purple*). **c** Superposition of XAC ligand (*purple*) and compound **9** (*orange*) in the binding site of A_{2A} receptor

pyrimidinepurinedione derivatives showing affinity towards A_{2A} AR. However, there were also compounds showing anticonvulsant activity but were devoid of A_{2A} AR affinity. The most active A_{2A} AR ligand, the diazepine derivative **28** and the dipropyl derivative **35** were not active as anticonvulsants. Comparing *in vitro* and *in vivo* activities of the studied tricyclic xanthine derivatives, there is no clear correlation between A_1/A_{2A} AR affinity and anticonvulsant activity of the investigated ligands. So our previous observation that A_{2A} selectivity of AR antagonists may be important for anticonvulsant activity has not been confirmed in the present study [19].

Conclusions

Tricyclic pyrimido- and diazepinopurinediones are moderately potent A_1 and A_{2A} AR antagonists. Enlargement of the annelated ring caused an increase in affinity but not selectivity. Pyrimidopurinediones showed anticonvulsant activity in mice (*i.p.*) and rats (*p.o.*), but lower homologs were toxic. Enlargement of the annelated ring was unprofitable for anticonvulsant activity and led practically to a lack of the activity. Although there is no apparent correlation between anticonvulsant activity and adenosine receptor affinity, it seems that a lipophilic substituent is necessary for both activities.

Acknowledgments The authors wish to acknowledge Professor James P. Stables for providing pharmacological data through the Antiepileptic Drug Development Program National Institutes of Health. Financial support of this research by the Ministry of Science and Higher Education and National Science Centre, Poland (grant No. 4 P05F 014 18, grant No. NN 405 297 836 and grant DEC-2012/04/M/NZ4/00219) and by the BMBF, Germany (01EW0911) in the frame of ERA-NET NEURON is gratefully acknowledged. We would like to thank Mrs. Inge Renner and Katharina Thiemke for the technical assistance in the biological experiments.

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Appendix

Experimental section

Chemistry

Melting points were determined on a MEL-TEMP II apparatus, and they are uncorrected. IR spectra were taken as KBr discs on an FT Jasco IR 410 spectrometer. $^1\text{H-NMR}$ spectra (in CDCl_3) for compounds **10**, **13**, **14** and **22** were

obtained at ambient temperature with a Bruker VM 250, for other compounds with a Varian Mercury 300-MHz spectrometer, using signal of undeuterated solvent as an internal standard. UV spectra were recorded on a Jasco UV/Vis V-530 apparatus in 10^{-5} mole/L in methanol. Elemental analyses (CHN) were performed on an Elementar Vario-EL III apparatus and agreed with theoretical values within $\pm 0.4\%$. TLC data were obtained with Merck silica gel 60 F_{254} aluminium sheets with benzene: acetone (7:3) as developing system. Spots were detected under UV light. Syntheses of compounds **5**, **25** and **33** were described elsewhere [25, 26].

Synthetic procedures and analytical data of tricyclic alkylimidazo-, pyrimido- and diazepinopurinediones

General procedure for the synthesis of 9-substituted 1,3-dimethyl-6,7,8,9-tetrahydropyrimido[2,1-*f*]purine-2,4(1*H*,3*H*)-dione (**6–19**, **23**, **24**). A mixture of (0.66 g, 2 mmol) of 7-(3-chloropropyl)-8-bromotheophylline (**2**) and amine derivative (4–66 mmol) was heated under reflux in the appropriate solvent (ethanol, propanol, butanol and DMF) or without solvent for 5 to 15 h. After cooling, the precipitate was separated. If the crude compound included a little amount of unreacted 7-(3-chloropropyl)-8-bromotheophylline (monitoring by TLC), it was purified by dissolving in 10 % HCl, alkalinizing to pH8 with 10 % NaOH and washing with water. If DMF was used as a reaction medium, the product was precipitated by adding water to the reaction mixture. Compound **17** did not crystallize after cooling and was obtained by removing the excess of 2-propynylamine by steam distillation and cooling the water solution. All compounds were purified by crystallization.

1,3,9-Trimethyl-6,7,8,9-tetrahydropyrimido *[2,1-*f*]purine-2,4(1*H*,3*H*)-dione (6)*

Reaction medium, ethanol; time of heating under reflux, 15 h; excess of amine, **33**. Crystallization with ethanol gave pure compound **6**. Yield 47 %, mp 251–252 °C; TLC: R_f : 0.31; Anal. Calcd for $\text{C}_{11}\text{H}_{15}\text{O}_2\text{N}_5$ (**6**): C, 53.00; H, 6.07; N, 28.10. Found: C, 52.77; H, 5.91; N, 27.82. $^1\text{H-NMR}$ δ (ppm): 2.12–2.20 (m, 2H, $\text{N}_5\text{CH}_2\text{CH}_2\text{CH}_2\text{N}_9$), 3.12 (s, 3H, CH_3), 3.29–3.33 (m, 3H+2H, N_3CH_3 + CH_2N_9), 3.48 (s, 3H, N_1CH_3), 4.19 (t, $J=6.04\text{Hz}$, 2H, N_5CH_2). IR KBr (cm^{-1}): 2,950–2,871— CH_2 , 1,699—CO (2), 1,655 (4), 744— CH_2 . UV λ_{max} , $\log \epsilon$: 300.5, 4.15.

9-Ethyl-1,3-dimethyl-6,7,8,9-tetrahydropyrimido *[2,1-*f*]purine-2,4(1*H*,3*H*)-dione (7)*

Reaction medium, ethanol; time of heating under reflux, 5 h; excess of amine, **29**. Crystallization with ethanol (30 %) gave pure compound **7**. Yield 34 %, mp 215–216 °C; TLC:

R_f : 0.49; Anal. Calcd for $C_{12}H_{17}O_2N_5$ (**7**): C, 54.74; H, 6.52; N, 26.60. Found: C, 55.00; H, 6.52; N, 26.25. 1H -NMR δ (ppm): 21.22 (t, $J=7.14$ Hz, 3H, CH_2CH_3), 2.13–2.20 (m, 2H, $N_5CH_2CH_2CH_2N_9$), 3.34–3.37 (m, 3H+2H, $N_3CH_3+CH_2N_9$), 3.50 (s, 3H, N_1CH_3), 3.54–3.63 (m, 2H, CH_2CH_3), 4.20 (t, $J=6.04$ Hz, 2H, N_5CH_2). IR KBr (cm^{-1}): 2,967–2,872— CH_2 , 1,697—CO (2), 1,666 (4), 784— CH_2 . UV λ_{max} , $\log\epsilon$: 301.5, 4.28.

1,3-Dimethyl-9-propyl-6,7,8,9-tetrahydropyrimido[2,1-f] purine-2,4(1H,3H)-dione (**8**)

Reaction medium, ethanol; time of heating under reflux, 5 h; excess of amine, 12. Crystallization with ethanol (30 %) gave pure compound **8**. Yield 60 %, mp 188–190 °C; TLC: R_f : 0.52; Anal. Calcd for $C_{13}H_{19}O_2N_5$ (**8**): C, 56.31; H, 6.91; N, 21.25. Found: C, 56.25; H, 6.97; N, 24.96. 1H -NMR δ (ppm): 0.95 (t, $J=7.42$ Hz, 3H, CH_2CH_3), 1.60–1.83 (m, 2H, $CH_2CH_2CH_3$), 2.11–2.19 (m, 2H, $N_5CH_2CH_2CH_2N_9$), 3.34–3.46 (m, 3H+2H, $N_3CH_3+CH_2N_9$), 3.49–3.59 (m, 2H+3H, $CH_2CH_3+N_1CH_3$), 4.21 (t, $J=5.91$ Hz, 2H, N_5CH_2). IR KBr (cm^{-1}): 2,958–2,871— CH_2 , 1,697—CO (2), 1,664 (4), 784— CH_2 . UV λ_{max} , $\log\epsilon$: 302, 4.34.

9-Butyl-1,3-dimethyl-6,7,8,9-tetrahydropyrimido[2,1-f] purine-2,4(1H,3H)-dione (**9**)

Reaction medium, solvent-free; time of heating under reflux, 5 h; excess of amine, 15. Crystallization with ethanol gave pure compound **9**. Yield 81 %, mp 174–176 °C; TLC: R_f : 0.37; Anal. Calcd for $C_{14}H_{21}O_2N_5$ (**9**): C, 57.71; H, 7.27; N, 24.04. Found: C, 57.63; H, 7.08; N, 23.84. 1H -NMR δ (ppm): 0.95 (t, $J=7.28$ Hz, 3H, $(CH_2)_3CH_3$), 1.31–1.38 (m, 2H, CH_2CH_3), 1.54–1.62 (m, 2H, $CH_2CH_2CH_3$), 2.09–2.17 (m, 2H, $N_5CH_2CH_2CH_2N_9$), 3.31–3.34 (m, 3H+2H, $N_3CH_3+CH_2N_9$), 3.48–3.53 (m, 2H+3H, $CH_2CH_3+N_1CH_3$), 4.19 (t, $J=6.04$ Hz, 2H, N_5CH_2). IR KBr (cm^{-1}): 2,958–2,873— CH_2 , 1,702—CO (2), 1,666 (4), 784— CH_2 . UV λ_{max} , $\log\epsilon$: 302, 4.25.

1,3-Dimethyl-9-pentyl-6,7,8,9-tetrahydropyrimido[2,1-f] purine-2,4(1H,3H)-dione (**10**)

Reaction medium, solvent-free; time of heating under reflux, 5 h; excess of amine, 20. Crystallization with ethanol (80 %) gave pure compound **10**. Yield 59 %, mp 183–185 °C; TLC: R_f : 0.43; Anal. Calcd for $C_{15}H_{23}O_2N_5$ (**10**): C, 58.99; H, 7.60; N, 22.93. Found: C, 58.76; H, 7.26; N, 23.08. 1H -NMR δ (ppm): 0.92 (t, $J=6.25$ Hz, 3H, $(CH_2)_4CH_3$), 1.27–1.65 (m, 6H, $(CH_2)_3CH_3$), 2.10–2.19 (m, 2H, $N-CH_2CH_2CH_2N_9$), 3.33–3.37 (m, 3H+2H, $N_3CH_3+CH_2N_9$), 3.46–3.54 (m, 2H+3H, $CH_2CH_3+N_1CH_3$), 4.24 (t, $J=6.25$ Hz, 2H, N_5CH_2). IR KBr

(cm^{-1}): 2,952–2,869— CH_2 , 1,699—CO (2), 1,662 (4), 784— CH_2 . UV λ_{max} , $\log\epsilon$: 301, 4.34.

9-Hexyl-1,3-dimethyl-6,7,8,9-tetrahydropyrimido[2,1-f] purine-2,4(1H,3H)-dione (**11**)

Reaction medium, solvent-free; time of heating under reflux, 5 h; excess of amine, 12. Crystallization with ethanol gave pure compound **11**. Yield 57 %, mp 149 °C; TLC: R_f : 0.55; Anal. Calcd for $C_{16}H_{25}O_2N_5$ (**11**): C, 60.16; H, 7.89; N, 21.93. Found: C, 60.13; H, 3.94; N, 22.04. 1H -NMR δ (ppm): 0.90 (t, $J=5.77$ Hz, 3H, $(CH_2)_5CH_3$), 1.33–1.64 (m, 8H, $(CH_2)_4CH_3$), 2.11–2.18 (m, 3H+2H, $N_3CH_3+CH_2N_9$), 3.49–3.53 (m, 2H+3H, $CH_2CH_3+N_1CH_3$), 4.21 (t, $J=6.04$ Hz, 2H, N_5CH_2). IR KBr (cm^{-1}): 2,952–2,869— CH_2 , 1,698—CO (2), 1,664 (4), 742— CH_2 . UV λ_{max} , $\log\epsilon$: 301.5, 4.32.

9-Isopropyl-1,3-dimethyl-6,7,8,9-tetrahydropyrimido[2,1-f] purine-2,4(1H,3H)-dione (**12**)

Reaction medium, propanol; time of heating under reflux, 5 h; excess of amine, 12. Crystallization with ethanol (30 %) gave pure compound **12**. Yield 65 %, mp 241–242 °C; TLC: R_f : 0.48; Anal. Calcd for $C_{13}H_{19}O_2N_5$ (**12**): C, 56.31; H, 6.91; N, 25.25. Found: C, 56.10; H, 6.58; N, 25.05. 1H -NMR δ (ppm): 1.20 (d, $J=6.86$ Hz, 6H, $CH(CH_3)_2$), 2.05–2.13 (m, 2H, $N_5CH_2CH_2CH_2N_9$), 3.26 (s, 3H, N_3CH_3), 3.48 (s, 3H, N_1CH_3), 4.17 (t, $J=6.04$ Hz, 3H, N_1CH_3), 4.62–4.71 (m, 1H, N_9CH). IR KBr (cm^{-1}): 2,967–2,848— CH_2 , 1,704—CO (2), 1,662 (4), 742— CH_2 . UV λ_{max} , $\log\epsilon$: 302, 4.29.

9-Isobutyl-1,3-dimethyl-6,7,8,9-tetrahydropyrimido[2,1-f] purine-2,4(1H,3H)-dione (**13**)

Reaction medium, solvent-free; time of heating under reflux, 5 h; excess of amine, 25. Crystallization with ethanol (50 %) gave pure compound **13**. Yield 68 %, mp 213–215 °C; TLC: R_f : 0.44; Anal. Calcd for $C_{14}H_{21}O_2N_5$ (**13**): C, 57.71; H, 7.27; N, 24.04. Found: C, 58.05; H, 6.99; N, 23.79. 1H -NMR δ (ppm): 0.94 (d, $J=6.50$ Hz, 6H, $CH(CH_3)_2$), 2.01–2.19 (m, 1H+2H, $CH(CH_3)_2+N_5CH_2CH_2CH_2N_9$), 3.31–3.37 (m, 2H+2H, $CH_2N_9CH_2$), 3.39 (s, 3H, N_3CH_3), 3.50 (s, 3H, N_1CH_3), 4.22 (t, $J=6.00$ Hz, 2H, N_5CH_2). IR KBr (cm^{-1}): 2,964–2,844— CH_2 , 1,698—CO (2), 1,662 (4), 742— CH_2 . UV λ_{max} , $\log\epsilon$: 301.5, 4.34.

9-Sec-butyl-1,3-dimethyl-6,7,8,9-tetrahydropyrimido[2,1-f] purine-2,4(1H,3H)-dione (**14**)

Reaction medium, solvent-free; time of heating under reflux, 5 h; excess of amine, 25. Crystallization with ethanol

(50 %) gave pure compound **14**. Yield 98 %, mp 188–190 °C; TLC: R_f : 0.48; Anal. Calcd for $C_{14}H_{21}O_2N_5$ (**14**): C, 57.71; H, 7.27; N, 24.04. Found: C, 57.90; H, 7.43; N, 23.87. 1H -NMR δ (ppm): 0.90 (t, $J=7.38$ Hz, 3H, CH_2CH_3), 1.19 (d, $J=6.60$ Hz, 3H, $=CHCH_3$), 2.06–2.16 (m, 2H, $N_5CH_2CH_2CH_2N_9$), 3.22–3.27 (m, 2H, CH_2N_9), 3.37 (s, 3H, N_3CH_3), 3.50 (s, 3H, N_1CH_3), 4.18–4.24 (m, 2H, N_5CH_2), 4.39–4.48 (m, 1H, $N_1CH=$). IR KBr (cm^{-1}): 2,969–2,875— CH_2 , 1,704—CO(2), 1,652 (4), 744— CH_2 . UV λ_{max} , $\log\epsilon$: 302.5, 4.26.

1,3-Dimethyl-9-(pentan-2-yl)-6,7,8,9-tetrahydropyrimido[2,1-f]purine-2,4(1H,3H)-dione (15)

Reaction medium, DMF; time of heating under reflux, 10 h; excess of amine, 4. Crystallization with ethanol/ H_2O gave pure compound **15**. Yield 91 %, mp 143–144 °C; TLC: R_f : 0.65; Anal. Calcd for $C_{15}H_{23}O_2N_5$ (**15**): C, 59.00; H, 7.60; N, 22.94. Found: C, 58.90; H, 7.57; N, 22.99. 1H -NMR δ (ppm): 0.93 (t, $J=7.29$ Hz, 3H, CH_2CH_3), 1.17 (d, $J=6.88$ Hz, 3H, CH_3CH), 1.25–1.59 (m, 4H, $CH_2CH_2CH_3$), 2.05–2.13 (m, 2H, $N_5CH_2CH_2CH_2N_9$), 3.20–3.25 (m, 2H, CH_2N_9), 3.35 (s, 3H, N_3CH_3), 3.50 (s, 3H, N_1CH_3), 4.18–4.24 (m, 2H, N_5CH_2), 3.49 (s, 3H, N_1CH_3), 4.14–4.24 (m, 2H, N_5CH_2), 4.51–4.58 (m, 1H, $N_1CH=$). IR KBr (cm^{-1}): 2,963–2,872— CH_2 , 1,697—CO (2), 1,666 (4), 743— CH_2 . UV λ_{max} , $\log\epsilon$: 302, 4.28.

1,3-Dimethyl-9-(4-methylpentan-2-yl)-6,7,8,9-tetrahydropyrimido[2,1-f]purine-2,4(1H,3H)-dione (16)

Reaction medium, DMF; time of heating under reflux, 5 h; excess of amine, 2. Crystallization with ethanol/ H_2O gave pure compound **16**. Yield 59 %, mp 116–117 °C; TLC: R_f : 0.58; Anal. Calcd for $C_{16}H_{25}O_2N_5$ (**16**): C, 60.16; H, 7.89; N, 21.93. Found: C, 60.22; H, 8.01; N, 21.96. 1H -NMR δ (ppm): 0.89–0.94 (m, 6H, $CH(CH_3)_2$), 1.15 (d, $J=6.60$ Hz, 3H, N_9CHCH_3), 1.19–1.26 (m, 1H, $CH(CH_3)_2$), 1.40–1.58 (m, 2H, $CH_2CH(CH_3)_2$), 2.03–2.15 (m, 2H, $N_5-CH_2-CH_2-CH_2-N_9$), 3.14–3.20 (m, 2H, CH_2N_9), 3.33 (s, 3H, N_3CH_3), 3.47 (s, 3H, N_1CH_3), 4.12–4.22 (m, 2H, N_5CH_2), 4.57–4.67 (m, 1H, N_9CHCH_3). IR KBr (cm^{-1}): 2,952–2,867— CH_2 , 1,699—CO (2), 1,660 (4), 742— CH_2 . UV λ_{max} , $\log\epsilon$: 301.5, 4.21.

1,3-Dimethyl-9-(2-methylhexyl)-6,7,8,9-tetrahydropyrimido[2,1-f]purine-2,4(1H,3H)-dione (17)

Reaction medium, DMF; time of heating under reflux, 5 h; excess of amine, 4. Crystallization with ethanol/ H_2O gave pure compound **17**. Yield 81 %, mp 122–124 °C; TLC: R_f : 0.63; Anal. Calcd for $C_{17}H_{27}O_2N_5$ (**17**): C, 61.24; H, 8.17; N, 21.01. Found: C, 61.51; H, 8.19; N, 21.12. 1H -NMR δ (ppm): 0.84–0.89 (m, 6H, 2 CH_3), 1.18–1.59 (m, 8H, $CH_2CH(CH_2)_3$

CH_2CH_3), 1.40–1.58 (m, 2H, $CH_2CH(CH_3)_2$), 2.06–2.13 (m, 2H, $N_5CH_2CH_2CH_2N_9$), 3.17–3.21 (m, 2H, CH_2N_9), 3.36 (s, 3H, N_3CH_3), 3.48 (s, 3H, N_1CH_3), 4.21 (t, $J=6.05$ Hz, 2H, N_5CH_2), 4.29–4.35 (m, 1H, N_9CH). IR KBr (cm^{-1}): 2,960–2,861— CH_2 , 1,699—CO (2), 1,659 (4), 750— CH_2 . UV λ_{max} , $\log\epsilon$: 302, 4.31.

9-(Heptan-2-yl)-1,3-dimethyl-6,7,8,9-tetrahydropyrimido[2,1-f]purine-2,4(1H,3H)-dione (18)

Reaction medium, DMF; time of heating under reflux, 10 h; excess of amine, 2. Crystallization with ethanol/ H_2O gave pure compound **18**. Yield 57 %, mp 94–96 °C; Anal. Calcd for $C_{17}H_{27}O_2N_5$ (**18**): C, 61.24; H, 8.17; N, 21.01. Found: C, 60.95; H, 7.90; N, 21.05. 1H -NMR δ (ppm): 0.84–0.94 (m, 3H, $(CH_2)_4-CH_3$), 1.17 (d, $J=6.87$ Hz, 3H, $CHCH_3$), 1.26–1.59 (m, 8H, $(CH_2)_4CH_3$), 2.05–2.13 (m, 2H, $N_5CH_2CH_2CH_2N_9$), 3.20–3.25 (m, 2H, CH_2N_9), 3.35 (s, 3H, N_3CH_3), 3.49 (s, 3H, N_1CH_3), 4.13–4.24 (m, 2H, N_5CH_2), 4.50–4.56 (m, 1H, N_9CH). IR KBr (cm^{-1}): 2,955–2,859— CH_2 , 1,701—CO (2), 1,656 (4), 745— CH_2 . UV λ_{max} , $\log\epsilon$: 302, 4.30.

1,3-Dimethyl-9-(6-methylheptan-2-yl)-6,7,8,9-tetrahydropyrimido[2,1-f]purine-2,4(1H,3H)-dione (19)

Reaction medium, DMF; time of heating under reflux, 5 h; excess of amine, 2. Crystallization with ethanol/ H_2O gave pure compound **19**. Yield 60 %, mp 99–101 °C; TLC: R_f : 0.48; Anal. Calcd for $C_{18}H_{29}O_2N_5$ (**19**): C, 61.24; H, 8.17; N, 21.01. Found: C, 61.99; H, 8.21; N, 20.19. 1H -NMR δ (ppm): 0.83–0.86 (m, 6H, $CH(CH_3)_2$), 1.18 (d, $J=6.87$ Hz, 3H, N_9CHCH_3), 1.23–1.38 (m, 1H, $CH(CH_3)_2$), 1.43–1.61 (m, 6H, $(CH_2)_3$), 2.06–2.14 (m, 2H, CH_2N_9), 3.36 (s, 3H, N_3CH_3), 3.50 (s, 3H, N_1CH_3), 4.12–4.27 (m, 2H, N_5CH_2), 4.46–4.58 (m, 1H, N_9CHCH_3). IR KBr (cm^{-1}): 2,956–2,858— CH_2 , 1,708—CO (2), 1,651 (4), 742— CH_2 . UV λ_{max} , $\log\epsilon$: 302.5, 4.25.

9-Allyl-1,3-dimethyl-6,7,8,9-tetrahydropyrimido[2,1-f]purine-2,4(1H,3H)-dione (23)

Reaction medium, solvent-free; time of heating under reflux, 5 h; excess of amine, 20. Crystallization with ethanol (10 %) gave pure compound **23**. Yield 65 %, mp 157–159 °C; TLC: R_f : 0.35; Anal. Calcd for $C_{13}H_{17}O_2N_5$ (**23**): C, 56.71; H, 6.22; N, 24.44. Found: C, 56.88; H, 6.48; N, 25.12. 1H -NMR δ (ppm): 2.10–2.18 (m, 2H, $N_5CH_2CH_2CH_2N_9$), 3.29 (t, $J=5.65$ Hz, 2H, CH_2N_9), 3.34 (s, 3H, N_3CH_3), 3.48 (s, 3H, N_1CH_3), 4.16 (t, $J=6.04$ Hz, 2H, N_9CH_2), 4.20 (t, $J=6.04$ Hz, 2H, N_5CH_2), 5.20–5.26 (m, 2H, $CH_2=CH$), 5.75–5.88 (m, 1H, $CH_2=CH$). IR KBr (cm^{-1}): 3,081— $CH_2-CH=CH_2$, 2,944–2,902 CH_2 , 1,695—CO (2), 1,664 (4), 750— CH_2 . UV λ_{max} , $\log\epsilon$: 303.5, 4.30.

1,3-Dimethyl-9-(prop-2-ynyl)-6,7,8,9-tetrahydropyrimido[2,1-f]purine-2,4(1H,3H)-dione (24)

Reaction medium, n-butanol; time of heating under reflux, 10 h; excess of amine, 5. Crystallization with ethanol (30 %) gave pure compound **24**. Yield 93 %, mp 203–206 °C; TLC: R_f : 0.57; Anal. Calcd for $C_{13}H_{15}O_2N_5$ (**24**): C, 57.14; H, 5.54; N, 25.62. Found: C, 57.44; H, 5.66; N, 25.79. 1H -NMR δ (ppm): 2.18–2.29 (m, 3H, $N_5CH_2CH_2CH_2+CH\equiv$), 3.35 (s, 3H, N_3CH_3), 3.35–3.42 (m, 2H, CH_2N_9), 3.50 (s, 3H, N_1CH_3), 4.36 (t, $J=6.05$ Hz, 2H, N_9CH_2). IR KBr (cm^{-1}): 3,123— $CH_2-CH=CH_2$, 2,952— CH_2 , 1,701—CO (2), 1,665 (4), 744— CH_2 . UV λ_{max} , $\log\epsilon$: 296, 4.19.

Synthesis of 9-ethenyl 1,3-dimethyl-6,7,8,9-tetrahydropyrimido[2,1-f]purine-2,4(1H,3H)-dione (22)

A mixture of 9-(2-bromoethyl)-1,3-dimethyl-6,7,8,9-tetrahydropyrimido[2,1-f]purine-2,4(1H, 3H)-dione (0.68 g, 2 mmol) (**21**) [27] and KOH (0.11 g, 2 mmol) in 10 ml ethanol was heated at reflux for 20 h. After cooling, the precipitate was separated and washed with water and crystallized from methoxyethanol. Yield 70 %, mp 268–271 °C; TLC: R_f : 0.55; Anal. Calcd for $C_{12}H_{15}O_2N_5$ (**22**): C, 55.19; H, 5.79; N, 26.80. Found: C, 55.47; H, 5.55; N, 26.45. 1H -NMR δ (ppm): 2.17–2.28 (m, 2H, $N_5CH_2CH_2CH_2N_9$), 3.38 (s, 3H, N_3CH_3), 3.53 (s, 3H, N_1CH_3), 3.54–3.59 (m, 2H, N_9CH_2), 4.26–4.52 (m, 4H, $N_5CH_2+CH_2=$), 7.38–7.48 (m, 1H, $CH=$). IR KBr (cm^{-1}): 3,108–3,089— $CH=CH_2$, 2,983–2883 CH_2 , 1,701—CO (2), 1,652 (4), 746— CH_2 . UV λ_{max} , $\log\epsilon$: 304, 4.40.

General procedure for the synthesis of 10-substituted 1,3-dimethyl-6,7,8,9-tetrahydrodiazepino[2,1-f]purine-2,4(1H,3H)-diones (**26–32**)

A mixture of 7-(4-bromobutyl)-8-bromotheophylline (0.79 g, 2 mmol) (**3**) [23] and an amine derivative (4–72 mmol) was heated under reflux without solvent or with ethanol, isobutanol or DMF for 5–10 h. Compounds **30** and **31** precipitated by adding water to the reaction mixture; compounds **26** and **27** were obtained by removing the solvent and excess of amine by distillation under reduced pressure and adding water to the residue. Compounds **28**, **29** and **32** precipitated by removing the excess of amine by steam distillation and cooling the water solution. All compounds were purified by crystallization from diluted ethanol or by dissolving in ethanol and precipitation with water.

10-Ethyl-1,3-dimethyl-6,7,8,9-tetrahydrodiazepino[2,1-f]purine-2,4(1H,3H)-dione (26)

Reaction medium, ethanol; time of heating under reflux, 10 h; excess of amine, 36. Crystallization with ethanol

(30 %) gave pure compound **26**. Yield 38 %, mp 124–125 °C; TLC: R_f : 0.62; Anal. Calcd for $C_{13}H_{19}O_2N_5$ (**26**): C, 56.29; H, 6.91; N, 25.24. Found: C, 56.08; H, 6.64; N, 25.50. 1H -NMR δ (ppm): 1.22–1.26 (t, $J=7.01$ Hz, 3H, CH_3CH_2), 1.27–1.92 (s, 4H, $N_5CH_2(CH_2)_2CH_2$), 3.30–3.23 (m, 2H, CH_2CH_3), 3.37 (s, 3H, N_3CH_3), 3.51 (s, 3H, N_1CH_3), 3.54–3.56 (m, 2H, CH_2N_{10}), 4.28–4.31 (m, 2H, N_5CH_2). IR KBr (cm^{-1}): 2,952–2873— CH_2 , 1,693—CO(2), 1,661 (4), 740— CH_2 . UV λ_{max} , $\log\epsilon$: 300, 4.22.

1,3-Dimethyl-10-propyl-6,7,8,9-tetrahydrodiazepino[2,1-f]purine-2,4(1H,3H)-dione (27)

Reaction medium, isobutanol; time of heating under reflux, 10 h; excess of amine, 11. Crystallization with ethanol (30 %) gave pure compound **27**. Yield 46 %, mp 153–154 °C; TLC: R_f : 0.64; Anal. Calcd for $C_{14}H_{21}O_2N_5$ (**27**): C, 57.71; H, 7.27; N, 24.03. Found: C, 57.76; H, 6.98; N, 23.66. 1H -NMR δ (ppm): 0.91 (t, $J=7.43$ Hz, 3H, $(CH_2)_2CH_3$), 1.57–1.73 (m, 2H, $CH_2CH_2CH_3$), 1.87–1.91 (m, 4H, $N_5CH_2(CH_2)_2CH_2N_{10}$), 3.24 (t, $J=5.45$ Hz, 2H, $CH_2CH_2CH_3$), 3.37 (s, 2H, N_3CH_3), 3.42–3.47 (s, 2H, CH_2N_{10}), 3.51 (s, 3H, N_1CH_3), 4.31 (t, $J=5.08$ Hz, 2H, N_5CH_2). IR KBr (cm^{-1}): 2,935–2870— CH_2 , 1,692—CO(2), 1,653 (4), 750— CH_2 . UV λ_{max} , $\log\epsilon$: 300, 4.30.

10-Butyl-1,3-dimethyl-6,7,8,9-tetrahydrodiazepino[2,1-f]purine-2,4(1H,3H)-dione (28)

Reaction medium, solvent-free; time of heating under reflux, 5 h; excess of amine, 25. Crystallization with ethanol/ H_2O gave pure compound **28**. Yield 72 %, mp 84–86 °C; TLC: R_f : 0.70; Anal. Calcd for $C_{15}H_{23}O_2N_5$ (**28**): C, 58.99; H, 7.60; N, 22.93. Found: C, 58.89; H, 7.22; N, 22.60. 1H -NMR δ (ppm): 0.95 (t, $J=7.28$ Hz, 3H, $(CH_2)_3CH_3$), 1.30–1.42 (m, 2H, $CH_2CH_2CH_2CH_3$), 1.57–1.67 (m, 2H, $CH_3CH_2CH_2CH_3$), 1.87–1.88 (m, 4H $N_5CH_2(CH_2)_2CH_2N_{10}$), 3.23 (t, $J=5.01$ Hz, 2H $N_{10}CH_2$), 3.37 (s, 3H, N_3CH_3), 3.48 (t, $J=7.48$ Hz, 2H, CH_2N_{10}), 3.51 (s, 3H, N_1CH_3), 4.30 (t, $J=5.09$ Hz, 2H, N_5CH_2). IR KBr (cm^{-1}): 2,957–2,871— CH_2 , 1,700—CO(2), 1,658 (4), 745— CH_2 . UV λ_{max} , $\log\epsilon$: 300, 4.28.

10-Sec-butyl-1,3-dimethyl-6,7,8,9-tetrahydrodiazepino[2,1-f]purine-2,4(1H,3H)-dione (29)

Reaction medium, solvent-free; time of heating under reflux, 10 h; excess of amine, 10. Crystallization with ethanol (50 %) gave pure compound **29**. Yield 57 %, mp 138–140 °C; TLC: R_f : 0.53; Anal. Calcd for $C_{15}H_{23}O_2N_5$ (**29**): C, 58.99; H, 7.60; N, 22.93. Found: C, 58.67; H, 7.30; N, 22.75. 1H -NMR δ (ppm): 0.95 (t, $J=1.10$ Hz, 3H, CH_2CH_3), 1.22 (d, $J=7.60$ Hz, 3H, $CHCH_3$), 1.45–1.72

(m, 2H, CH₃CH₂), 1.72–2.00 (m, 4H, N₅CH₂(CH₂)₂CH₂N₁₀), 2.98–3.28 (m, 2H, CH₂N₁₀), 3.37 (s, 2H, N₃CH₃), 3.48 (t, *J*=7.48 Hz, 2H, CH₂N₁₀), 3.51 (s, 3H, N₁CH₃), 4.01–4.12 (m, 2H, N₅CH₂), 4.51–4.58 (m, 1H, CHCH₃). IR KBr (cm⁻¹): 2,966–2,876—CH₂, 1,701—CO(2), 1,660 (4), 745—CH₂. UV λ_{max}, logε: 300, 4.20.

1,3-Dimethyl-10-(4-methylpentan-2-yl)-6,7,8,9-tetrahydrodiazepino[2,1-f]purine-2,4(1H,3H)-dione (30)

Reaction medium, DMF; time of heating under reflux, 10 h; excess of amine, 2. Crystallization with ethanol/H₂O gave pure compound **30**. Yield 61 %, mp 96–100 °C; TLC: *R_f*: 0.72; Anal. Calcd for C₁₇H₂₇O₂N₅ (**30**): C, 61.22; H, 8.17; N, 21.02. Found: C, 61.52; H, 8.09; N, 20.90. ¹H-NMR δ (ppm): 0.92–0.94 (m, 6H, CH(CH₃)₂), 1.19–1.28 (m, 4H CHCH₃+CH(CH₃)₂), 1.50–1.67 (m, 2H, CH₂CH), 1.73–1.97 (m, 4H, N₅CH₂(CH₂)₂CH₂N₁₀), 2.99–3.27(m, 2H, CH₂N₁₀), 3.36 (s, 3H, N₃CH₃), 3.50 (s, 3H, N₁CH₃), 4.02–4.10 (m, 1H, N₅CH), 4.27–4.34 (m, 2H, N₅CH₂), 4.48–4.55 (m, 1H, N₁₀CH). IR KBr (cm⁻¹): 2,946–2,876—CH₂, 1,695—CO(2), 1,664 (4), 747—CH₂. UV λ_{max}, logε: 300, 4.37.

1,3-Dimethyl-10-(pentan-2-yl)-6,7,8,9-tetrahydrodiazepino[2,1-f]purine-2,4(1H,3H)-dione (31)

Reaction medium, DMF; time of heating under reflux, 10 h; excess of amine, 2. Crystallization with ethanol/H₂O gave pure compound **31**. Yield 78 %, mp 113–115 °C; TLC: *R_f*: 0.70; Anal. Calcd for C₁₆H₂₅O₂N₅ (**31**): C, 60.17; H, 7.90; N, 21.93. Found: C, 60.19; H, 7.85; N, 21.78. ¹H-NMR δ (ppm): 0.90–0.94 (m, 3H, CH₂CH₃), 1.21 (d, *J*=6.88 Hz, 3H, CHCH₃), 1.32–1.64 (m, 4H, (CH₂)₂CH₃), 1.74–1.98 (m, 4H, N₅CH₂(CH₂)₂CH₂N₁₀), 2.99–3.07 (m, 1H, CH₂N₁₀), 3.20–3.28 (m, 1H, CH₂N₁₀), 3.50 (s, 3H, N₁CH₃), 4.01–4.09 (m, 1H, N₅CH), 4.17–4.24 (m, 2H, N₅CH₂), 4.49–4.57 (m, 1H, N₁₀CH). IR KBr (cm⁻¹): 2,949–2,871—CH₂, 1,700—CO(2), 1,661 (4), 745—CH₂. UV λ_{max}, logε: 300, 4.29.

10-Allyl-1,3-dimethyl-6,7,8,9-tetrahydrodiazepino[2,1-f]purine-2,4(1H,3H)-dione (32)

Reaction medium, solvent-free; time of heating under reflux, 5 h; excess of amine, 20. Crystallization with ethanol/H₂O gave pure compound **32**. Yield 65 %, mp 228–230 °C; TLC: *R_f*: 0.62; Anal. Calcd for C₁₇H₂₅O₂N₅ (**32**): C, 58.11; H, 6.63; N, 24.20. Found: C, 58.23; H, 6.48; N, 24.36. ¹H-NMR δ (ppm): 1.83–1.90 (m, 4H, N₅CH₂(CH₂)₂CH₂N₁₀), 3.15 (t, *J*=4.95 Hz, 2H, CH₂N₁₀), 3.37 (s, 3H, N₃CH₃), 3.51 (s, 3H, N₁CH₃), 4.07 (d, *J*=4.95 Hz, 2H, N₁₀CH₂), 4.33 (t, *J*=4.95 Hz, 2H, N₅CH₂), 5.21–5.30 (m, 2H, CH₂=CH), 5.89–6.00 (m, 1H, CH₂=CH).

IR KBr (cm⁻¹): 3,078–3,010—allyl, 2,936—CH₂, 1,696—CO(2), 1,652 (4), 750—CH₂. UV λ_{max}, logε: 298, 4.23.

8-Isopropyl-1,3-dimethyl-6,7-dihydro-1H-imidazo[2,1-f]purine-2,4(1H,3H)-dione (34)

A mixture of (0.73 g, 2 mmol) of 7-(2-bromoethyl)-8-bromothetheophylline (**1**) [16, 21] and isopropyl amine (1.42 g, 24 mmol) was heated under reflux for 5 h. After cooling, the precipitate was separated and crystallized from 30 % ethanol to give analytically pure compound **34**. Yield 50 %, mp 157–159 °C; Anal. Calcd for C₁₂H₁₇O₂N₅ (**34**): C, 54.74; H, 6.50; N, 26.60. Found: C, 54.35; H, 6.50; N, 26.28. ¹H-NMR δ (ppm): 1.83–1.90 (m, 4H, N₅CH₂(CH₂)₂CH₂N₁₀), 3.15 (t, *J*=4.95 Hz, 2H, CH₂N₁₀), 3.37 (s, 3H, N₃CH₃), 3.51 (s, 3H, N₁CH₃), 4.07 (d, *J*=4.95 Hz, 2H, N₁₀CH₂), 4.33 (t, *J*=4.95 Hz, 2H, N₅CH₂), 5.21–5.30 (m, 2H, CH₂=CH), 5.89–6.00 (m, 1H, CH₂=CH). IR KBr (cm⁻¹): 2,960–2,870—CH₂, 1,696—CO(2), 1,653—CO (4), 749—CH₂. UV λ_{max}, logε: 302, 4.21.

9-Butyl-1,3-dipropyl-6,7,8,9-tetrahydropyrimido[2,1-f]purine-2,4(1H,3H)-dione (35)

A mixture of (0.78 g, 2 mmol) of 1,3-dipropyl-7-(3-chloropropyl)-8-bromo-purine-2,4(3H,6H)-dione (**4**) [16, 17] and butyl amine (2.20 g, 30 mmol) was heated under reflux for 5 h. After cooling, the precipitate was separated and crystallized from ethanol to give analytically pure compound **35**. Yield 45 %, mp 77–79 °C; Anal. Calcd for C₁₈H₂₉O₂N₅×H₂O (**29**): C, 59.14; H, 8.42; N, 19.16. Found: C, 58.85; H, 8.81; N, 19.10. ¹H-NMR δ (ppm): 0.91–0.98 (m, 8H, CH₃CH₂CH₂+CH₃-CH₂CH₂CH₂), 1.28–1.41 (m, 6H, CH₃CH₂CH₂N₁+CH₃CH₂CH₂CH₂), 3.24 (t, *J*=5.64 Hz, 2H, N₉-CH₂), 3.52 (t, *J*=7.15Hz, 2H, N₉-CH₂), 3.89–3.98 (m, 4H, N₁,N₃CH₂CH₂CH₃), 4.20 (t, *J*=5.19Hz, 2H, N₅CH₂). IR KBr (cm⁻¹): 3,518–2,870 H₂O, 2,959–2,874—CH₂, 1,689—CO (2), 1,655—CO (4), 754—CH₂. UV λ_{max}, logε: 303, 4.30.

Pharmacology

Adenosine receptor binding studies

[³H]CCPA (58 Ci/mmol) was purchased from NEN Life Sciences; [³H]MSX-2 (84 Ci/mmol), [³H]PSB-11 (28 Ci/mmol) and [³H]PSB-603 (73 Ci/mmol) were obtained from Quotient Biosciences (custom synthesis). The non-radioactive precursors of [³H]MSX-2 (MSX-1) [33, 48], [³H]PSB-11 (PSB-10) [36, 49] and [³H]PSB-603 (PSB-603) [35] were synthesized in our laboratory. Frozen rat brains obtained from Pel Freez®, Rogers, AR, USA, were dissected to obtain cortical membrane preparations

for A_1 assays and striatal membrane preparations for A_{2A} assays as described [50]. CHO cells stably transfected with the human adenosine A_1 , A_{2B} and A_3 receptors were used for membrane preparations as previously described [50, 51]. For A_{2A} adenosine receptor assays, commercially available membrane preparations containing the human A_{2A} AR were obtained from Perkin Elmer Life Sciences (Boston, MA, USA). Stock solutions of the compounds were prepared in dimethylsulfoxide (DMSO); the final concentration of DMSO in the assays did not exceed 2.5 %. Initial screening was performed at a single concentration of 25 or 10 μM , depending on target receptor. Binding assays were performed as previously described [33, 50–54]. Curves were determined using six to seven different concentrations of test compounds spanning three orders of magnitude. At least three separate experiments were performed, each in triplicate. For non-linear regression analysis, the Cheng–Prusoff equation and K_D values of 0.2 nM (rat A_1) and 0.6 nM (human A_1), respectively, for [^3H]CCPA [34]; 8 nM (rat A_{2A}) and 7 nM (human A_{2A}), respectively, for [^3H]MSX-2 [33, 55]; 0.41 nM (human A_{2B}) for [^3H]PSB-603 (human A_{2B}) [35] and 4.9 nM (human A_3) for [^3H]PSB-11 [52] were used to calculate K_i values from IC_{50} values.

Anticonvulsant screening

The anticonvulsant evaluation was carried out using reported procedures [39, 40, 47]. Male albino mice (Carworth Farms Nr 1, 18–25 g) and male albino rats (Sprague-Dawley, 100–150 g) were used as experimental animals. A group of three to five mice was used in MES and ScMet tests, and a group of four to eight animals was used in rotorod tests. For the evaluation of activity after oral administration, a group of four rats was used. The test compounds were suspended in 0.5 % methylcellulose water mixture.

In the preliminary screening, each compound was administered as an i.p. injection at three dose levels (30, 100 and 300 mg/kg), with anticonvulsant activity and neurotoxicity assessed at 30-min and 4-h intervals after administration. For some compounds, also intervals of 15 min and 1 h were used. Anticonvulsant efficacy was measured by MES, ScMet and neurological deficit in the rotorod tests, and the data are presented in Table 4. Some selected derivatives were examined for oral activity in the MES and ScMet and neurotoxicity screen (rotorod test) 30 and 50 mg/kg doses. The results are summarized in Table 5.

The pharmacological parameters estimated in the preliminary screening were quantified for compounds **9** and **13** in mice (i.p.) (Table 7) and for comp. **9** also in rats (p.o.) (Table 6).

Anticonvulsant activity was expressed in terms of the median effective dose (ED_{50}) MES and ScMet in mice and ED_{50} ScMet in rats, and neurotoxicity was expressed as the

median toxic dose (TD_{50}). For determination of ED_{50} and TD_{50} , groups of eight mice and rats were given a range of i.p. or p.o. doses of the test drug until at least three points were established in the range of 10–90 % seizure protection or minimal observed neurotoxicity. From the plot of these data, the respective ED_{50} , TD_{50} value (95 % confidence intervals, slope of the regression line and the standard error of the slope were calculated by a computer program based on [38] methods described by Finney. For comp. **9**, the hippocampal kindling model of focal seizures was applied (Table 6). In this procedure, the bipolar electrodes were placed surgically in the ventral hippocampus of adult male Sprague-Dawley rats. Stage five behavioural seizures are produced by using a stimulus consisting of a 50-Hz, 10-s train of 1-ms biphasic 200- μA pulses delivered every 30 min for 6 h. A single dose of comp. **9** is then administered intraperitoneally, 15 min following the last stimulation. The anticonvulsant activity of **9** is assessed every 30 min for 2 h, starting 15 min after administering the test material. After each stimulation, seizure scores and after discharge durations were recorded (Table 6).

Molecular modelling

The 3D molecule models of **5–35** and caffeine were built using Schrödinger Maestro molecular modelling environment [56] basing on presented crystal structures of chosen pyrimido- and diazepinopurinediones (**10** and **30**, respectively). The geometry optimization was performed using the multiple minimization method as implemented in MacroModel with MMFF force field and Truncated Newton Conjugate Gradient options and terminated when the root means square (RMS) of conjugate gradient was below $0.05 \text{ kJ mol}^{-1} \text{ \AA}^{-1}$. The minimization was carried out for water solvent, with distance-dependent dielectric constant as a way to treat electrostatic interactions.

The template 3REY was prepared to docking using the Protein Preparation Wizard option as a part of Schrodinger package. The ligand binding cavity was confined to a box with the crystal ligand in the center (size of the box set automatically), according to the receptor grid generation procedure. The studies of docking to the rigid receptor binding pocket were performed using Glide program, with standard precision (SP) mode.

For validation, redocking of XAC ligand was done. After initial docking, 10 poses for each ligand, with RMS deviation higher than 0.5 \AA , were kept, and post-minimized, RMSD values between predicted and crystallographic positions of the ligand were calculated. Redocking of XAC was performed twice: (1) without any constraints and (2) with the amino hydrogen atom of the Asn253 (6.55) side chain used as a constrained H-bond donor for a carbonyl group of xanthine.

Molecular docking of A_{2A} AR antagonists

Docking simulations of low-energy conformations for all of the synthesized compounds and caffeine were performed with SP mode to both templates with limitation used during redocking: the constrained H-bond between the side chain amino group of Asn253 (6.55) and the ligand. Five poses obtained after docking for each ligand (RMS deviation higher than 0.5 Å) were post-minimized, and final poses were kept and analysed according to the obtained docking score values.

For the graphic presentation of the selected structures with the highest docking scores, representing individual clusters of poses, PyMOL [57] software was used.

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