

City University of New York (CUNY)

CUNY Academic Works

Publications and Research

Lehman College

2007

mda-7/IL-24, novel anticancer cytokine: Focus on bystander antitumor, radiosensitization and antiangiogenic properties and overview of the phase I clinical experience (Review)

IRINA V. LEBEDEVA

LUNI EMDAD

ZAO-ZHONG SU

PANKAJ GUPTA

MOIRA SAUANE
CUNY Lehman College

See next page for additional authors

[How does access to this work benefit you? Let us know!](#)

More information about this work at: https://academicworks.cuny.edu/le_pubs/63

Discover additional works at: <https://academicworks.cuny.edu>

This work is made publicly available by the City University of New York (CUNY).
Contact: AcademicWorks@cuny.edu

Authors

IRINA V. LEBEDEVA, LUNI EMDAD, ZAO-ZHONG SU, PANKAJ GUPTA, MOIRA SAUANE, DEVANAND SARKAR, MICHELLE R. STAUDT, SHI-JIAN LIU, MOHIUDDIN M. TAHER, RUOYU XIAO, PAOLA BARRAL, SEOK-GEUN LEE, DONGNING WANG, NICOLLAQ VOZHILLA, EUN SOOK PARK, LEJUAN CHATMAN, HABIB BOUKERCHE, RAJAGOPAL RAMESH, SATOSHI INOUE, SUNIL CHADA, RONG LI, ANTHONY L. DE PASS, PARAMESHWAR J. MAHASRESHTI, IGOR P. DMITRIEV, DAVID T. CURIEL, ADLY YACOUB, STEVEN GRANT, PAUL DENT, NEIL SENZER, JOHN J. NEMUNAITIS, and PAUL B. FISHER

***mda-7/IL-24*, novel anticancer cytokine: Focus on bystander antitumor, radiosensitization and antiangiogenic properties and overview of the phase I clinical experience (Review)**

IRINA V. LEBEDEVA¹, LUNI EMDAD^{1,2}, ZAO-ZHONG SU¹, PANKAJ GUPTA¹, MOIRA SAUANE¹, DEVANAND SARKAR^{1,2}, MICHELLE R. STAUDT¹, SHI-JIAN LIU¹, MOHIUDDIN M. TAHER¹, RUOYU XIAO¹, PAOLA BARRAL¹, SEOK-GEUN LEE¹, DONGNING WANG^{1,3}, NICOLLAQ VOZHILLA¹, EUN SOOK PARK¹, LEJUAN CHATMAN¹, HABIB BOUKERCHE^{2,11}, RAJAGOPAL RAMESH⁴, SATOSHI INOUE⁴, SUNIL CHADA⁵, RONG LI⁶, ANTHONY L. DE PASS⁶, PARAMESHWAR J. MAHASRESHTI⁷, IGOR P. DMITRIEV⁷, DAVID T. CUIEL⁷, ADLY YACOUB⁸, STEVEN GRANT⁹, PAUL DENT⁸, NEIL SENZER¹⁰, JOHN J. NEMUNAITIS¹⁰ and PAUL B. FISHER¹⁻³

Departments of ¹Urology, ²Neurosurgery and ³Pathology, Herbert Irving Comprehensive Cancer Center, Columbia University Medical Center, College of Physicians and Surgeons, New York, NY 10032; ⁴Department of Thoracic and Cardiovascular Surgery, M.D. Anderson Cancer Center, Houston, TX 77030; ⁵Introgen Therapeutics, Inc., Houston, TX 77030; ⁶Department of Biology, Long Island University-Brooklyn, Brooklyn, NY 11201; ⁷Division of Human Gene Therapy, Departments of Medicine, Pathology and Surgery, University of Alabama, Birmingham, AL 35294; Departments of ⁸Biochemistry and ⁹Medicine, Virginia Commonwealth University, Richmond, VA 23284; ¹⁰Mary Crowley Research Center, Dallas, TX 75246, USA

Received January 2, 2007; Accepted March 27, 2007

Abstract. Subtraction hybridization applied to a 'differentiation therapy' model of cancer employing human melanoma cells resulted in the cloning of melanoma differentiation associated gene-7/interleukin-24 (*mda-7/IL-24*). Initial studies confirm an inverse correlation between *mda-7* expression and melanoma development and progression. Forced expression of *mda-7* by means of a plasmid or via a replication incompetent adenovirus (Ad.*mda-7*) promotes growth suppression and induces apoptosis in a broad array of human cancers. In contrast, *mda-7* does not induce growth suppressive or toxic effects in normal cells. Based on structure (containing an IL-10 signature motif), secretion by cells (including subsets of T-cells) and location on chromosome 1q (in an area containing IL-10-family genes), *mda-7* has now been renamed *mda-7/IL-24*. Studies by several laboratories have uncovered many of *mda-7/IL-24*'s unique properties, including cancer-specific apoptosis-

induction, cell cycle regulation, an ability to inhibit angiogenesis, potent 'bystander antitumor activity' and a capacity to enhance the sensitivity of tumor cells to radiation, chemo-

Abbreviations: Ad, adenovirus; APC, adenomatous polyposis coli gene; bFGF, basic fibroblast growth factor; CLL, chronic lymphocytic leukemia; DOTAP:Chol, 1,2-dioleoyl-3-(trimethylammonio) propane; Cholesterol; COX-2, cyclooxygenase-2; CR, complete response; ECD, endothelial cell differentiation; ER, endoplasmic reticulum; FADD, Fas-associated death domain; FAK, focal adhesion kinase; FGF, fibroblast growth factor; FISP, IL-4-induced secreted protein; FITC, fluorescein isothiocyanate; *GADD* genes, growth arrest and DNA damage inducible genes; GSK-3 β , glycogen synthase kinase-3 β ; GM-CSF, granulocyte macrophage colony-stimulating factor; HUVEC, human umbilical vein endothelial cells; IKK, I κ B α kinase; IR, ionizing radiation; JAK-STAT, janus kinase signal transducer and activator of transcription; *mda-7*, melanoma differentiation associated gene-7; IFN, interferon; IL, interleukin; LPS, lipopolysaccharide; MEF, mouse embryonic fibroblasts; MMP, matrix metalloproteinase; NSCLC, non-small cell lung cancer; PBMC, peripheral blood mononuclear leukocytes; *PEG-3*, progression elevated gene-3; PHA, phytohemagglutinin; PI3K, phosphatidylinositol 3-kinase; PKR, double-stranded RNA-dependent protein kinase; PR, partial response; ROS, reactive oxygen species; SAE, serious adverse effects; SCCHN, squamous cell carcinoma of the head and neck; SD, stable disease; SMS, smooth muscle cell; TCF/LEF, T-cell factor/lymphoid enhancer factor; TGF- β , transforming growth factor- β ; TNF- α , tumor necrosis factor- α ; TUNEL, terminal deoxynucleotidyltransferase dUTP nick-end labeling; UPR, unfolded protein response; VEGF, vascular endothelial growth factor; vp, viral particles

Correspondence to: Professor Paul B. Fisher, Departments of Pathology, Urology and Neurosurgery, Columbia University Medical Center, College of Physicians and Surgeons, BB1501, 630 West 168th Street, New York, NY 10032, USA
E-mail: pbf1@columbia.edu

Present address: ¹¹National de la Sante et de la Recherche Medicale (INSERM)1, Lyon Cedex, France

Key words: *mda-7/IL-24*, apoptosis, bystander effect, radiosensitization, antiangiogenesis, phase I clinical trial

therapy and monoclonal antibody therapy. Moreover, based on its profound cancer tropism, substantiated by *in vivo* human xenograft studies in nude mice, *mda-7/IL-24* (administered as Ad.*mda-7*) was evaluated in a phase I clinical trial in patients with melanomas and solid cancers. These studies document that *mda-7/IL-24* is well tolerated and demonstrates evidence of significant clinical activity. In these contexts, *mda-7/IL-24* represents a unique cytokine gene with potential for therapy of human cancers. The present review focuses on three unique properties of *mda-7/IL-24*, namely its potent 'bystander anti-tumor activity', ability to sensitize tumor cells to radiation, and its antiangiogenesis properties. Additionally, an overview of the phase I clinical trial is provided. These studies affirm that *mda-7/IL-24* has promise for the management of diverse cancers.

Contents

1. Introduction
2. Mechanisms of antiapoptotic properties of *mda-7/IL-24*
3. *mda-7/IL-24* expression radiosensitizes cancer cells
4. Ectopic expression of *mda-7/IL-24* inhibits invasion and migration of cancer cells
5. Antiangiogenic activity of *mda-7/IL-24*
6. Immune properties of *mda-7/IL-24*
7. 'Bystander antitumor' activity of *mda-7/IL-24*
8. Translation of *mda-7/IL-24* into the clinic - A good beginning
9. Future applications for improving the therapeutic index of *mda-7/IL-24*

1. Introduction

A novel melanoma differentiation associated (*mda*) gene, *mda-7*, was identified with a subtraction hybridization protocol (1) using temporally-spaced cDNA libraries isolated from terminally differentiated human melanoma cells treated with β -interferon and the protein kinase C activator mezerein (2). The *mda-7* cDNA encodes a 24-kDa protein, which contains an interleukin (IL)-10 signature motif at amino acids 101-121 (SDAESCYLVHTLLEFYLKTVF) shared by other members of the IL-10 family of cytokines (3,4). Based on sequence homology, chromosomal localization, and its functional properties, the *mda-7* gene is now classified as a member of the IL-10 family of cytokines and named IL-24 (4-8). The MDA-7/IL-24 protein contains a consensus signal sequence and a proteolytic cleavage site permitting secretion from cells. Expression of MDA-7/IL-24 protein in normal cells is restricted to cells of the immune system and melanocytes (5,7,9). MDA-7/IL-24 expression is decreased during pathologic progression of melanocytes to melanomas as demonstrated by both RT-PCR (2) and immunohistochemistry (10). This correlation between the loss of gene expression and tumor invasion suggests that *mda-7/IL-24* might function as a tumor suppressor (2,11,12). Further studies *in vitro* indicated that the functional antitumor attributes of *mda-7/IL-24* were dependent on MDA-7/IL-24 expression level (13). When present at low physiological levels, *mda-7/IL-24* exhibited specific immunostimulatory properties (see details below).

However, when this gene was overexpressed in diverse cancer cells *in vitro* by transfection with a *mda-7/IL-24* cDNA or by adenoviral infection with a recombinant replication incompetent adenovirus, Ad.*mda-7*, it displayed potent tumor killing and apoptosis-inducing properties (reviewed in refs. 4,6,8,14-20).

Infection with Ad.*mda-7* results in apoptosis induction and cell death in a wide variety of solid tumors including melanoma, malignant glioma, fibrosarcoma and carcinomas of the breast, cervix, colorectum, liver, lung, ovary and prostate. The tumor suppressing ability of *mda-7/IL-24* is now well established and is independent of the status of other tumor suppressor genes, such as *p53* and *Rb*, or apoptosis regulating genes, such as *bax* or *caspases*, in tumor cells (12,21-25). A provocative discovery was that overexpression of *mda-7/IL-24* in normal cells, such as normal human epithelial cells, melanocytes, astrocytes or fibroblasts, did not affect their growth or viability. This *in vitro* tumor-selective cytotoxicity was confirmed by *in vivo* experiments in animal models containing human breast, lung and colorectal carcinomas and in glioma xenografts (23-26). Based on the results obtained using *in vitro* studies and in *in vivo* preclinical animal modeling, *mda-7/IL-24* emerged as a promising new and potentially widely applicable antitumor therapeutic agent, possessing direct tumor growth inhibitory effects as well as 'bystander antitumor' properties. Findings from a completed phase I dose-escalation trial on 22 advanced cancer patients who received intratumoral injection of a non-replicating adenovirus vector carrying the *mda-7/IL-24* transgene (INGN 241) were recently reviewed (6,17,18,27-29). These results confirm that the clinical activities of INGN 241 (Ad.*mda-7*) include apoptotic and antitumor activities and systemic immune activation and can generate both direct tumor growth inhibition and 'bystander antitumor' effects.

2. Mechanisms of antiapoptotic properties of *mda-7/IL-24*

IL-20/IL-22 receptor independence. Within the IL-10 family, IL-19, IL-20 and IL-24 exhibit substantial sharing of receptor complexes; all three are capable of signaling through IL-20R1/IL-20R2, and both IL-20 and IL-24 can also use IL-22R1/IL-20R2 (30). Nevertheless, the biological activities of these three cytokines appear quite distinct; since only *mda-7/IL-24* induces tumor-specific apoptosis, and this effect can be receptor-independent (31). The MDA-7/IL-24 protein binds to IL-20 and IL-22 receptor complexes resulting in JAK/STAT activation (30,32,33). However, treatment with tyrosine kinase-specific inhibitors (Genistein and AG18) or a JAK-selective inhibitor (AG490) does not alter Ad.*mda-7*-induced apoptosis in diverse cancer cell lines (31). In addition, there is no correlation between the pattern of expression of IL-20R1, IL-20R2, and IL-22R mRNA and susceptibility to Ad.*mda-7*-induced cell death in different cell lines. Therefore, signaling events leading to Ad.*mda-7*-induced apoptosis might be tyrosine kinase independent and can be distinguished from *mda-7/IL-24* cytokine function-related properties mediated by the IL-20/IL-22 receptor complexes that require JAK/STAT kinase activity (31,34,35).

Mitochondrial pathway and reactive oxygen species (ROS) production. *In vitro* studies were performed to address the mechanism(s) by which overexpression of *mda-7/IL-24*

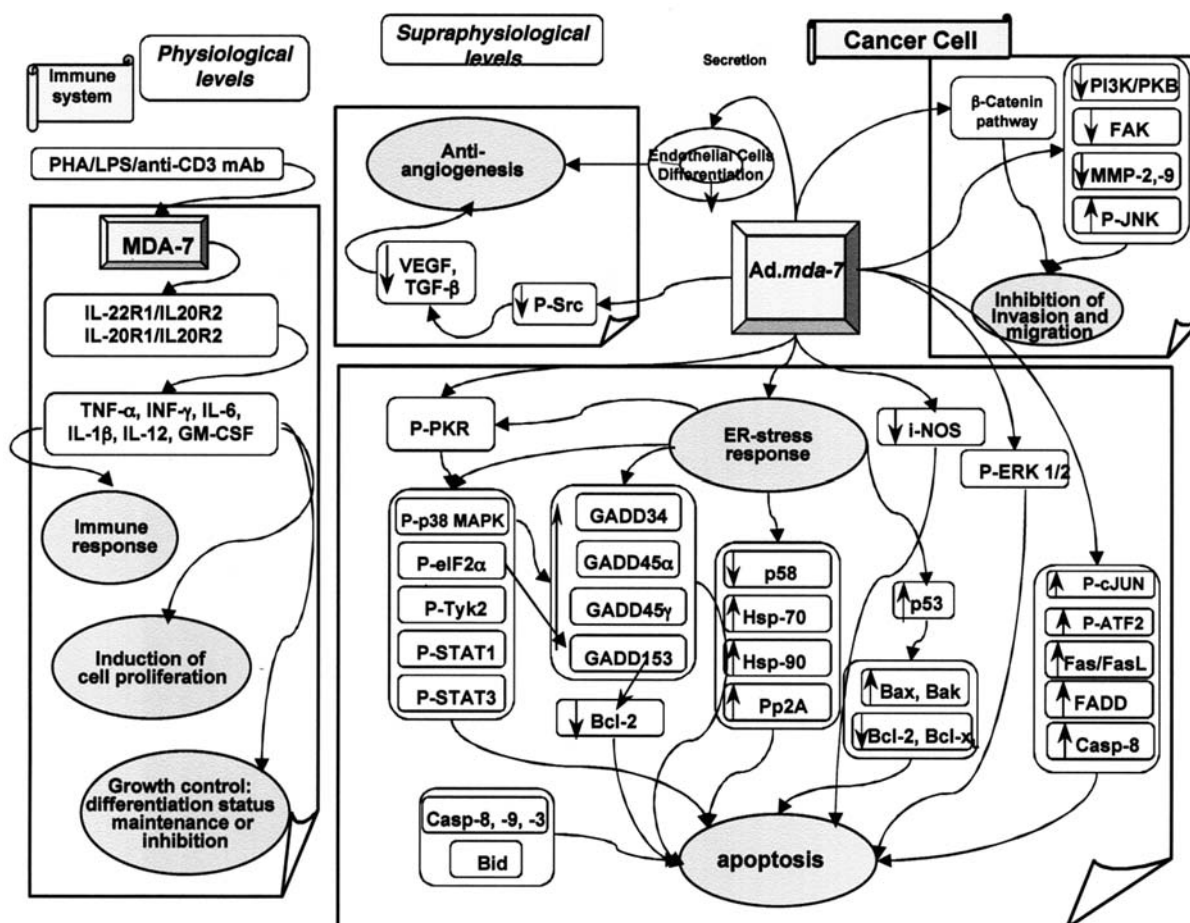


Figure 1. Model of *mda-7/IL-24*-induced apoptosis in cancer cells. Outline of proposed physiologic (left) and ectopic (right) effects of *mda-7/IL-24* on normal and cancer cells. When normally or ectopically overexpressed, MDA-7/IL-24 localizes to the ER/Golgi compartments, whether or not the protein contains a secretory signal. Accumulation of MDA-7/IL-24 protein in this compartment triggers apoptosis that could involve induction of ER stress and/or reactive oxygen species in mitochondria. Secreted MDA-7/IL-24 protein employs the IL-20/IL-22 receptors to activate signal transduction pathways and/or potentially enter cancer cells and activate proapoptotic pathways by localization and accumulation in the ER/Golgi compartment and/or by inducing mitochondrial dysfunction. A combination of pathways triggered by *mda-7/IL-24* results in transformed cell-specific apoptosis. Modified from Lebedeva *et al* (17).

induces death selectively in cancer cells. Despite these efforts, the precise pathways of *mda-7/IL-24*-induced apoptosis remain to be defined. Evidences from earlier studies suggested an involvement of the intrinsic/mitochondrial pathway of apoptosis. Transfection of breast carcinoma cells with an *mda-7/IL-24* cDNA led to proapoptotic Bax up-regulation and subsequent apoptosis induction (25,36). The onset of apoptosis was blocked by antiapoptotic Bcl-2 overexpression. Further studies in melanoma (12), glioblastoma multiforme (26,37,38), renal carcinoma (39) and prostate carcinomas (21,40,41) confirmed the involvement of the Bcl-family of proteins in *mda-7/IL-24*-induced apoptosis. In most tumor cell contexts, overexpression of MDA-7/IL-24 protein leads to the significant down-regulation of antiapoptotic (Bcl-2 and Bcl-x_L) and/or to the up-regulation of proapoptotic (Bax and Bak) members of the Bcl-family. It should be noted that Ad.*mda-7* can induce apoptosis in *bax*-null DU-145 prostate cancer cells (21); therefore, apoptosis can be mediated by a *bax*-independent pathway. Moreover, overexpression of antiapoptotic Bcl-family members differentially protects prostate cancer cells from the *mda-7*-induced apoptosis (21). Bcl-x_L overexpression prevents Ad.*mda-7*-induced apoptosis in DU-145 and PC-3 prostate carcinoma cells, while Bcl-2 overexpression protects LNCaP cells from *mda-7/IL-24*-induced

apoptosis. The reasons for this disparity are not understood and are currently being investigated.

Detailed studies in prostate cancer cells identify reactive oxygen species (ROS) as an active component in *mda-7/IL-24*-induced apoptosis (Fig. 1) (39,42,43). When overexpressed inside a cancer cell MDA-7/IL-24 protein directly or indirectly affects mitochondria, causing mitochondrial dysfunction and the inner mitochondria membrane potential to drop thereby leading to ROS production and to apoptosis induction. All of the changes described above are absent after Ad.*mda-7* infection of normal prostate epithelial cells. Antioxidants (*N*-acetyl-L-cysteine and Tiron) and inhibitors of mitochondrial permeability transition (cyclosporine A and bongkreikic acid) inhibit Ad.*mda-7*-induced mitochondrial dysfunction and apoptosis in prostate cancer cells. Conversely, agents augmenting ROS production (arsenic trioxide, NSC656240, and PK11195) facilitate Ad.*mda-7*-induced apoptosis. Ectopic expression of Bcl-2 and Bcl-x_L inhibits mitochondrial changes, ROS production, and apoptosis providing additional support for an association between mitochondrial dysfunction and Ad.*mda-7* action. These studies present definitive evidence that changes in mitochondrial function and ROS production represent key components associated with selective killing of prostate cancer cells by *mda-7/IL-24*.

The ability of MDA-7/IL-24 to produce ROS may prove extremely important for treatment of pancreatic cancer. Pancreatic cancer cells are resistant to *mda-7/IL-24*-induced apoptosis due to a diminished capacity to convert *mda-7/IL-24* mRNA into protein (44). This translational block can be reversed by combinational treatment with Ad.*mda-7/IL-24* and agents that increase the ROS levels within cells, such as arsenic trioxide, N-(4-hydroxyphenyl) retinamide, or dithiophene (42). Induction of apoptosis *in vitro* and suppression of tumorigenesis *in vivo* in nude mice are induced in pancreatic cancers independently of their *K-ras* status upon combinatorial treatment with Ad.*mda-7/IL-24* and a ROS-inducing agent. ROS inhibitors, including N-acetyl-cysteine and Tyron, block this effect.

ER-stress and its link to mitochondria. In addition to modification of mitochondrial function, *mda-7/IL-24* has also been shown to kill cancer cells by causing endoplasmic reticulum (ER) stress (45-47). ER stress may be caused by misfolded protein accumulation followed by the activation of a highly conserved unfolded protein response (UPR). These events lead to apoptosis through the induction of growth arrest and DNA-damage-inducible (*GADD*) genes (48,49). The hypothesis of a role for *mda-7/IL-24*-induced ER stress and cancer cell apoptosis is supported by the fact that *mda-7/IL-24* induces *GADD* genes and further activates p38MAPK in the context of transformed cells (46,50).

Previous studies document that signaling events leading to Ad.*mda-7*-induced transformed cell apoptosis are tyrosine kinase-independent (31). These results suggest that *mda-7/IL-24* cancer cell-specific activity can occur through mechanisms independent of binding to its currently recognized cognate receptors and might even occur independent of receptor function. An adenovirus vector expressing a non-secreted version of MDA-7/IL-24 protein was generated by deletion of its signal peptide (45). This non-secreted protein was as effective as wild-type secreted MDA-7/IL-24 in inducing apoptosis in prostate carcinoma cell lines and displayed transformed cell specificity and localization of MDA-7/IL-24 in the Golgi/ER compartments. These results indicate that *mda-7/IL-24*-mediated apoptosis can be triggered through a combination of intracellular as well as secretory mechanisms and could occur efficiently in the absence of protein secretion.

Treatment of susceptible prostate cancer cell lines with Ad.*mda-7* as well as Ad.SP*mda-7* induces killing to a comparable extent through ERK1/2-dependent and JAK/STAT-independent pathways (45). The fact that both secreted and non-secreted forms of MDA-7/IL-24 protein have comparable apoptosis-inducing activity was unanticipated, adding an additional level of complexity in understanding how this novel molecule works. Localization of full-length MDA-7/IL-24 protein in the ER/Golgi compartments is consistent with the signal peptide hypothesis (51) and the currently known and predicted secreted cytokine nature of the protein (6,14). Because the signal-peptideless mutant of MDA-7/IL-24 protein does not contain an export signal, it is predicted to remain in the cytosol. We have, however, confirmed through confocal immunofluorescence studies that a significant fraction of this protein is able to enter the ER and Golgi apparatus and that proteins derived from wild-type and mutant viruses

appear to have overlapping patterns of localization within the cell (45). It is not possible to rule out cryptic internalization signals that become active in the absence of the actual signal peptide, as the identity of these cryptic sites is currently hypothetical. Western blot analyses performed on protein-derived cytosolic and extracellular fractions of cells infected with both viruses indicated that only full-length MDA-7/IL-24 was processed and secreted. It is also possible that adenovirus infection produced relatively large amounts of protein that even in the absence of a specific targeting sequence possesses the ability to cross membranes and accumulate in the ER/Golgi because of charge and/or tertiary structure. However, because localization of MDA-7/IL-24 is similar in both normal (P69) and cancer (DU-145) cells, differences in cellular localization of this protein can be excluded as a direct mechanism underlying the differential apoptosis-inducing activity of MDA-7/IL-24 toward cancer cells. There are two possible explanations of differential MDA-7/IL-24-induced killing in tumor and normal cells. One is that enhanced sensitivity may be due to the 'activated' or 'destabilized' nature of tumor compared to normal cells, which enhances cell death after the ER-stress response is triggered. The other possibility is that *mda-7/IL-24* not only induces the classical ER-stress response that favors apoptosis, but it also induces additional specific pathways that cause apoptosis only in transformed cell lines. We demonstrated that *mda-7/IL-24* localizes to the ER compartment both in normal and cancer cells and is therefore in a position to induce this pathway irrespective of the transformation status of the cell (45). Current data provide some support for both hypotheses, however additional studies are required. *GADD* family gene induction as well as p38 MAPK activation is induced only in transformed and not in normal cells (46,50) indicating that the disparate response might be due to differential activation, either in strength or in duration of the ER-stress response. However, this does not rule out the activation of additional pathways, specifically in cancer cells. In a microarray-based study, *mda-7/IL-24* was shown to induce expression of ER-stress response genes such as *BiP/GRP78*, *PP2A*, *HSJ1* and *TRAI* in H1299 lung carcinoma cells (52). Nevertheless, with the exception of *BiP/GRP78*, which is selectively up-regulated in cancer cells (46), no comparative data for most of these genes is available contrasting normal and cancer cells.

MDA-7/IL-24 modulates multiple apoptotic signaling pathways. Expanding studies in diverse systems reveal that the signal transduction pathways mediating *mda-7/IL-24*-induced apoptosis are varied and multiple signaling pathways are activated in different tumor cell lines upon Ad.*mda-7* infection or treatment with recombinant GST-MDA-7 protein (53) (summarized in Fig. 2). Experiments in melanoma cell lines demonstrated that SB203580, a specific inhibitor of p38 MAPK pathway, protects these cells from Ad.*mda-7*-induced apoptosis. Ad.*mda-7* infection resulted in phosphorylation of p38 MAPK and induction of the *GADD* family of genes in melanoma cells, but not in normal melanocytes (50). In general, overexpression of each *GADD* gene from the family in combination (*GADD34*, *GADD45 α* , *GADD45 β* , *GADD45 γ* and *GADD153*) led to synergistic or cooperative antiproliferative effects (54). Ad.*mda-7* infection resulted in significant

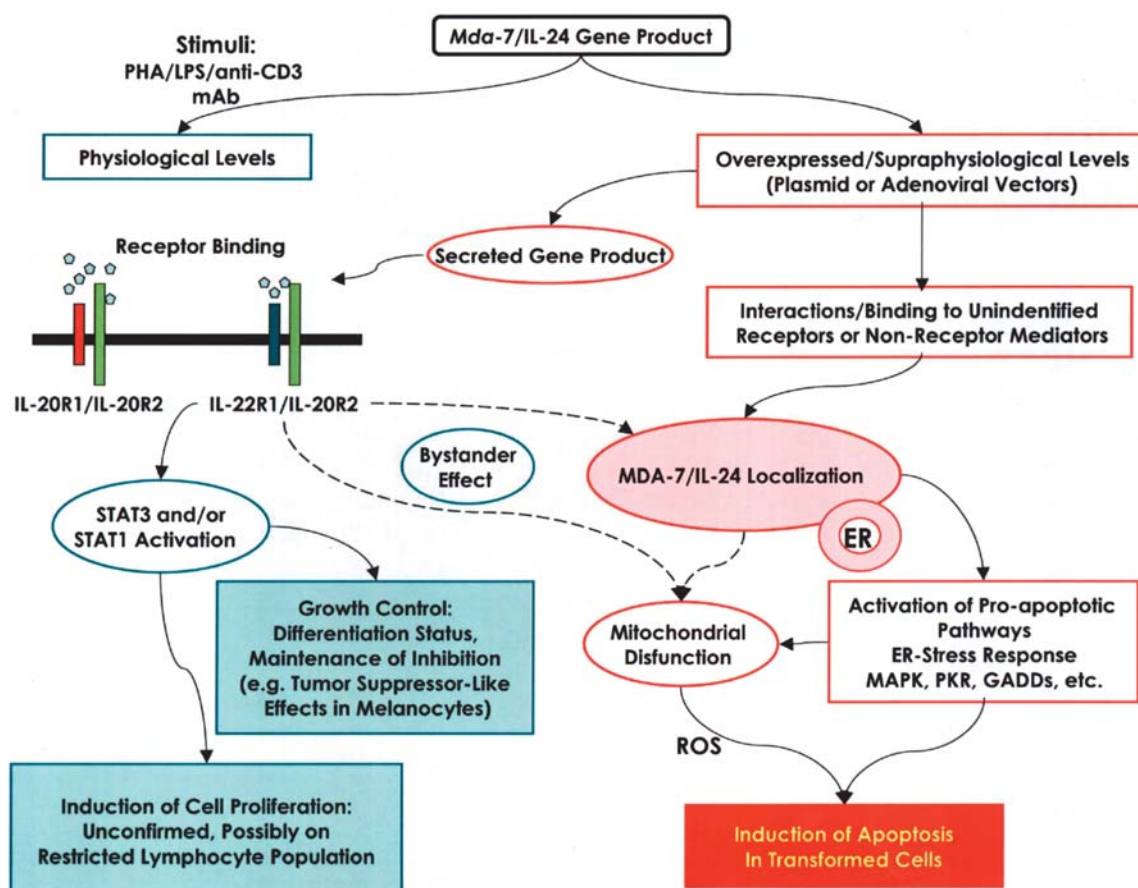


Figure 2. Signaling pathways involved in *mda-7/IL-24* activity in cancer cells, endothelial cells and immune system cells. Overview of the signaling pathways associated with Ad.*mda-7* and MDA-7/IL-24 activity in cancer cells and in the immune system. Modified from Sauane *et al* (45).

induction of *GADD153*, *GADD45a* and *GADD34*, and a moderate induction of *GADD45γ* (50). Both inhibition of the p38 MAPK pathway (either pharmacologically with SB203580 or by an adenovirus expressing a dominant negative p38 MAPK) and inhibition of the *GADD* family of genes by an antisense approach rescued melanoma cells from Ad.*mda-7*-induced apoptosis. Activation of the p38 MAPK pathway followed by induction of the *GADD* family of genes also plays a crucial role in Ad.*mda-7*-modulated apoptosis in glioblastoma multiforme (37), prostate cancer (31,41,46), and breast cancer cells (unpublished data).

Paradoxically, p38 MAPK phosphorylation by MDA-7/IL-24 in chronic lymphocytic leukemia (CLL)-B-cells promoted survival of these malignant cells (55). Both *mda-7/IL-24* mRNA and protein were overexpressed in the CLL B-cells examined, and p38 MAPK, a downstream *mda-7/IL-24* signaling target, was highly phosphorylated in all CLL cells, but not in normal B-cells. Obviously, phospho-p38 MAPK has no proapoptotic functions in CLL cells and may instead be required for survival, as suggested from these studies. Correspondingly, phosphorylation of p38 MAPK following transfection of CLL cells with *mda-7/IL-24* promoted CLL cell survival (55). These studies do not exclude the possibility of *mda-7/IL-24* signaling through a p38 MAPK-independent mechanism; however, their data suggest a competition between *mda-7/IL-24* and SB203580 (a specific inhibitor of p38 MAPK) for p38 MAPK activation.

In non-small cell lung carcinoma (NSCLC) cells, Ad.*mda-7* induced apoptosis via up-regulation of double stranded RNA-dependent protein kinase (PKR). Infection with Ad.*mda-7* led to the phosphorylation of PKR and also its downstream targets eIF2 α , Tyk2, Stat1, Stat3, and p38 MAPK (56,57). Ad.*mda-7* treatment activated caspases 3, 8 and 9 and cleavage of Bid and PARP in NSCLC cells. The activation of PKR appeared to be upstream of caspase activation because pre-treatment with caspase inhibitors failed to prevent PKR phosphorylation. Treatment with a specific serine/threonine kinase inhibitor 2-aminopurine blocked Ad.*mda-7*-induced apoptosis and also activation of PKR and eIF2 α (56). A recent study by Chada *et al* (58) reported PKR up-regulation in breast cancer cell lines upon Ad.*mda-7* infection. Further investigation of the interaction between MDA-7/IL-24 and PKR in NSCLC cells suggested a role for posttranslational regulation of PKR by MDA-7/IL-24 (59). A physical interaction of MDA-7/IL-24 and PKR was confirmed by immunofluorescence and coimmunoprecipitation studies. In both studies (56,59), the authors indicate an inability of Ad.*mda-7* to induce apoptosis in PKR null (-/-) mouse embryonic fibroblasts (MEF), but not in PKR wild-type MEFs, providing additional proof of PKR involvement in Ad.*mda-7*-modulated apoptosis. Nevertheless, this finding requires further independent confirmation given that Ad.*mda-7* does not induce apoptosis in a wide array of normal human or rodent cells.

Activation of p38 MAPK following Ad.*mda-7* infection appears to be a common event linking the PKR and p38 MAPK pathways (50,56,57). It is possible that in melanoma cells p38 MAPK activation is downstream of PKR activation, although in melanoma cells the post-p38 signal transduction changes seem to be more important in Ad.*mda-7*-induced apoptosis. Since eIF2 α phosphorylation activates the transcription factor ATF4, which in turn activates *GADD153* (60), there is a significant level of cross talk between the PKR and the p38 MAPK signal transduction pathways. Additional studies are required to identify the upstream molecules within PKR and p38 MAPK pathways involved in Ad.*mda-7*-induced apoptosis.

The c-Jun NH₂-terminal kinase (JNK) pathway is another important signaling pathway involved in *mda-7/IL-24*-induced apoptosis. Infection with Ad.*mda-7* radiosensitizes malignant glioma (37,61) and prostate cancer cells (40). A combination of Ad.*mda-7* and γ -irradiation activates JNK in glioma and prostate cancer cells, and treatment with a specific JNK inhibitor, SP600125, prevents apoptosis after the combination treatment (40,61). JNK activation leads to down-regulation of antiapoptotic Bcl-x_L and/or Bcl-2 in glioma and prostate carcinoma cells (38,40) and to the up-regulation of proapoptotic Bax and Bak proteins in prostate carcinoma cells (40). Blocking of JNK phosphorylation with SP600125 abrogates these changes in the levels of Bcl-family proteins. In NSCLC cells, curcumin (dietary pigment that inhibits JNK activation) prevents phosphorylation of c-*jun* and radiosensitization by Ad.*mda-7* (62).

A potential negative regulation of β -catenin and PI3K/Akt signaling pathways, which are implicated in cell-cell adhesion, cytoskeletal rearrangements, and membrane trafficking in breast and lung cancer cells, by MDA-7/IL-24 was reported (63). Using microarray analysis, increased protein expression was evident from tumor suppressor genes such as *E-cadherin*, *APC*, *GSK-3 β* , and *PTEN* after Ad.*mda-7* infection. At the same time, expression of proto-oncogenes involved in β -catenin and PI3K signaling was decreased. Ad.*mda-7* treatment led to a redistribution of cellular β -catenin from the nucleus to the plasma membrane. As a result, LEF/TCF transactivation was significantly reduced, and E-cadherin- β -catenin adhesion complex was up-regulated in a tumor cell-specific manner. Furthermore, Ad.*mda-7* infection of breast and lung cancer cells down-regulated the expression of PI3K pathway members (p85 PI3K, FAK, ILK-1, Akt, and PLC- γ). Experiments did not indicate if overexpression of β -catenin or activation of the PI3K pathway protects these cells from Ad.*mda-7*-induced apoptosis. In conclusion, Ad.*mda-7* appears to negatively regulate both the PI3K and the β -catenin signaling pathways in breast and lung cancer cells resulting in restoration of apoptosis induction. Moreover, PI3K activation has been implicated in chemoresistance and radioresistance in tumor cells; thus, Ad.*mda-7* treatment should serve as a chemo- and radiosensitizer. Ad.*mda-7* regulates multiple members of the β -catenin and PI3K pathways that are considerably redundant and simultaneously produce antiproliferative, proapoptotic, and antimetastatic phenotypes. Although it is not clear where these changes are initiated, *mda-7/IL-24* appears to act upstream of PI3K, PLC- γ , and PTEN. It is significant that the β -catenin and PI3K pathways were not altered by *mda-7/IL-24* expression in normal HUVEC cells.

Activation of the Fas-FasL signaling pathway occurred in the human ovarian cancer cell line MDAH 2774 following infection with Ad.*mda-7*, where Ad.*mda-7* significantly inhibited cell proliferation and induced apoptosis. Early *mda-7/IL-24*-induced activation of the transcription factors c-Jun and activating transcription factor 2 (ATF2), which stimulated the transcription of an immediate downstream target, the death-inducer Fas ligand (FasL), and its cognate receptor Fas. The activation of NF- κ B and induction of Fas-associated factor 1, FADD, and caspase-8 were associated with the activation of Fas-FasL (64). However, another study by Emdad *et al* (65) reported p38 MAPK activation upon adenovirus-mediated *mda-7* infection in several ovarian cancer cell lines. The level of phosphorylated p38 MAPK correlated with the killing effect of *mda-7/IL-24* in these cells, and SB203580 (selective p38 MAPK inhibitor) significantly abolished this killing effect. Thus, activation of p38 MAPK presents a more general effect than induction of the Fas-FasL pathway and might be a key element in *mda-7/IL-24*-induced apoptosis in ovarian carcinoma cell lines as well as in other cancer cell lines (46,50,61,65).

3. *mda-7/IL-24* expression radiosensitizes cancer cells

Radiotherapy is a common option for cancer therapy; however, because of high toxicity at therapeutic doses the efficiency of this treatment is often suboptimal. Ionizing radiation (IR) generates hydroxyl radicals in water that can impact the function of mitochondria in cells, resulting in amplification of the amount of reactive oxygen and nitrogen species (66,67). In studies using human malignant glioma (26,37,38,61) and prostate carcinoma (40), the ability of both IR and *mda-7/IL-24* to generate ROS was directly linked to the radiosensitizing properties of this gene. Additional studies in ovarian carcinoma (65), NSCLC (23,62) and breast cancer cells (58) confirmed the radiosensitizing ability of *mda-7/IL-24* gene. Studies *in vitro* highlighted the importance of JNK activation as a mediator of radiosensitization effect and its p53 independence (6,26,37,38,40,61,62).

In glioma cells, the antiproliferative effects of Ad.*mda-7* were enhanced by IR in a greater than additive manner (37). The radiosensitizing effects were not observed in non-transformed primary human fetal astrocytes. Growth inhibition of glioma cells *in vitro* correlated with an increased number of cells in the G₁/G₀ and G₂/M phases of the cell cycle, confirming cell cycle independence of the radiosensitization effect (61). Irradiation of cells, but not Ad.*mda-7* infection itself, dramatically enhanced JNK 1/2 activity. Accordingly, inhibition of JNK 1/2 with its specific inhibitor SP600125 abolished radiosensitization properties of MDA-7/IL-24. Both Ad.*mda-7* and recombinant GST-MDA-7/IL-24 protein demonstrated radiosensitizing properties in malignant glioma cell lines, and this effect correlated with increased expression of specific members of the *GADD* gene family, particularly, *GADD153* (37). Similarly, antiproliferative and cytotoxic properties of recombinant GST-MDA-7/IL-24 protein were enhanced by radiation in primary (non-established) human glioma cells (38). *In vivo*, Ad.*mda-7* enhanced the survival of Fischer 344 rats implanted intracranially with rat malignant glioma cells. Radiation did not alter survival in control-infected

animals, whereas it prolonged survival in animals infected with Ad.*mda-7* (26).

GST-MDA-7/IL-24 protein also demonstrated radiosensitizing effect *in vitro* in breast cancer cells MDA-MB-231 (53). Moreover, ectopic *mda-7/IL-24* expression (or treatment with purified recombinant GST-MDA-7/IL-24 protein) in combination with IR was able to overcome both radioresistance and *de novo* resistance to *mda-7/IL-24*-induced apoptosis in prostate cancer cells overexpressing Bcl-2/Bcl-x_L (40). JNK activation was documented to be a central event in the radiosensitization of cancer cells and blocking of JNK activation ameliorated this radiation enhancement effect (40). Considering the fact that elevated expression of Bcl-x_L and Bcl-2 are frequent events in prostate cancer development and progression, the results of this study support the use of IR in combination with *mda-7/IL-24* to augment the therapeutic benefit of this gene in prostate cancer, particularly in the context of tumors displaying resistance to radiation therapy owing to Bcl-2 family members overexpression.

In NSCLC, Ad.*mda-7* infection enhanced radiosensitivity independently of the *p53* status of the cells and independently of any ability to up-regulate the expression of Fas or BAX. Phosphorylated c-Jun expression was demonstrated in NSCLC cell lines treated with both IR and Ad.*mda-7*, but not in normal human lung fibroblasts, CCD-16 (62). Nishikawa *et al* (23) demonstrated that not only did Ad.*mda-7* infection in combination with radiation enhance apoptosis in tumors, but also that secreted MDA-7/IL-24 protein sensitized HUVEC cells to IR thus inhibiting angiogenesis. Further studies revealed that Ad.*mda-7* sensitizes NSCLC cells to IR by suppressing the activity of non-homologous end-joining pathway of double strand break repair (68). A recent study by Chada *et al* (58) described a synergistic effect of the combination of Ad.*mda-7* and IR on breast cancer cells *in vitro* and *in vivo*, however, the mechanism of this effect was not investigated.

Emdad *et al* (65) demonstrated that *mda-7/IL-24*-induced apoptosis in ovarian cancer cells could be significantly enhanced by combination with ionizing radiation. Additionally, *mda-7/IL-24* gene delivery under the control of a minimal promoter region of progression elevated gene-3 (*PEG-3*), which functions selectively in diverse cancer cells with minimal activity in normal cells (69-74), displayed a selective radiosensitizing effect in ovarian cancer cells (65).

In summary, studies in multiple human tumor model systems, including malignant glioma, and carcinomas of the lung, breast, prostate and ovary (26,37,38,40,58,61,62,65), suggest that the therapeutic index of *mda-7/IL-24* can be enhanced by its use in combination with IR. The ability of secreted *mda-7/IL-24* to inhibit tumor angiogenesis, which is promoted by IR, provides another reason for using this combinatorial approach for cancer therapy. Of additional importance, the ability of the combination of *mda-7/IL-24* and IR to reverse resistance in specific cancers, such as prostate carcinoma cells overexpressing Bcl-2 or Bcl-x_L, to either agent used alone, suggests expanded applications of this novel cytokine in the context of potentially resistant tumor cells. Further studies are necessary to confirm improved antitumor activity in patients by combining *mda-7/IL-24* with IR.

4. Ectopic expression of *mda-7/IL-24* inhibits invasion and migration of cancer cells

Abnormalities in smooth muscle cell (SMC) proliferation and differentiation underlie the pathogenesis of proliferative vascular diseases. Due to *mda-7/IL-24*'s involvement in apoptosis and differentiation processes in diverse cancers, this gene was used to modulate SMC growth and migration (75). Overexpression of *mda-7/IL-24* suppressed growth of PAC1 rat cells (exhibiting multiple chromosomal aberrations) and induced apoptosis in this cell line in a dose-dependent manner; this treatment had no effect on normal primary human coronary artery cells or rat aortic SMC despite the high levels of MDA-7/IL-24 expression in all SMC. Ad.*mda-7* also inhibited serum-stimulated PAC1 cell migration. In contrast, recombinant MDA-7/IL-24 protein did not elicit death or STAT3 activation in PAC1 cells that lacked IL-20/IL-22 receptors suggesting that the effects of Ad.*mda-7* were mediated through intracellular pathways.

When applied to human non-small cell lung carcinoma cells (NSCLC), Ad.*mda-7* inhibited migration and invasion of H1299 and A549 cells (76). Mechanistic studies revealed down-regulation of PI3K/PKB, FAK and MMP-2 and -9 following Ad.*mda-7* infection. Furthermore, tumor cells treated with Ad.*mda-7* *ex vivo* or with DOTAP:Chol-*mda-7/IL-24* complex *in vivo* formed significantly less tumors in an experimental lung metastasis model. Both migration and invasion processes played pivotal roles in cancer metastasis. Thus, MDA-7/IL-24-based drugs may provide a novel therapeutic strategy that can inhibit tumor growth directly by inducing apoptosis, and may also prevent tumor invasion, ultimately reducing metastasis.

Sauane *et al* (45) compared the effect of secreted and non-secreted forms of MDA-7/IL-24 on C8161 tumor cell invasiveness. The C8161 cell line is characterized by very high invasive capacity *in vitro* and by tumorigenic and metastatic properties *in vivo* in nude mice (77). Infection of C8161 cells with Ad.SP-*mda-7* (an adenovirus producing a version of *mda-7/IL-24* that is not secreted because of the absence of the secretory signal peptide at amino acids 1-49) or with Ad.*mda-7* inhibited to a similar extent the ability of these cells to invade through Matrigel-coated membrane inserts as compared with Ad.*vec* infected cells. In these experimental settings, no effect was observed on C8161 cell growth in monolayer, thereby confirming that invasiveness was not inhibited because of loss of cell viability (45).

Moreover, MDA-7/IL-24 secretion by normal cells inhibited invasion of cancer cells expressing the complete set of IL-20/IL-22 receptors, such as DU-145 or BxPC-3, but not A549 cells that lack a full complement of IL-20/IL-22 receptors (35). In these experiments, P69 immortalized normal human prostate epithelial cells were infected with Ad.*mda-7* and cocultivated with different cancer cells for 48 h and then evaluated for invasion. Under these conditions, no decrease in cell growth or viability was evident, and no apoptosis was induced in these cancer cells (35). These results indicate 'antitumor (invasion inhibition) bystander effect' of secreted MDA-7/IL-24 protein can be expected to enhance the therapeutic properties of this cytokine and promote biologically relevant effects at a distance from its initial cellular site of synthesis and secretion.

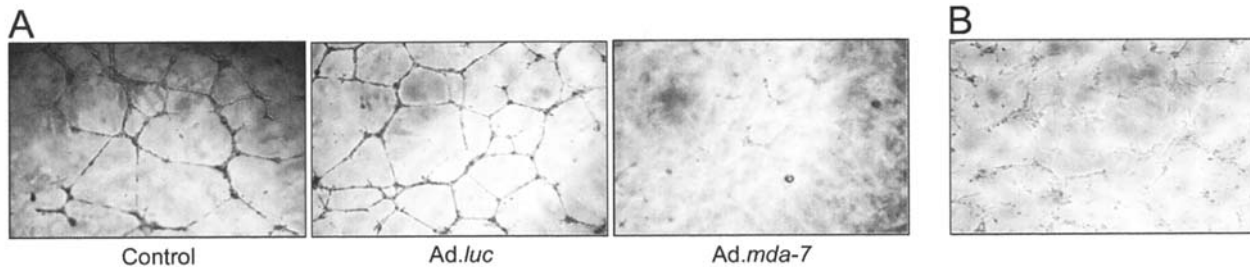


Figure 3. Demonstration of *mda-7/IL-24*-mediated antiangiogenic activity by tumor-endothelial cell mixing experiment *in vitro*. A, Human lung tumor cells (H1299) treated with PBS, Ad.*luc* or Ad.*mda-7* were plated in 96-well plates and overlaid with Matrigel. HUVEC seeded on top of the Matrigel were analyzed for ECD or tube-formation. Significant inhibition of ECD was observed in wells that contained Ad.*mda-7* treated tumor cells compared to PBS or Ad.*luc* treated tumor cells. B, Photomicrograph showing H1299 cells at the bottom of the Matrigel coated wells.

5. Antiangiogenic activity of *mda-7/IL-24*

Angiogenesis, a complex process that involves many cell types and a finely orchestrated series of events that include endothelial cell proliferation, new capillary formation, attraction of pericytes and macrophages, disruption of existing extracellular matrix and deposition of new matrix, is a critical process required by solid tumors to support their growth (78,79). The role of angiogenesis in growth and metastasis of tumors is well established and recent studies support the concept that metastasis from solid tumors is facilitated by angiogenesis of the primary tumor (80). In these contexts, inhibition of tumor angiogenesis should prove to be an effective means of inhibiting cancer growth and spread.

A number of angiogenesis inhibitors have been identified and found to block tumor growth by inhibiting tumor angiogenesis in preclinical studies (81-91). Angiogenesis may be regulated on multiple levels: some inhibitors may function to impede the formation of new blood vessels (antiangiogenic), whereas others may disrupt or modify existing vessels (anti-vascular). Although several antiangiogenic molecules have been developed and tested only a few have demonstrated a therapeutic effect in clinical trials (81,82). Based on the potential relevance of this approach and its limited current success, there has been a need to identify and characterize new and novel agents that target the tumor and its vasculature. Below, we summarize preclinical findings showing the antiangiogenic activity of *mda-7/IL-24*.

Initial studies testing the antitumor activity of *mda-7/IL-24* using an adenoviral vector (Ad.*mda-7*) demonstrated that Ad.*mda-7* was cytotoxic to a panel of human lung tumor cells, but not to endothelial cells *in vitro* (24). However, *in vivo* molecular analysis of the growth inhibitory effects showed Ad.*mda-7*-treated tumors had reduced tumor vascularization compared to control vector-treated tumors (24). These results suggested that the reduced tumor vascularization in Ad.*mda-7*-treated tumors could be either due to direct tumor killing or antiangiogenic activity. To test whether *mda-7*-mediated an antiangiogenic activity, *in vitro* and *in vivo* experiments were conducted. In *in vitro* studies, Ad.*mda-7*-treated human H1299 lung tumor cells that served as an ectopic source for MDA-7/IL-24 protein were mixed with human umbilical vein endothelial cells (HUVEC) and plated onto Matrigel coated 96-well plates and observed for endothelial cell differentiation (ECD), an assay that is routinely used to test for antiangiogenic activity. Tumor cells that were treated with PBS

or infected with Ad.*luc* (vector control) and mixed with HUVEC served as controls. A marked inhibition of ECD was observed in wells containing Ad.*mda-7*-treated tumor cells (Fig. 3). No inhibition of ECD was observed in wells that contained PBS or Ad.*luc*-treated tumor cells. These results showed that tumor cells exogenously expressing MDA-7/IL-24 protein when in close proximity to endothelial cells could inhibit ECD. These observations confirm that *mda-7/IL-24* has antiangiogenic activity.

Although results from *in vitro* mixing experiments supported *in vivo* findings for *mda-7/IL-24*-mediated antiangiogenic activity, the exact mechanism was not known. Furthermore, previous studies had shown that tumor angiogenesis could be inhibited directly by suppressing tumor blood vessel formation and indirectly by suppressing the production of tumor-derived growth factors, such as vascular endothelial growth factor (VEGF), fibroblast growth factor (FGF) and interleukin-8 (IL-8) (78-81). Thus, it was possible that *mda-7/IL-24* exerted its antiangiogenic activity by a direct or an indirect mechanism or by a combination of both processes. Confirmation of direct and indirect inhibitory effects of *mda-7/IL-24* on tumor angiogenesis is discussed below.

Direct inhibitory effect. To test whether *mda-7/IL-24* directly affected the tumor vasculature, experiments were performed using affinity purified human MDA-7/IL-24 protein (7). *In vitro*, MDA-7/IL-24 protein selectively inhibited ECD with no effect on cell proliferation (92). The inhibitory effect on ECD was dose-dependent with complete inhibition occurring at concentrations above 10 ng/ml. Note, at the concentrations of MDA-7/IL-24 protein used no significant cytotoxicity against receptor-negative lung tumor cells was observed indicating selective activity against receptor-positive endothelial cells. That the inhibitory effect was specific to MDA-7/IL-24 protein was demonstrated by immunodepletion studies (92). Additionally, the inhibitory effect of MDA-7/IL-24 protein on ECD was 10-50 times more potent than endostatin, interferon- γ and interferon-inducible protein-10 when tested at equimolar concentrations *in vitro* (92). Additional studies showed that MDA-7/IL-24 protein effectively inhibited in a dose-dependent manner VEGF and FGF-induced endothelial cell migration (92). These results demonstrate direct antiangiogenic activities for the MDA-7/IL-24 protein.

Molecular analysis of MDA-7/IL-24 protein-mediated antiangiogenic activity showed a requirement for IL-22R1, one of the two receptors for MDA-7/IL-24 (32,33,92). The

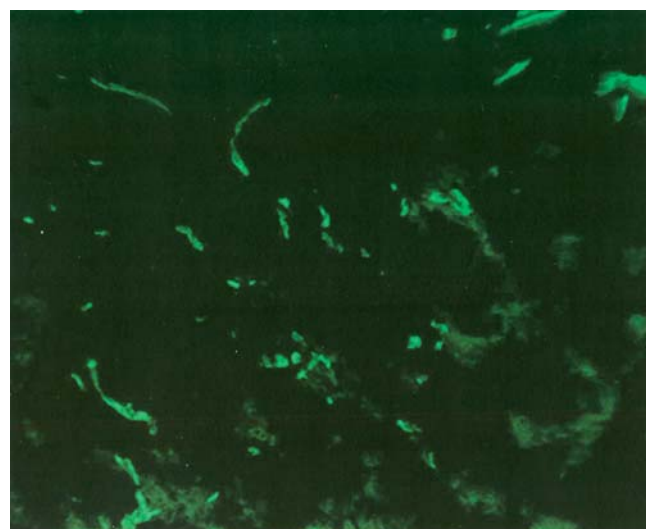


Figure 4. Antiangiogenic activity of *mda-7/IL-24* *in vivo*. Human A549 lung tumor cells were mixed with parental 293 cells (A549 + 293) or mixed with 293-*mda-7* cells (A549 + 293-*mda-7*) and implanted subcutaneously into the lower right flank of nude mice. Tumor growth was measured at regular intervals and experiments terminated after 3 weeks. A significant inhibition of tumor growth was observed in mice implanted with a mixture of A549 + 293-*mda-7* cells (bottom) compared to mice implanted with a mixture of A549 + 293 (top).

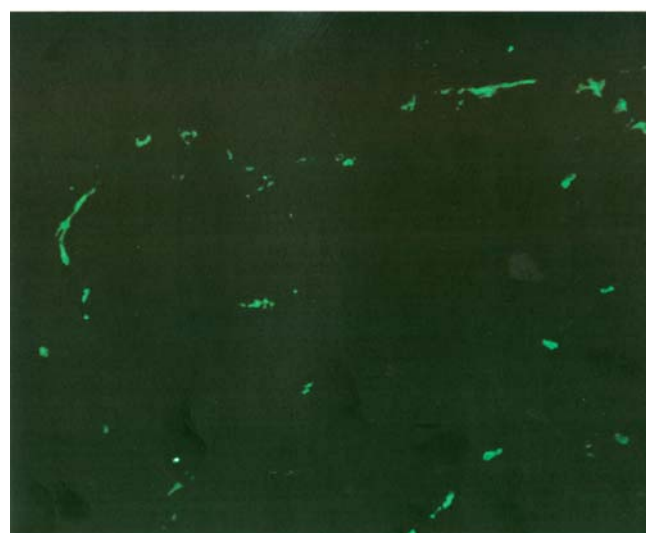
involvement of IL-22R for MDA-7/IL-24 protein-mediated antiangiogenic activity was demonstrated by activation of STAT-3 in HUVEC as a measure of receptor-ligand interaction and by receptor-blocking studies (92). Although involvement of IL-22R in MDA-7/IL-24 protein-mediated antiangiogenic activity has been demonstrated, the role of the IL-20 receptor in *mda-7/IL-24*-mediated antiangiogenic activity has yet to be determined. Additionally, the downstream signaling mechanisms regulated by *mda-7/IL-24* in endothelial cells has not been studied. One possibility is the activation of STAT-1, a molecule that is associated with antiangiogenic phenotype (93). Another possibility is *mda-7/IL-24*-mediated inhibition of the PI3K/AKT pathway that is associated with a proangiogenic phenotype. Recent studies have demonstrated PI3K/AKT inhibition in Ad.*mda-7*-treated lung and breast cancer cells (63,94).

Evidence for direct effects of MDA-7/IL-24-mediated antiangiogenic activity *in vivo* has been obtained. Subcutaneous implantation of MDA-7/IL-24 producing 293 cells (293-*mda-7*) mixed with receptor negative A549 lung tumor cells (1:1 ratio) in nude mice demonstrated suppression of tumor growth compared to tumor growth in mice implanted with a mixture of parental 293 cells and tumor cells [Fig. 4; (92)]. Tumor growth inhibition was accompanied by apoptosis of tumor endothelial cells and tumor cells, reduced hemoglobin content, and less CD31 positive endothelial cells (92). These results demonstrate the direct antiangiogenic activity of MDA-7/IL-24 protein.

To test whether MDA-7/IL-24 protein can inhibit tumor growth at a distant site systemically, A549 lung tumor xenografts were established subcutaneously in the lower right flank of nude mice. Subsequently when the tumors had grown to a size of 50-100 mm³, Matrigel containing parental 293 cells or Matrigel containing 293-*mda-7* cells were implanted subcutaneously into the upper right flank of tumor-bearing animals and the effects of *mda-7/IL-24* on tumor growth were measured. A significant growth inhibition with 40-50% reduction in tumor size was observed in mice that were implanted with Matrigel containing 293-*mda-7* cells compared to mice that were implanted with Matrigel containing



Tumor + 293 cells



Tumor + 293-*mda-7* cells

Figure 5. Systemic inhibition of tumor vascularization by *mda-7/IL-24*. Subcutaneous A549 lung tumor xenografts were established in nude mice in the lower right flank. When the tumors were measurable, Matrigel encapsulated 293 or 293-*mda-7* cells were implanted in the upper right flank. At 3-4 weeks after initiation of the experiment animals were euthanized, tumors harvested and tissue sections prepared, and stained for CD31 using FITC-conjugated antimouse CD31 antibody. A significant decrease in CD31 positive blood vessels was observed in tumors that were obtained from mice implanted with 293-*mda-7* cells compared to tumors that were obtained from mice implanted with 293 cells.

293 cells (92) (Fig. 5). That the observed tumor growth inhibition was due to systemic inhibitory effects of MDA-7/IL-24 protein on tumor angiogenesis was demonstrated by detecting the protein in the blood and a reduction in CD31 positive blood vessels (92). Furthermore, no gross pathological changes were observed in the animals implanted with 293-*mda-7* cells suggesting that the protein had no toxic side effects. These results demonstrated MDA-7/IL-24 protein systemically inhibits tumor growth by inhibiting tumor angiogenesis. Although demonstration of the antiangiogenic activity of *mda-7/IL-24* is exciting, it should be noted that

these studies were conducted in an immunodeficient nude mouse model. It is of interest to test the systemic effects of *mda-7/IL-24* on tumor angiogenesis in an immunocompetent mouse model as well as to evaluate for toxicity.

Intratumoral or systemic administration of *mda-7*-nanoparticles into immunocompetent C3H mice bearing syngeneic UV2237m subcutaneous tumors or experimental lung metastases resulted in tumor growth inhibition (20,76). Associated with growth inhibition was the demonstration of reduced vascularization demonstrating *mda-7/IL-24*-mediated antiangiogenic activity. Additionally, no measurable toxicity was observed. The results from this study showed that the anti-tumor activity of *mda-7/IL-24* in immunocompetent tumor-bearing mice is partly mediated by inhibiting angiogenesis with no toxicity, a phenomenon akin to that seen in immunodeficient tumor-bearing mice.

Indirect inhibitory effect. Although direct antiangiogenic activity for MDA-7/IL-24 protein was demonstrated (92), MDA-7/IL-24 like many other antiangiogenic agents can also indirectly exert its antiangiogenic activity by inhibiting tumor-derived growth factor (IL-8, bFGF, VEGF) expression (95-97). Thus, inhibition of these growth factors will result in the suppression of tumor growth due to its inability to support neovascularization. Supporting this possibility is the recent report by Nishikawa *et al* (23) who showed inhibition of VEGF, bFGF, and IL-8 in *Ad.mda-7*-treated human lung tumor xenografts. In the same study, combining radiation therapy with *Ad.mda-7* documented enhanced radiosensitization of lung tumor xenografts that was associated with a significant inhibition of VEGF, bFGF, IL-8 and tumor neovascularization. In fact, combination therapy showed greater inhibition of these proangiogenic factors compared to radiation or *Ad.mda-7* treatment alone. Thus, a role for indirect antiangiogenic activity for *mda-7/IL-24* was established. However, the molecular mechanism by which *mda-7/IL-24* exerted its inhibitory activity on these growth factors was neither examined nor reported.

Recent studies demonstrate that *mda-7/IL-24* inhibits VEGF expression at the transcriptional level resulting in reduced VEGF protein expression [Fig. 6; (98)]. Molecular analysis revealed that Src kinase regulated VEGF transcription and protein expression and that *mda-7/IL-24* directly inhibited Src kinase activity, thereby resulting in reduced VEGF transcription and expression. Additionally, tissue culture supernatant collected from *mda-7/IL-24*-treated tumor cells that contained reduced VEGF, as determined by ELISA, when added to actively growing endothelial cells showed reduced VEGFR2 signaling and Akt activation leading to inhibition of cell proliferation and induction of apoptosis (98). In contrast, addition of culture supernatants from tumor cells that were treated with PBS or *Ad.luc* (vector control) and contained high levels of VEGF resulted in increased endothelial cell proliferation that was associated with increased VEGFR2 and Akt activation. Thus, *mda-7/IL-24*-mediated inhibition of VEGF expression in tumor cells results in the inability of the tumor to support endothelial cell survival. These results showed that *mda-7/IL-24* could inhibit angiogenesis indirectly by inhibiting tumor-derived growth factor expression, such as VEGF. Additional growth factors that were inhibited by *mda-7/IL-24* include IL-8, FGF, and TGF- β (Inoue *et al*,

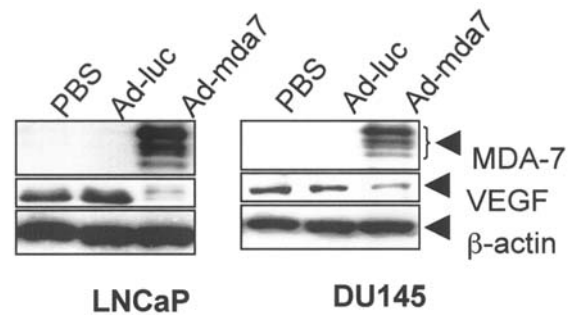


Figure 6. Inhibition of VEGF by *mda-7/IL-24*. Human prostate tumor cells (LNCaP and DU-145) seeded in 6-well tissue culture plates were treated with PBS, *Ad.luc* or *Ad.mda-7*. At 72-h after treatment, cells were harvested, cell lysate prepared and subjected to Western blotting for VEGF protein expression. Inhibition of VEGF that correlated with MDA-7/IL-24 protein expression was observed in *Ad.mda-7*-treated tumor cells and not in PBS- or *Ad.luc*-treated cells. VEGF was inhibited in both LNCaP and DU-145 tumor cells. β -actin was used as a loading control.

unpublished data). In conclusion, *mda-7/IL-24* exerts its antiangiogenic activity both by direct and indirect mechanisms that are mediated by the extracellular and intracellular protein, respectively.

6. Immune properties of *mda-7/IL-24*

Several features of *mda-7/IL-24* characterize this gene as a cytokine and a member of the IL-10 family of cytokines. These features include chromosomal location on chromosome 1q31-32 in a region containing a cluster of IL-10 family member genes (3,5), translational regulation (42,44,99-101), a secretory motif, and an IL-10 signature motif. Several experimental studies document that MDA-7/IL-24 protein can be secreted and that purified MDA-7/IL-24 protein can bind to IL-20R1/IL-20R2 and IL-20R1/IL-22 receptor complexes and activate STAT-signaling pathways (7,12,34,35). Physiological expression of the *mda-7/IL-24* gene occurs in a highly restricted manner in human cells, and associates with cells having immunological functions (5,7). Several studies show expression of *mda-7/IL-24* both on an mRNA and protein level in melanocytes and a progressive loss of expression in melanoma (2,5,10-13). Loss of *mda-7/IL-24* expression in progressing melanoma might contribute to the ineffective immune response to melanoma (102).

Earlier studies based on multiple tissue Northern blots containing poly(A) RNAs from normal human tissue demonstrated *mda-7/IL-24* tissue specific expression in thymus, spleen and peripheral blood mononuclear cells (PBMCs) compared to several other tissues (5). Two broad classes of cells were shown to endogenously express *mda-7/IL-24*, including melanocytes and subpopulations of PBMCs. These included normal unstimulated or LPS stimulated monocytes, anti-CD3 stimulated T-cells, and normal human melanocytes (5,7,9).

A potential role for *mda-7/IL-24* as a cytokine and its involvement in the immune system has been highlighted in two studies (7,9). *Mda-7/IL-24* expression could be induced in PBMCs upon treatment with LPS or PHA (7). In addition, *mda-7/IL-24* mRNA could be detected by real-time PCR in monocytes and its expression was up-regulated in monocytes

by treatment with LPS or in T-cells, especially in CD4⁺ naïve and memory cells, following activation by anti-CD3 mAb (9). No expression has been shown in NK or B-cells before or after stimulation. Further studies in T-cells demonstrated that at earlier time points (6 h), *mda-7/IL-24* expression was down-regulated in cells undergoing T1 differentiation (mediating cellular immunity) and slightly up-regulated in cells undergoing T2 differentiation (regulating humoral immunity). At 66 h, the expression increased in cells committed to T1 differentiation (9). However, the expression pattern in mouse T cells is different from that in humans. The mouse counterpart of *mda-7*, FISP, was shown to have exclusive expression in Th2 lymphocytes (103).

Treatment of PBMCs with MDA-7/IL-24 protein, purified from conditioned media of a 293-cell line stably expressing *mda-7/IL-24*, resulted in the induction of IL-6, IFN- γ , TNF- α , IL-1 β , IL-12, and GM-CSF (granulocyte macrophage-colony stimulating factor) (7). These inductions were blocked, either completely or partially, by simultaneous administration of IL-10, which might occur due to sharing of receptor subunits by IL-10 family and due to a 10-fold higher affinity of IL-10 for its receptors (9). However, treatment with MDA-7/IL-24 did not modulate the proliferative functions of PBMCs (7). It was hypothesized that the secondary cytokines induced by MDA-7/IL-24 might activate antigen presenting cells to present tumor antigen, thereby triggering an antitumor immune response (7). In PBMC cultures, the cytokine activity of MDA-7/IL-24 was antagonized by the prototypic immunosuppressive cytokine IL-10. These results suggest that even though MDA-7/IL-24 and IL-10 are related family members and share 19% amino acid homology, they bind to different receptors and appear to have opposing roles in regulating immunity against incipient tumors. In melanoma, pathologic progression correlates with increased IL-10 levels (104), therefore IL-10 can function as an autocrine growth factor for melanoma. Expression of IL-10 in melanoma and other tumors may facilitate loss of immune surveillance by blocking tumor antigen presentation and thereby inhibiting cell-mediated immunity (9). In contrast, MDA-7/IL-24 induces cell cycle arrest and death in melanoma cells, therefore *mda-7/IL-24*-based drugs may be especially efficient for melanoma treatment because they restore a protein involved in the genesis and progression of this disease.

Garn *et al* (105) demonstrated expression of *mda-7/IL-24* in response to stimulation by various members of the IL-10 family of cytokines in mouse and human macrophages. LPS or IL-4 (but not TNF- α) treatment of rat alveolar macrophages resulted in *mda-7/IL-24* mRNA and protein expression. Induction of *mda-7/IL-24* mRNA correlated with expression of IL-1, IL-6, and TNF- α in cultured human monocytes infected with influenza virus confirming a proinflammatory role for this molecule.

More recent studies of *mda-7/IL-24* induction in human PBMCs stimulated by PHA and LPS or by allogeneic major histocompatibility complex confirmed data presented by Caudell *et al* (7), Wolk *et al* (9), and Garn *et al* (105) and demonstrated that the expression of *mda-7/IL-24* in human PBMCs results from cytokine stimulation and is regulated at the post-transcriptional level through stabilization of *mda-7/IL-24* mRNA (106). Individual cytokines including IL-2, IL-7,

IL-15, TNF- α , GM-CSF and IL-1 β stimulate the expression of *mda-7/IL-24* mRNA and protein, whereas interferons and Th2 type cytokines fail to induce substantial *mda-7/IL-24*. Since cytokines stimulating *mda-7/IL-24* production were secreted from cells involved in an inflammatory response, the authors hypothesized that *mda-7/IL-24* might play an important role in the inflammatory response in the skin occurring as a result of a danger signal initiated by viral or bacterial infection or tumor cell growth. Even though MDA-7/IL-24 protein induced inflammatory cytokines and activates inflammatory and apoptotic signaling proteins, it did not activate NF- κ B alone, but significantly enhanced TNF- α -induced NF- κ B activation and NF- κ B-regulated expression of cyclin D1 and COX-2 (107). Potentiation of TNF- α -induced NF- κ B activation by MDA-7/IL-24 may occur through MDA-7/IL-24-induced PKR activation, since it is known that PKR activates NF- κ B by phosphorylation of I κ B α (108) or by interacting with IKK (109,110). The effects described by Aggarwal *et al* (107) appear to be receptor-independent.

In addition to direct tumor cell killing of transformed human and rodent cells (14,17,18), *mda-7/IL-24* gene transfer can also elicit immunotherapeutic activity against syngeneic murine melanoma tumors (111). UV2237m murine fibrosarcoma cells infected with Ad.*mda-7* were not able to grow after injection into syngeneic immunocompetent C3H mice, in contrast with immunocompromised nude mice, in which all mice developed tumors after the injection. The authors of this study hypothesized that Ad.*mda-7* treatment of tumors would result in local MDA-7/IL-24 protein secretion, which would induce or promote systemic antitumor immunity. As predicted, when challenged with parental tumor cells, these tumor-free C3H mice that previously received Ad.*mda-7/IL-24* treatment experienced no tumor growth. Splenocytes prepared from vaccinated C3H mice demonstrated higher proliferative activity and produced elevated levels of TH1 cytokines, such as TNF- α , GM-CSF, IL-6 and IFN- γ . These findings corroborate the *in vitro* data of Caudell *et al* (7), which showed induction of TH1 cytokines in human PBMCs following treatment with purified human MDA-7/IL-24 protein. Production of IFN- γ , TNF- α and IL-6 was also reported in the serum of patients treated intratumorally with Ad.*mda-7/IL-24* (27). An *in vitro* subset analysis of splenocytes from vaccinated mice demonstrated a significant increase in the CD3⁺CD8⁺ but not the CD3⁺CD4⁺ cell population. These data are also in agreement with the results from a recently finished clinical trial where patients that received an intratumoral injection of Ad.*mda-7* (INGN 241) experienced a marked increase in CD3⁺CD8⁺ T-cells after treatment (27). The exact mechanism of *mda-7/IL-24*-induced systemic immunity is still unknown. One possibility is that antigen-presenting cells take up for presentation T-cell immunogenic apoptotic bodies resulting from *mda-7/IL-24*-mediated apoptosis. Another possibility considers activation of immune accessory and effector cells present within the tumor microenvironment and at distant sites by secreted MDA-7/IL-24 protein (111).

7. 'Bystander antitumor' activity of *mda-7/IL-24*

Multiple experiments performed *in vitro* and *in vivo* confirm the remarkable ability of *mda-7/IL-24* to induce apoptosis in

most cancers. However, pancreatic cancer cells display an inherent resistance to the adenovirus-mediated *mda-7* infection (44). Pancreatic cancer is a complex disease where multiple subsets of genes undergo genetic changes during tumor development and progression. One of the most frequent and earliest genetic alterations in pancreatic cancer (85-95%) involves activation of the *K-ras* oncogene (112). The lack of activity of Ad.*mda-7* is not a consequence of failure to express *mda-7/IL-24* mRNA. Rather, it involves a translational suppression in conversion of this mRNA into protein (42,44,99). When expression of mut *K-ras* is down-regulated by means of antisense oligonucleotides, plasmid or adenovirus expressing antisense *K-ras*, pancreatic cancer cells with mut *K-ras* become sensitive to Ad.*mda-7*-induced apoptosis (44). This observation itself provides a basis for developing rational targeted therapies for pancreatic cancer. Furthermore, these original studies using antisense phosphorothioate oligonucleotides or DNA plasmid transfection to down-regulate *K-ras* lead to another important and provocative observation of 'bystander antitumor' activity *in vitro* and *in vivo*. Tumor formation was suppressed when Ad.*mda-7*-infected cells (estimated to be ~100% of the cells) were combined with transfection with an antisense *K-ras* plasmid (likely transducing ~3-5% of the cells). Nevertheless, the growth of the whole population of tumor cells was efficiently suppressed, a phenomenon also observed in the *in vitro* setting. This supports a 'bystander antitumor' effect in which tumor cells receiving both *mda-7/IL-24* and antisense *K-ras*-induced changes, likely resulting from the secretion of MDA-7/IL-24 protein, that led to cell death in tumor cells with none or only one of these two genetic manipulations. Additional early evidence of 'bystander antitumor' effect derives from experiments where primary rodent hepatocytes were infected with Ad.*mda-7* resulting in the production and secretion of bioactive MDA-7/IL-24, which when applied to DU-145 cells suppressed their growth *in vitro* (Su *et al*, unpublished data). Understanding the mechanism underlying this 'bystander antitumor' effect could provide new therapeutic insights as well as help in defining ways of improving the anticancer properties of *mda-7/IL-24* (14).

Evidence for 'bystander antitumor' activity was recently provided *in vivo* in human tumor xenograft nude mouse animal studies (73). A conditionally replication competent adenovirus (CRCA) was produced in which replication (E1A function) was controlled by the cancer-specific promoter derived from the *PEG-3* gene (71,113) to drive replication in tumor cells and *mda-7/IL-24* expression was controlled by the cytomegalovirus promoter in the E3 region of the adenovirus, Ad.PEG-E1A-*mda-7* (73). Human breast carcinoma cells were inoculated into the right and left flanks of athymic nude mice producing tumors at both sites. Tumors that formed in the left side of the animal were injected with Ad.PEG-E1A-*mda-7* resulting in eradication of this tumor (reflective of a primary tumor) as well as elimination of the tumor on the right side of the animal (reflective of a metastatic state). Additionally, injection with a non-replicating Ad.*mda-7* into the left tumor resulted in tumor regression and a statistically significant decrease, although not a complete eradication, was also evident in the right tumor, confirming 'bystander antitumor' activity of the secreted MDA-7/IL-24. The potent

inhibitory effects on distant tumors could be explained by a direct action of the secreted cytokine following interaction with its cognate IL-20/IL-22 receptors that activate signal transduction pathways mediating antitumor activity (34,35). Alternatively, the potent 'bystander antitumor' effect of MDA-7/IL-24 on distant tumor cells may include stimulation of NK cells, since athymic nude mice used in the experiments still have a spleen and a liver and display potent NK cells activity (114).

Three recent studies investigated the mechanism(s) of the 'antitumor bystander' effect exerted by MDA-7/IL-24 (34,35, 115). Transduction of both tumor and normal cells with Ad.*mda-7* gene results in secretion of glycosylated MDA-7/IL-24 protein (reviewed in refs. 4,14-16,18). While Ad.*mda-7* and GST-MDA-7 recombinant protein were able to selectively kill cancer cells in a receptor-independent manner involving both a secretory and non-secretory mode of action (45,53), the ability of secreted MDA-7/IL-24 to suppress tumor cell growth, inhibit invasion and radiosensitize cancer cells appears to be receptor-dependent. As a cytokine belonging to the IL-10 family, secreted MDA-7/IL-24 protein can bind to two distinct type II cytokine heterodimeric receptor complexes, type 1 IL-20R (IL-20R1/IL-20R2), and type 2 IL-20R (IL-22R1/IL-20R2) (32,33). Ligand-mediated assembly of both receptor chains rapidly activates STAT3 and, to a lesser extent, STAT1 (30). In melanoma cells, both type 1 and type 2 IL-20R mediate transient STAT3 activation by secreted MDA-7/IL-24 protein. This activation is inhibited by >90% by anti-MDA-7/IL-24 neutralizing antibodies (34). Although MDA-7/IL-24 induces STAT3 activation and cytotoxicity in melanoma cells, these events are uncoupled. Moreover, even though all of the IL-10 family members tested elicited STAT3 activation in melanoma cells, MDA-7/IL-24 was the only cytokine that produced cell death. Using antibodies against type 1 IL20R and type 2 IL20R, Chada *et al* (34) demonstrated, also that MDA-7/IL-24-induced apoptosis in melanoma cells is mediated by type 1 IL-20R receptor, even though engagement of both type of receptors resulted in STAT3 activation. The authors hypothesized that the downstream signaling between MDA-7/IL-24 and other members of the IL-10 family differs or that MDA-7/IL-24 may use a co-receptor not utilized by other family members to activate apoptosis signaling. In a similar study, Zheng *et al* (115) demonstrated that apoptosis induced by exogenous MDA-7/IL-24 protein in breast cancer cells is mediated through IL-20R1/IL-20R2 heterodimeric receptor complex and is independent of STAT3 phosphorylation. Coadministration of the related cytokine IL-10 inhibited MDA-7/IL-24-induced killing and MDA-7/IL-24-induced up-regulation of the tumor suppressor proteins *p53* and *p27^{Kip1}* (115).

Su *et al* (35) provided evidence of a 'bystander antitumor' effect of *mda-7/IL-24* resulting from secretion of this cytokine by normal cells. Normal cells were infected with Ad.*mda-7* and directly co-cultured with cancer cells or co-cultured with cancer cells that were separated by an agar overlay, providing a diffusion model for testing for cytokine efficacy. Cell viability and apoptosis (direct co-cultivation), anchorage independent growth (agar diffusion), radiosensitivity (agar diffusion) and invasion (direct co-cultivation) of cancer cells *in vitro* were evaluated using appropriate assays. Primary and immortalized normal cells (PFHA, FM-516SV and P69)

were infected with Ad.*mda-7* resulting in the production and secretion of MDA-7/IL-24 protein, which modulated the anchorage-independent growth, invasiveness, survival and radiosensitivity of cancer cells that contained functional IL-20/IL-22 receptors, but not cancer cells that lacked a complete set of receptors. Moreover, the combination of radiation and MDA-7/IL-24 secreted from normal cells evoked a 'bystander antitumor' effect not only in cancer cells inherently radiosensitive or permissive for *mda-7/IL-24* killing, but also in tumor cells overexpressing the antiapoptotic proteins BCL-2 or BCL-x_L and displaying resistance to either treatment alone (35,40). Furthermore, since this effect can occur even in the presence of small amounts of secreted MDA-7/IL-24 protein (that are barely detectable by Western blotting using polyclonal antibodies to MDA-7/IL-24), secretion of this cytokine from normal cells should have a profound impact on tumor growth and radiosensitization *in vivo* by directly affecting tumor cell survival and by inhibiting angiogenesis. As described above, the recent study by Su *et al* (35) highlights the significance of normal cells as contributors to the potent antitumor activity of MDA-7/IL-24. A novel application of this cytokine to enhance its cancer therapeutic efficacy could exploit normal cells as a repository for producing low levels of MDA-7/IL-24 protein to facilitate both localized and systemic cancer therapies. If successful and non-toxic *in vivo*, constitutively expressing low levels of *mda-7/IL-24* would allow a maximization of benefit of this novel anticancer gene therapy, potentially providing a means of preventing, in real-time, tumor and metastasis development.

As described previously, another potential mechanism underlying *mda-7/IL-24*-mediated 'bystander antitumor' effect, including inhibition of cancer cell growth in distant tumors in an *in vivo* context, might be activation of the immune system. *Mda-7/IL-24* expression is restricted to melanocytes and those tissues associated with the immune system, such as spleen, thymus, and PBMCs. Expression of this gene induced in PBMCs with purified recombinant MDA-7/IL-24 protein resulted in the induction of multiple potent immunomodulatory agents such as IL-6, IFN- γ , TNF- α , IL-1 β , IL-12, and granulocyte macrophage-colony stimulating factor (7,27). These secondary cytokines induced by *mda-7/IL-24* might activate antigen-presenting cells to present tumor antigens, thereby triggering an antitumor immune response. Studies in a phase I clinical trial involving intratumoral injection of Ad.*mda-7* (INGN 241) suggest that these *in vitro* effects are recapitulated in the context of patients supporting the immune modulating properties of this cytokine (27). Although current studies were performed in athymic nude mice that are immunocompromised, these mice still have a spleen and a liver and display potent natural killer activity (74,114).

Another aspect of the 'bystander antitumor' activity of *mda-7/IL-24*, also elaborated on previously, relates to its potent antiangiogenic activity (24,92). This possibility is exemplified by studies by Nishikawa *et al* (23) using A549 non-small cell lung carcinoma cells that lack a complete set of IL-20/IL-22 receptors. Established tumors in animals were injected with Ad.*mda-7* and then exposed to radiation. Histological examination of tumors revealed a significant reduction in angiogenic factors (bFGF, VEGF) and microvessel density with enhanced apoptosis in tumors receiving the combination

of Ad.*mda-7* and radiation. Analysis of clonogenic survival of HUVECs, A549 and normal human lung fibroblasts pretreated with conditioned medium from 293 cells stably expressing *mda-7/IL-24* indicated sensitization of the HUVECs to radiation without affecting A549 or normal human lung fibroblasts. These results confirm that a significant enhancement in killing of A549 tumor cells was observed that was a consequence of secreted MDA-7/IL-24 that was inhibiting HUVECs and tumor angiogenesis. These results suggest increased antitumor activity of Ad.*mda-7* when combined with radiation in non-small cell lung carcinoma, as has also been shown in malignant gliomas and prostate and breast cancers (26,37,38,40,53,61).

A seminal proof of the 'bystander antitumor' effects of *mda-7/IL-24* derives from the recent phase I clinical trial (27,28). Administration of Ad.*mda-7* (INGN 241) intratumorally in patients with advanced carcinomas and melanomas resulted in apoptosis in tumor cells receiving the *mda-7/IL-24* transgene as well as death in tumor cells not initially receiving the virally-transduced gene. Immunohistochemical staining demonstrated that ~30% of the tumor mass received the *mda-7/IL-24* gene, however, ~70% of the tumor cells showed detectable apoptosis. This can reflect both a direct effect of secreted MDA-7/IL-24 or an immune-mediated antitumor response, or a combination of both processes.

The ability of a gene therapeutic to effectively manage cancer is dependent upon a number of factors and technical issues. Induction of direct tumor cell killing with the potential to be transmitted in the circulation to elicit an effect not only on a primary tumor, but also metastases, is a profoundly important component required for an effective cancer gene therapeutic. As highlighted in this review, *mda-7/IL-24* not only induced apoptosis selectively in cancer cells, but it also has immune modulatory, radiation enhancing, antiangiogenic properties and potent 'bystander antitumor' effects, thereby making it an ideal candidate for promoting effective cancer gene therapy (6,7,14,17,24,27,28,35,44,92). Because of these essential qualities as a potential therapeutic gene for cancer, a replication-incompetent adenovirus expressing *mda-7/IL-24* (Ad.*mda-7*; INGN 241) was evaluated for safety in a phase I clinical trial and based on these results (6,17,27-29), confirming safety with a demonstrable clinical benefit, *mda-7/IL-24* is currently being evaluated for clinical efficacy in a phase II clinical trial.

8. Translation of *mda-7/IL-24* into the clinic - A good beginning

Initial clinical testing of the *mda-7/IL-24* gene as a cancer therapeutic involved a non-replicating adenoviral (Ad) vector deleted in the E1 region (INGN 241) as the delivery vehicle (2,25). The initial trial was designed to prove the principle of mechanistic activity of Ad.*mda-7* and to demonstrate tolerability. Preclinical models justified single-agent testing of Ad.*mda-7* (4,6,8,11-13,15-17,22,24,25,37,116). Combination with chemotherapy, radiation therapy, and biological therapy demonstrated no increase in toxic profile of *mda-7/IL-24*-related adverse effects and suggested enhancement in immune-related activity. Preclinical toxicology studies of Ad.*mda-7* in mice demonstrated adverse effects only at the highest dose

Table I. Patient characteristics and dosing.

Cohort	No. of patients ^a	Age (range)	Previous treatments ^b	Dose (vp)	Adverse events ^c
1	1 (1)	49	S, C, Ca, N, H, G	2x10 ¹⁰	2
2	1 (1)	44	S, F, I	2x10 ¹¹	1
3	3 (3)	74 (66-76)	S, RT, T, P, C, V, D, Fl, Cl	2x10 ¹²	0/1/1
4	3 (3)	45 (38-60)	S, A, C, T, Ta, N, P, RT, E	2x10 ¹²	0/1/0
5	4 (3)	75 (65-75)	S, Ta, RT, I, D	2x10 ¹²	0/2/2
6	1 (1)	57	S, A, T, Ta	2x10 ¹² /divided doses	0
7	7 (5)	86 (76-92)	S, M, V, A, P, T, G, RT, F, I	2x10 ¹²	2/0/0/2/2
8	8 (5)	64 (62-91)	S, RT, IF, IT, P, T, F	2x10 ¹² repeated 2x week for 3 weeks	2/2/2/1/1 ^d

^aNo. of patients enrolled per cohort is indicated. The no. of patients completing at least one cycle of treatment is shown in parentheses.

^bPrior treatments: S, surgery; A, adriamycin; C, cyclophosphamide; Ca, capecitabine; Cl, chlorambucil; D, dacarbazine; E, etoposide; F, 5-fluorouracil; Fl, fluradabine; G, gemcitabine; H, herceptin; I, irinotecan; IF, IFN- γ ; IT, immunotherapy; M, methotrexate; N, navelbine; P, platinum; RT, radiotherapy; T, taxane; Tz, tamoxifen; V, vincristine. ^cAdverse events possibly or probably related to INGN 241 administration are indicated (\leq grade 3). ^dOne cohort 8 patient experienced a grade 3 SAE and withdrew from the study.

[5x10¹² viral particles (vp)/kg; equivalent to a human dose of 3x10¹⁴ vp], which consisted of decreases in body weight, mild liver toxicity, and transient decreases in platelet count (117,118). These results suggest that *mda-7/IL-24* may be optimally used therapeutically when combined with other treatment modalities (6,14,17-19) and supported initiation of clinical investigation. Thus far, however, clinical investigation has been limited and focused on sorting out mechanistic, pharmacodynamic, and pharmacokinetic issues using monotherapy. A single trial has been completed (28).

Safety. Twenty-two patients with refractory cancer and accessible tissue for surgical biopsy were entered into the first trial with *Ad.mda-7* (Table I) (28). This was a dose escalation study in which patients received 2x10¹⁰-2x10¹² vp delivered to the central region of the target tumor. Twenty-two patients completed 1 complete cycle (30 days). Treatment schedules ranged from a minimum of a single injection to two injections per week for 3 weeks. Of the evaluable patients there were 13 females and 9 males. Age ranged from 38 to 92 years, with a median age of 66 years. All patients had been heavily pretreated. Malignant melanoma (21.4%), squamous cell carcinoma of the head and neck (18%), breast carcinoma (14%), and colorectal carcinoma (7%) were the most frequent tumor types treated primarily due to accessibility of injectable tumor lesions. Adverse events were generally mild; transient fever being most common. There was one grade 3 serious adverse event (SAE) involving fatigue in a high dose-treated patient, which was thought possibly to be due to the study drug. The most common mild adverse event specifically related to injection of the study drug was included injection site pain. These effects were seen more consistently at higher doses and delivered volumes of *Ad.mda-7* and they generally resolved within 48 h after injection. In two patients in the highest dose schedule cohort (twice-weekly injections), marked skin erythema was noted surrounding the injected lesion within

24 h of the injection. This resolved over the next 96 h. A maximum tolerated dose was not attained in this study as the maximum monofactorable concentration was delivered. Overall, *Ad.mda-7* was well tolerated when administered in single or multidose regimens.

Clinical response. In the first six patient cohorts, injected tissue was excised within 24-96 h after injection. No clinical activity was observed within 24-96 h, although minor changes in morphology in the injected lesions were noted. Three patients receiving the highest single-dose therapy via intratumoral injection exhibited SD (stable disease), with 0, 0, and 23% reduction in tumor size (melanoma, colorectal carcinoma, and SCCHN, respectively). Off-study tumor measurements were taken 5-7 weeks from screening. Time from first injection to death for these patients was 347, 351, and 401 days, respectively.

Eight patients in cohort 8 received injections twice weekly for 3 weeks (in a 28-day cycle for 1 or more cycles); five patients completed at least one cycle of treatment. All patients in this cohort had failed multiple prior therapies (Table II). Two of five patients demonstrated a clinically-significant response to *Ad.mda-7* injections consisting of at least partial regression of the injected lesion. The most dramatic of these responses was in a 64-year-old female with widely metastatic melanoma (at study entry, she had >10 distinct lesion). Her initial site of treatment was a superclavicular node measuring 2x2 cm at baseline. No appreciable change was noted for the first five injections but by the sixth (and final) injection, a clear decrease in the size of the lesion was apparent and associated erythema was evident. Regression continued over the next 2 weeks until there was no clinical evidence of disease at that site (29) (Fig. 7). Subsequently, a second course of injections was begun on a lesion on the dorsum of her right hand. The baseline measurement was 1.8x2.3 cm and regression was evident by the fifth injection (84% reduction in lesion area). After

Table II. Responses in cohort 8 patients.

Patient	Gender/age	Diagnosis	Date diagnosed	Previous treatments ^a	No. of injections	Response ^b	Time to death ^c
81	M/71	Adenocarcinoma	1998	P, T, G	2 ^d	SD	nd
83	F/64	Melanoma	1994	S, RT, IT, IF	24 ^e	CR, PR, SD	>600
84	F/62	Melanoma	2000	S, RT, IT, IF	12	PR	309
85	M/64	Penile carcinoma	2002	S, RT, CDDP, T	6	PD	75
86	F/66	NSCLC	2001	RT, P, T	3 ^d	SD	180
87	M/62	SCCHN	1992	S, RT, CDDP, T, F, P	6	PD	185
88	M/91	Lip carcinoma	1982	S, RT	6	SD	181

^aPrior treatments: S, surgery; F, 5-fluorouracil; G, gemcitabine; IF, IFN- γ ; IT, immunotherapy; P, platinum; RT, radiotherapy; T, taxane.

^bSD, stable disease; CR, complete response; PR, partial response; PD, progressive disease. ^cTime in days from first injection of INGN 241 until death. ^dPatients 81 and 86 did not complete one full course of treatment (6 injections). ^ePatient received 12 injections on compassionate use protocol.

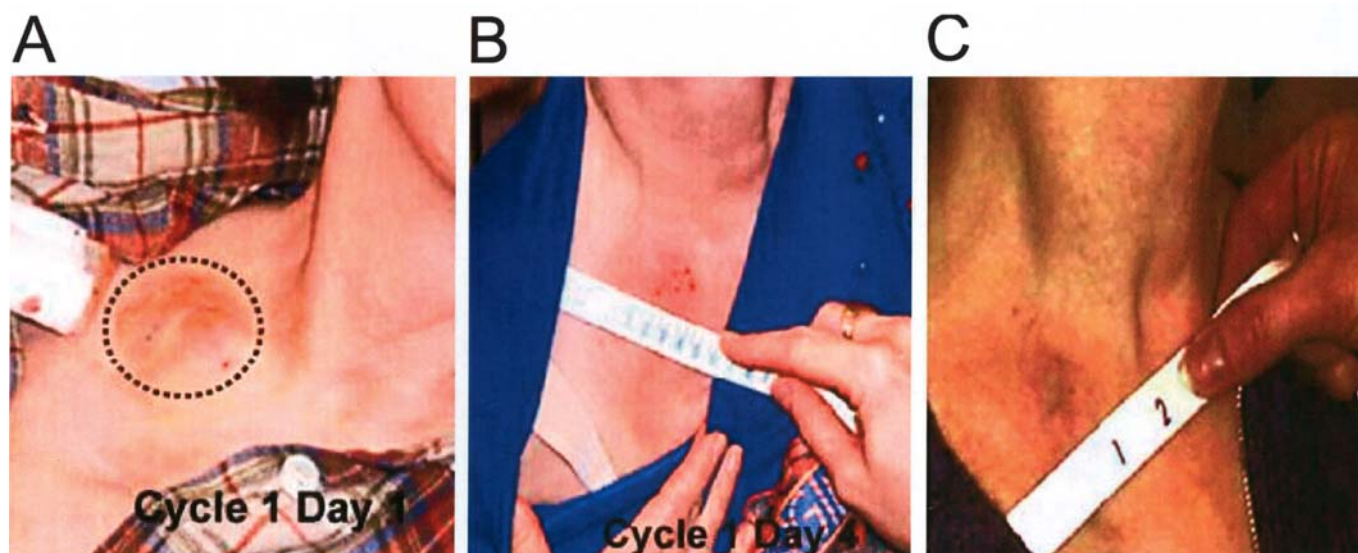


Figure 7. Objective clinical response to INGN 241 (Ad.mda-7) in cohort 8 patient with metastatic melanoma (patient 83). Injected lesion was on right clavicle (dashed circle in A). B, By day 4, region is inflamed. C, At the end of cycle 1 (day 30), lesion has completely regressed. This patient is still alive >600 days after initiating treatment.

completing six injections, the residual lesion was excised and on microscopic examination it was found to have marked inflammatory lymphoplasmacytic infiltrates throughout the residual nodule and surrounding tissue. There was also extensive coagulative necrosis in the tumor. A third lesion on the anterior right thigh was next treated in the same patient with two cycles of injections. Regression of this lesion was also observed after the first course of injections (baseline measurements 3.5x3.3 cm; 35% reduction), but a second set of injections produced no further response. Interestingly, several distant uninjected melanoma lesions also became erythematous during the course of injection of the target lesion, although clinical regression was not seen at these distant sites. This patient, a 64-year-old woman, survived 773 days after initiation of treatment with Ad.mda-7. An additional melanoma patient exhibited a partial response (33% decrease by RECIST criteria). Five of the eight cohort 8 patients

survived more than 2 years, four are still alive at 1643, 1550, 1267, and 1261 days.

We saw a less dramatic response in another patient with squamous cell carcinoma of the penis with multiple skin nodules in the groin and right hip area. Injection of Ad.mda-7 into one of the upper right hip lesions (2.5x3.0 cm at baseline) produced significant central necrosis with surrounding erythema by the fifth injection. However, the lesion continued to expand peripherally so that by completion of the first set of six injections, there was an indurated rim of erythematous tissue surrounding the central area and the total measurement of the lesion was 3.0x4.0 cm by the end of the 6th injection. A large central portion of the lesion (approximately two-thirds of the total area) remained blackened and necrotic at the end of cycle 1. Other, new lesions were rapidly appearing in the region, so the patient was removed from the study as PD, and he went on to other therapy. Three additional patients

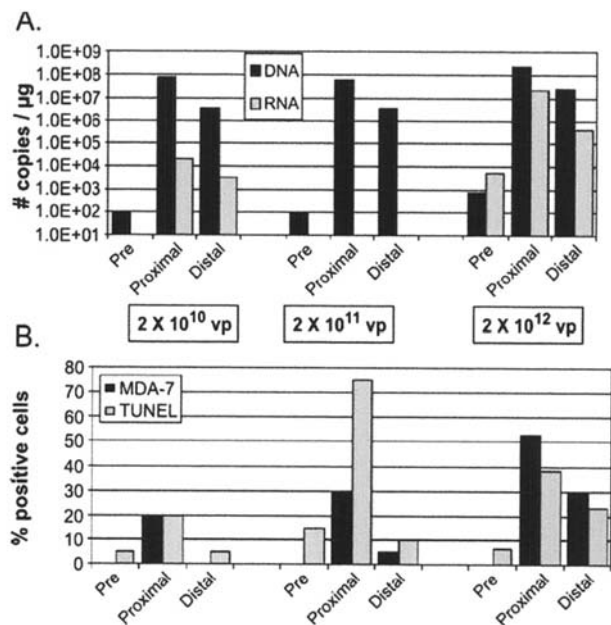


Figure 8. Pharmacodynamics of INGN 241 vector and expression. A, Dose-response of INGN 241 vector DNA and RNA. Tumor sections were obtained from preinjected (pre), proximal and distal sections from cohort 1 to 3 patients and vector-specific signals evaluated by quantitative DNA- and RT-PCR. RNA was not available from cohort 2 tumor. B, Dose-response of INGN 241 transgenic MDA-7/IL-24 expression and TUNEL reactivity. Tumor sections were obtained from preinjected (pre), proximal and distal sections from cohort 1 to 3 patients and vector-specific signals evaluated.

(with adenocarcinoma, NSCLC, and lip carcinoma) exhibited stable disease after Ad.*mda-7* injection and one patient with SCCHN exhibited disease progression (20% increase by RECIST). Of the nine lesions treated in cohort 8 patients, four demonstrated objective response (CR or PR by RECIST criteria).

Gene expression. Gene transfer and biologic effects in resected tumors were analyzed 1-4 days after injection. Quantitative studies of vector-specific DNA and RNA at the point of injection (center of lesion) and in serial sections to the periphery of each lesion were performed. To facilitate identification of the injection site, Isosulfan blue dye was admixed with vector prior to administration. Thus, at resection, the injection sites could be identified. All injected lesions showed high levels of vector-specific DNA signals in tumors (Fig. 8). Vector DNA, vector RNA, transgenic MDA-7/IL-24, and TUNEL signals across the dose range of 2×10^{10} - 2×10^{12} vp were compared (Fig. 8). Although the sample size was small, there was an indication of a dose-dependent increase in the number of Ad.*mda-7* vector DNA copies/ μ g DNA in the tumor [7×10^7 copies/ μ g in the proximal (central) sections of cohort 1 low dose compared to an average of 2.2×10^8 copies/ μ g] in the high dose cohort. A similar indication of dose response for vector RNA was also observed, with 2.0×10^4 vector RNA copies found in the central section of tumors injected with low dose INGN 241 (Ad.*mda-7*), whereas 2.3×10^7 copies/ μ g were observed in high-dose tumors (Fig. 8). Note that the distal regions of tumors uniformly showed lower vector signals than the proximal sections. Parallel sections for transgenic MDA-7/IL-24 protein and apoptosis induction

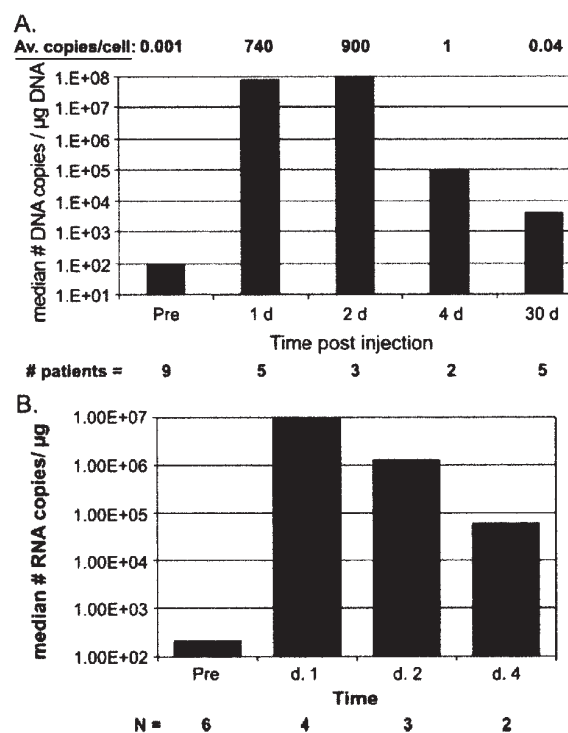


Figure 9. Intratumoral pharmacokinetics of vector DNA and mRNA. A, Decay of INGN 241 vector at injection site. The median number of DNA copies at each time point postinjection is shown; number of patients per sample is indicated below. The number of DNA copies/ μ g genomic DNA was converted to illustrate the average number of vector DNA copies per cell - shown above the graph. B, Decay of INGN 241 vector RNA at injection site. The median number of RNA copies/ μ g at each time point is shown; number of patients per sample is indicated below.

via TUNEL assay were also evaluated. All patients demonstrated undetectable MDA-7/IL-24 staining in the preinjected baseline samples, whereas MDA-7/IL-24 immunostaining was found in 20% of tumor cells from low dose, 30% of cells at intermediate dose, and 53% of cells from tumors injected with high dose of INGN 241. MDA-7/IL-24 staining in distal regions of tumors after injection with low dose vector was not observed, whereas up to 30% positive cells were found in mid- and high-dose tumors. Apoptosis induction showed higher trend with increased dose, except for the cohort 2, patient (colorectal carcinoma), who exhibited unusually high TUNEL reactivity of 75% after injection of Ad.*mda-7* (Fig. 8B).

Dynamics of vector spread and subsequent gene expression were also evaluated. The highest number of DNA vector copies were found at the area of injection (e.g., the center of injected lesions averaged greater than 1×10^8 vector DNA copies/ μ g) and signal decreased significantly ($p < 0.02$) with distance from the injection site, vector DNA levels had fallen by almost 90%. Low levels of vector DNA were detected up to 3-cm from the injection site; however, only 3 tumors showed DNA signals greater than 1×10^6 copies/ μ g at 1-cm from the injection site. Vector RNA levels were also quantitated and they showed a pattern of distribution similar to that of the vector DNA. Vector RNA was also distributed distally from the injection site and, similar to vector DNA, vector RNA signals were significantly reduced ($p < 0.05$) with distance from the injection site. Both vector DNA and RNA signals showed exponential decay with distance from the injection

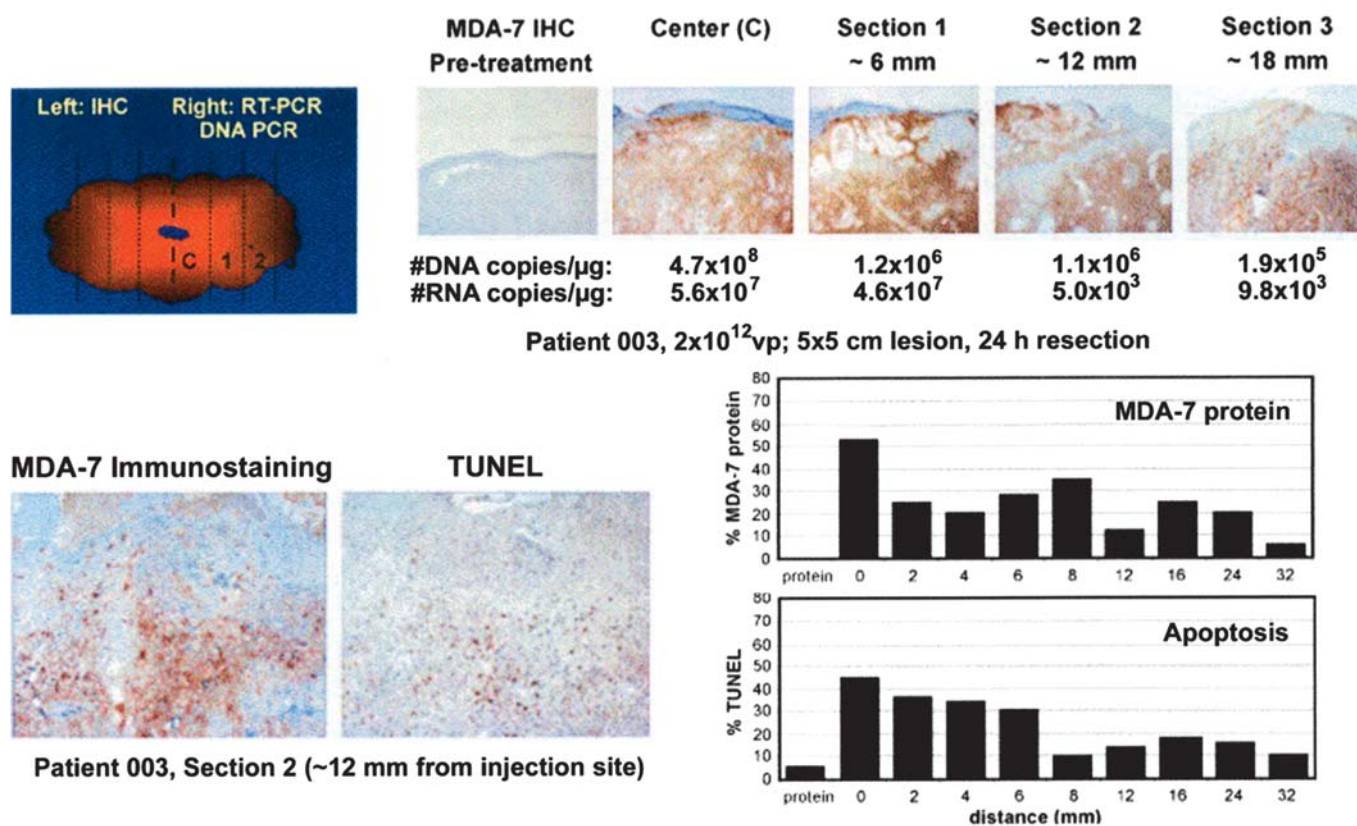


Figure 10. *Mda-7/IL-24* transgene expression correlates with distribution of vector throughout the tumor. One-half of the tumor was analyzed for MDA-7/IL-24 protein expression and the other half for vector-specific DNA and RNA levels. The number of DNA copies/ μ g genomic DNA and number of RNA copies/ μ g total RNA is shown for each tumor section. TUNEL reactivity was 50% in the central section and 30% at the periphery section 3.

site. Regression analyses indicated very strong correlations between vector DNA and RNA signals and distance from injection site (correlation coefficient = 0.82-0.9).

The kinetics of vector DNA and transgenic mRNA were evaluated for patients who received a single dose of INGN 241 (Ad.*mda-7*). Vector DNA reached maximum levels at the point of injection 24-48 h after injection. If one assumes these signals are cell-associated, then median signals approximated 1000 vector DNA copies per cell, a value 6 logs above pre-injection controls (Fig. 9A). Vector-specific mRNA exhibited a distribution and kinetic profile similar to that of vector DNA (Fig. 9B). Median vector-specified RNA signals were more than 4 logs greater in the center of injection lesions compared to uninjected control samples (Figs. 8A and 9B). Vector-specific RNA signals decreased by more than 2 logs by day 4 (Fig. 9B). Samples were not available for day 30 analysis.

Transgenic gene expression. Tumors were excised and serially-sectioned, then evaluated for transgenic MDA-7/IL-24 protein expression and apoptosis. Pretreatment samples were uniformly negative for MDA-7/IL-24 protein expression and most were negative for TUNEL reactivity (average TUNEL signal was <6% in cohorts 1-5). After Ad.*mda-7* injection, all tumors demonstrated substantial MDA-7/IL-24 immunostaining that ranged from 20 to 90% positive tumor cells in the center of the lesion. Transgenic MDA-7/IL-24 was detected beyond the injection site: five of eight (62%) tumors had detectable MDA-7/IL-24 at 1-cm from the single injection site, and

MDA-7/IL-24 expression was detected more than 3-cm from the injection site. Up to 25% of MDA-7/IL-24 stained cells were detected 16-mm from the point of injection. Reproducibly, within each tumor, MDA-7/IL-24 immunostaining was reduced with distance from the injection site. With the exception of patient 2, $\geq 90\%$ of MDA-7/IL-24 staining cells in all biopsies exhibited malignant histological features. The remaining MDA-7/IL-24-positive cells comprised infiltrating lymphocytes and/or histocytic/reticuloendothelial cells.

Apoptosis staining varied, with up to 80% of tumor cells at the center of the lesion demonstrating TUNEL reactivity. Apoptosis declined with distance from the injection site; five of seven (71%) tumors exhibited TUNEL reactivity beyond 1-cm from the injection site. Both MDA-7/IL-24 and TUNEL staining demonstrated significant ($p < 0.02$) linear decay with distance from the injection site. Regions of tumor exhibiting TUNEL staining strongly corresponded to those regions having MDA-7/IL-24 protein expression; those samples with distal MDA-7/IL-24 staining also showed distal TUNEL reactivity. Both protein expression and apoptosis reactivity reached maximum levels by 4-days postinjection but had returned to baseline, preinjection levels by day 30 [(28) and data not shown].

A representative example is shown in Fig. 10, which illustrates the high level of MDA-7/IL-24 immunostaining after Ad.*mda-7* injection and the decay in protein, DNA, and RNA signals with distance from the injection site. This patient with a 5x5-cm SCCHN lesion was injected with 2×10^{12} vp Ad.*mda-7* and resected at 24 h. No MDA-7/IL-24 signal was

observed in the pretreatment lesion, whereas the injection site (center section) exhibited up to 75% of tumor cells staining for MDA-7/IL-24. Staining intensity decreased with distance, but the distal lesion (18-mm from injection site) still showed regions of strong MDA-7/IL-24 staining (Fig. 10). Parallel sections were evaluated for apoptosis. The central section showed 50% TUNEL reactivity, whereas the distal section showed 30% staining. The levels of vector-specific DNA and RNA showed strong signals at the injection site, which decreased markedly (more than 100-fold) at the periphery (Fig. 10). Overall, vector-specific DNA and RNA, MDA-7/IL-24 protein levels, and TUNEL reactivity demonstrated similar dose response and kinetic and radial concentration gradients, although MDA-7/IL-24 and TUNEL signals persisted longer and showed enhanced distribution.

Conclusions from the phase I clinical trial. Direct injection of an adenovirus containing the *mda-7/IL-24* cDNA (INGN 241) into a variety of solid tumors was generally well tolerated, with injection site pain and erythema being noted locally in some patients. One patient experienced a grade 3 SAE of fatigue and was discontinued from the study.

Analysis of serial sections of injected lesions demonstrated that *mda-7/IL-24* DNA and RNA were detectable in 100% of the injected lesions, with the highest concentrations found at the site of injection, as expected. However, MDA-7/IL-24 protein was detectable in the periphery of injected lesions (greater than 3-cm from injection site), beyond the area of DNA spread, suggesting that the MDA-7/IL-24 protein can diffuse from transduced cells, which supports the *in vitro* 'bystander antitumor' studies (18,35,44).

Apoptosis, as measured by TUNEL assay, was significant in the injected lesions and also correlated geographically with MDA-7/IL-24 protein expression. Both MDA-7/IL-24 transgenic protein expression and apoptosis decayed with similar intratumoral dynamics. One hundred percent of injected tumors evaluated in the phase I clinical study demonstrated transgenic MDA-7/IL-24 expression and elevated apoptosis induction compared to untreated control tumors. In contrast, adenoviral-mediated gene transfer of p53 by injection into non-small cell lung cancer tumors resulted in vector detection by DNA PCR in 86% of patients' tumors and increased apoptosis in 46% of biopsy specimens (119). Indeed, as most previous gene transfer studies have not been structured to assess the geographic extent of vector distribution and biological effect, it is difficult to compare INGN 241's effects directly with other studies. One report has indicated that administration of Ad.p53 to glioma is hampered by the limited spread (<5 mm) of transduced cells (120). Both INGN 241 DNA and transgenic MDA-7/IL-24 protein distribute to a greater extent and demonstrate strong correlations with distance from injection site ($p < 0.02$). Nucleic acid signals decay by 50% at 4-6-mm from the injection site whereas MDA-7/IL-24 protein and apoptosis signals decay by 50% at 18-20-mm. Therefore, either INGN 241 (Ad.*mda-7*) is more potent than other therapeutic constructs at inducing apoptosis in human tumors or the study design allowed us to capture signals that were lost in other studies. The levels of apoptosis induction observed in this study are substantially higher (average = 45%) than reported with other anticancer drugs. Apoptosis induction correlates

with loss of Ki-67 staining (27). It is noteworthy that seven different tumor types were treated in cohorts 1-6 and all showed high levels of MDA-7/IL-24 expression and subsequent apoptosis induction. As discussed above, all of these patients had been heavily pretreated and failed multiple treatment regimens. It would be predicted that this group of chemo- and radio-resistant tumors would have acquired resistance to apoptosis. The fact that MDA-7/IL-24 induces apoptosis in such a spectrum of advanced tumor types mirrors preclinical studies and suggests that INGN 241 (Ad.*mda-7*) may have broad utility.

As discussed in other parts of this review, in addition to apoptosis induction of tumor cells and immunostimulation, additional 'bystander antitumor' activities have been reported for MDA-7/IL-24. Ad.*mda-7* was reported to inhibit endothelial differentiation *in vitro*, implying antiangiogenic activity (24). Further studies demonstrated antiangiogenic activity of Ad.*mda-7* *in vivo* (23), in that MDA-7/IL-24 expression significantly repressed angiogenic mediators, including VEGF, basic FGF, and IL-8. Studies using purified MDA-7/IL-24 demonstrated that it functions via the IL-22R1 on endothelial cells (92), and MDA-7/IL-24 was 50-fold more active than endostatin or angiostatin. When Ad.*mda-7* was combined with irradiation, synergistic inhibition of tumor growth was observed, with significant reduction in microvessel density and pronounced apoptotic response in the tumor (23). Two recent preclinical studies have provided support for intriguing observations about the role of MDA-7/IL-24 in melanoma disease progression and metastasis. Initial studies found that MDA-7/IL-24 mRNA and protein were expressed in normal human melanocytes and benign nevi, but expression was lost in metastatic melanoma (2,11). Subsequent studies evaluated larger patient groups and concluded that MDA-7/IL-24 protein expression is progressively lost as melanoma tumors invade and become more metastatic and aggressive (10). These authors speculated that MDA-7/IL-24 must play a role in maintenance of normal physiology of melanocytes and that inhibition of MDA-7/IL-24 results in transformation and progression from a local, non-metastatic primary tumor to a highly metastatic phenotype (2,10,11). Additional studies have shown that Ad.*mda-7* can inhibit cell migration and invasion of lung and breast tumor cells by down-regulating the PI3K pathway and inhibiting production of FAK and matrix metalloprotease (63,94). Tumor cells expressing MDA-7/IL-24 demonstrated significantly reduced lung metastases compared to control cells (76,94). It is clear that tumor cells can develop resistance to cytotoxic therapies, and reports are now emerging of acquisition of resistance to pathway-specific molecularly targeted therapeutics. The antitumor effects mediated by MDA-7/IL-24 encompass a variety of signaling pathways, and it is anticipated that redundant proapoptotic signals are activated (14). The ability of *mda-7/IL-24* to enhance the therapeutic activity of radiation, monoclonal antibody therapy and chemotherapy provide additional avenues worth exploring as a means of enhancing the therapeutic index of this novel cytokine (5,6,14,16,18,26,35,37,38,40,42,58,61,62,65,68,118,121).

Transient increases in circulating cytokines such as IL-6, IL-10, and TNF- α in response to MDA-7/IL-24 were observed. Significantly higher elevations of IL-6 and TNF- α were

observed in patients receiving repeat treatment who demonstrated evidence of activity related to Ad.*mda-7* (27). The majority of patients also showed a marked increase in CD3⁺ and CD8⁺ T cells at day 15 following injection, suggesting that Ad.*mda-7* may be associated with a TH1 response. Furthermore, MDA-7/IL-24 has been shown to inhibit G₂/M cell cycling (12,24,50). The effect of MDA-7/IL-24 on tumor suppressor genes, such as β -catenin and iNOS, was also evaluated (27). β -catenin expression was reduced in six of nine patients and iNOS expression was reduced in four of nine patients tested.

In summary, Ad.*mda-7* can induce a wide variety of effects associated with anticancer activity following intratumoral injection. Clinically significant responses were primarily seen with repeat injection. Future studies, therefore, will need to concentrate on repeat dosing of Ad.*mda-7*, particularly in malignant melanoma, in which the greatest clinical activity was seen in the first phase I study (29). Other future directions include developing systemic administration of Ad.*mda-7*, given its widespread tumor selectivity, exploration of its immunopotentiating effects, and its use in combinatorial strategies.

9. Future applications for improving the therapeutic index of *mda-7/IL-24*

In a remarkably short time frame, *mda-7/IL-24* has gone from a laboratory discovery into the clinic. The results achieved to date using this gene are very encouraging and suggest that *mda-7/IL-24* will provide significant benefit as a therapeutic for cancer. Although the results of the phase I clinical trial are provocative, it is clear that approaches to enhance the therapeutic benefit of *mda-7/IL-24* are necessary to maximize its effectiveness in patients. Several approaches to achieve this objective were recently highlighted (14). One strategy, which has already proven efficacious, is the application of conditionally replication competent bipartite adenoviruses that both replicate and express *mda-7/IL-24* in tumor cells (73,122, 123). These novel viruses have shown enhanced effectiveness in athymic nude mice containing human breast (73), colorectal (122) or therapy-resistant prostate (123) tumors in comparison with non-replicating adenoviruses expressing *mda-7/IL-24*. Augmentation of the antitumor properties of *mda-7/IL-24* has also been achieved by using this gene in combination with radiation, chemotherapy or monoclonal antibody therapy (reviewed in refs. 18,19). Combining *mda-7/IL-24* with agents that promote ROS production has been shown to augment therapeutic action of this gene in the context of prostate and pancreatic carcinomas (42,43). In this context, agents that can evoke this change in cancer cells may provide useful reagents to enhance therapeutic activity of *mda-7/IL-24*. Additionally, approaches that facilitate delivery of *mda-7/IL-24* to the tumor microenvironment and tumor cells, including the use of stem cells and dendritic cells, offer promise for increasing therapeutic index of this novel cytokine (14). Although requiring further development and testing, modifying normal cells to produce and secrete MDA-7/IL-24 protein could in principle permit an even further expansion of clinical utility for this interesting molecule (14,35). Additionally, using agents that can induce MDA-7/IL-24 expression in cancer cells, thereby resulting in apoptosis, represents a unique

therapy for cancer (124). Clearly, these and other approaches will require further study to define ways of more effectively using *mda-7/IL-24* as a cancer gene therapeutic. We remain cautiously optimistic relative to the use and effectiveness of this novel cytokine for the therapy of diverse human malignancies.

Acknowledgements

Support for the studies provided in this review have come from the NIH/NCI, Army DOD, and several private foundations. Support for P.B.F. and members of his laboratory (I.V.L., L.E., Z.-Z.S., P.G., M.S., D.S., M.R.S., T.M.M., P.B., S.-G.L., N.V., E.S.P., L.C., H.B.) was provided by NIH grants R01 CA35675, R01 CA97318, R01 CA108520, R01 CA98712 and P01 CA104177; Army DOD Grants DAMD 17-03-1-0290 and DAMD W81XWH-04-1-0433; the Lustgarten Foundation for Pancreatic Cancer Research; the Samuel Waxman Cancer Research Foundation and the Chernow Endowment. R.X. and D.W. were supported by the Army DOD Post-doctoral Fellowship PC 050545. Support for Z.-J.L. was provided by funds from New York State Department of Health (New York State Breast Cancer Research and Education Fund), contract #C020906. R.L., A.L.D. and P.B.F. were supported by NIH/NCI grants 5 U54 CA101388 and 5 U54 CA101598. Support for R.R., S.I. and S.C. was provided by NIH research grants CA102716, CA06294, CA89778, CA16672 and Sponsored Research Agreement with Introgen Therapeutics, Inc., Houston, TX. P.M., I.P.D. and D.T.C. were supported by P01 CA104177. Support for A.Y., S.G. and P.D. was provided by PHS grants R01 CA88906, P01 CA72955, R01 CA108520, R01 DK52825, and CA104177. P.B.F. is the Michael and Stella Chernow Urological Cancer Research Scientist and a SWCRF Investigator.

References

- Jiang H and Fisher PB: Use of a sensitive and efficient subtraction hybridization protocol for the identification of genes differentially regulated during the induction of differentiation in human melanoma cells. *Mol Cell Differ* 1: 285-299, 1993.
- Jiang H, Lin JJ, Su ZZ, Goldstein NI and Fisher PB: Subtraction hybridization identifies a novel melanoma differentiation associated gene, *mda-7*, modulated during human melanoma differentiation, growth and progression. *Oncogene* 11: 2477-2486, 1995.
- Pestka S, Krause CD, Sarkar D, Walter MR, Shi Y and Fisher PB: Interleukin-10 and related cytokines and receptors. *Annu Rev Immunol* 22: 929-979, 2004.
- Sauane M, Gopalkrishnan RV, Sarkar D, Su ZZ, Lebedeva IV, Dent P, Pestka S and Fisher PB: MDA-7/IL-24: novel cancer growth suppressing and apoptosis inducing cytokine. *Cytokine Growth Factor Rev* 14: 35-51, 2003.
- Huang EY, Madireddi MT, Gopalkrishnan RV, Leszczyniecka M, Su Z, Lebedeva IV, Kang D, Jiang H, Lin JJ, Alexandre D, Chen Y, Vozhilla N, Mei MX, Christiansen KA, Sivo F, Goldstein NI, Mhashilkar AB, Chada S, Huberman E, Pestka S and Fisher PB: Genomic structure, chromosomal localization and expression profile of a novel melanoma differentiation associated (*mda-7*) gene with cancer specific growth suppressing and apoptosis inducing properties. *Oncogene* 20: 7051-7063, 2001.
- Fisher PB, Gopalkrishnan RV, Chada S, Ramesh R, Grimm EA, Rosenfeld MR, Curiel DT and Dent P: *Mda-7/IL-24*, a novel cancer selective apoptosis inducing cytokine gene: from the laboratory into the clinic. *Cancer Biol Ther* 2: S23-S37, 2003.

7. Caudell EG, Mumm JB, Poindexter N, Ekmekcioglu S, Mhashilkar AM, Yang XH, Retter MW, Hill P, Chada S and Grimm EA: The protein product of the tumor suppressor gene, melanoma differentiation-associated gene 7, exhibits immunostimulatory activity and is designated IL-24. *J Immunol* 168: 6041-6046, 2002.
8. Sarkar D, Su ZZ, Lebedeva IV, Sauane M, Gopalkrishnan RV, Dent P and Fisher PB: *Mda-7* (IL-24): signaling and functional roles. *Biotechniques (Suppl)*: 30-39, 2002.
9. Wolk K, Kunz S, Asadullah K and Sabat R: Cutting edge: immune cells as sources and targets of the IL-10 family members? *J Immunol* 168: 5397-5402, 2002.
10. Ellerhorst JA, Prieto VG, Ekmekcioglu S, Broemeling L, Yekell S, Chada S and Grimm EA: Loss of MDA-7 expression with progression of melanoma. *J Clin Oncol* 20: 1069-1074, 2002.
11. Ekmekcioglu S, Ellerhorst J, Mhashilkar AM, Sahin AA, Read CM, Prieto VG, Chada S and Grimm EA: Down-regulated melanoma differentiation associated gene (*mda-7*) expression in human melanomas. *Int J Cancer* 94: 54-59, 2001.
12. Lebedeva IV, Su ZZ, Chang Y, Kitada S, Reed JC and Fisher PB: The cancer growth suppressing gene *mda-7* induces apoptosis selectively in human melanoma cells. *Oncogene* 21: 708-718, 2002.
13. Jiang H, Su ZZ, Lin JJ, Goldstein NI, Young CS and Fisher PB: The melanoma differentiation associated gene *mda-7* suppresses cancer cell growth. *Proc Natl Acad Sci USA* 93: 9160-9165, 1996.
14. Fisher PB: Is *mda-7/IL-24* a 'magic bullet' for cancer? *Cancer Res* 65: 10128-10138, 2005.
15. Gopalkrishnan RV, Sauane M and Fisher PB: Cytokine and tumor cell apoptosis inducing activity of *mda-7/IL-24*. *Int Immunopharmacol* 4: 635-647, 2004.
16. Chada S, Sutton RB, Ekmekcioglu S, Ellerhorst J, Mumm JB, Leitner WW, Yang HY, Sahin AA, Hunt KK, Fuson KL, Poindexter N, Roth JA, Ramesh R, Grimm EA and Mhashilkar AM: MDA-7/IL-24 is a unique cytokine-tumor suppressor in the IL-10 family. *Int Immunopharmacol* 4: 649-667, 2004.
17. Lebedeva IV, Sauane M, Gopalkrishnan RV, Sarkar D, Su ZZ, Gupta P, Nemunaitis J, Cunningham C, Yacoub A, Dent P and Fisher PB: *Mda-7/IL-24*: exploiting cancer's Achilles' heel. *Mol Ther* 11: 4-18, 2005.
18. Gupta P, Su ZZ, Lebedeva IV, Sarkar D, Sauane M, Emdad L, Bachelor MA, Grant S, Curiel DT, Dent P and Fisher PB: *Mda-7/IL-24*: multifunctional cancer-specific apoptosis-inducing cytokine. *Pharmacol Ther* 111: 596-628, 2006.
19. Sarkar D, Dent P and Fisher PB: Melanoma differentiation associated gene-7 (*mda-7*)/interleukin-24 (IL-24), *mda-7/IL-24*: current perspectives on a unique member of the IL-10 family of cytokines. Anti-inflammatory and anti-allergy agents in medicinal chemistry 5: 259-274, 2006.
20. Inoue S, Shanker M, Miyahara R, Gopalan B, Patel S, Oida Y, Branch CD, Munshi A, Meyn RE, Aandreeff M, Tanaka F, Mhashilkar AM, Chada S and Ramesh R: MDA-7/IL-24-based cancer gene therapy: translation from the laboratory to the clinic. *Curr Gene Ther* 6: 73-91, 2006.
21. Lebedeva IV, Sarkar D, Su ZZ, Kitada S, Dent P, Stein CA, Reed JC and Fisher PB: Bcl-2 and Bcl-x(L) differentially protect human prostate cancer cells from induction of apoptosis by melanoma differentiation associated gene-7, *mda-7/IL-24*. *Oncogene* 22: 8758-8773, 2003.
22. Mhashilkar AM, Schrock RD, Hindi M, Liao J, Sieger K, Kourouma F, Zou-Yang XH, Onishi E, Takh O, Vedvick TS, Fanger G, Stewart L, Watson GJ, Snary D, Fisher PB, Saeki T, Roth JA, Ramesh R and Chada S: Melanoma differentiation associated gene-7 (*mda-7*): a novel anti-tumor gene for cancer gene therapy. *Mol Med* 7: 271-282, 2001.
23. Nishikawa T, Ramesh R, Munshi A, Chada S and Meyn RE: Adenovirus-mediated *mda-7* (IL-24) gene therapy suppresses angiogenesis and sensitizes NSCLC xenograft tumors to radiation. *Mol Ther* 9: 818-828, 2004.
24. Saeki T, Mhashilkar A, Swanson X, Zou-Yang XH, Sieger K, Kawabe S, Branch CD, Zumstein L, Meyn RE, Roth JA, Chada S and Ramesh R: Inhibition of human lung cancer growth following adenovirus-mediated *mda-7* gene expression *in vivo*. *Oncogene* 21: 4558-4566, 2002.
25. Su ZZ, Madireddi MT, Lin JJ, Young CS, Kitada S, Reed JC, Goldstein NI and Fisher PB: The cancer growth suppressor gene *mda-7* selectively induces apoptosis in human breast cancer cells and inhibits tumor growth in nude mice. *Proc Natl Acad Sci USA* 95: 14400-14405, 1998.
26. Yacoub A, Mitchell C, Lister A, Lebedeva IV, Sarkar D, Su ZZ, Sigmon C, McKinstry R, Ramakrishnan V, Qiao L, Broaddus WC, Gopalkrishnan RV, Grant S, Fisher PB and Dent P: Melanoma differentiation-associated 7 (interleukin 24) inhibits growth and enhances radiosensitivity of glioma cells *in vitro* and *in vivo*. *Clin Cancer Res* 9: 3272-3281, 2003.
27. Tong AW, Nemunaitis J, Su D, Zhang Y, Cunningham C, Senzer N, Netto G, Rich D, Mhashilkar A, Parker K, Coffee K, Ramesh R, Ekmekcioglu S, Grimm EA, van Wart Hood J, Merritt J and Chada S: Intratumoral injection of INGN 241, a non-replicating adenovector expressing the melanoma-differentiation associated gene-7 (*mda-7/IL24*): biologic outcome in advanced cancer patients. *Mol Ther* 11: 160-172, 2005.
28. Cunningham CC, Chada S, Merritt JA, Tong A, Senzer N, Zhang Y, Mhashilkar A, Parker K, Vukelja S, Richards D, Hood J, Coffee K and Nemunaitis J: Clinical and local biological effects of an intratumoral injection of *mda-7* (IL24; INGN 241) in patients with advanced carcinoma: a phase I study. *Mol Ther* 11: 149-159, 2005.
29. Fisher PB, Sarkar D, Lebedeva IV, Emdad L, Gupta P, Sauane M, Su Z-Z, Grant S, Dent P, Curiel DT, Senzer N and Nemunaitis J: Melanoma differentiation associated gene-7/interleukin-24 (*mda-7/IL-24*): novel gene therapeutic for metastatic melanoma. *Toxicol Appl Pharmacol* 2006, Nov 23 [epub ahead of print].
30. Parrish-Novak J, Xu W, Brender T, Yao L, Jones C, West J, Brandt C, Jelinek L, Madden K, McKernan PA, Foster DC, Jaspers S and Chandrasekher YA: Interleukins 19, 20, and 24 signal through two distinct receptor complexes. Differences in receptor-ligand interactions mediate unique biological functions. *J Biol Chem* 277: 47517-47523, 2002.
31. Sauane M, Gopalkrishnan RV, Lebedeva I, Mei MX, Sarkar D, Su ZZ, Kang DC, Dent P, Pestka S and Fisher PB: *Mda-7/IL-24* induces apoptosis of diverse cancer cell lines through JAK/STAT-independent pathways. *J Cell Physiol* 196: 334-345, 2003.
32. Wang M, Tan Z, Zhang R, Kottenko SV and Liang P: Interleukin 24 (MDA-7/MOB-5) signals through two heterodimeric receptors, IL-22R1/IL-20R2 and IL-20R1/IL-20R2. *J Biol Chem* 277: 7341-7347, 2002.
33. Dumoutier L, Leemans C, Lejeune D, Kottenko SV and Renauld JC: Cutting edge: STAT activation by IL-19, IL-20 and *mda-7* through IL-20 receptor complexes of two types. *J Immunol* 167: 3545-3549, 2001.
34. Chada S, Mhashilkar AM, Ramesh R, Mumm JB, Sutton RB, Bocangel D, Zheng M, Grimm EA and Ekmekcioglu S: Bystander activity of Ad-*mda7*: human MDA-7 protein kills melanoma cells via an IL-20 receptor-dependent but STAT3-independent mechanism. *Mol Ther* 10: 1085-1095, 2004.
35. Su Z, Emdad L, Sauane M, Lebedeva IV, Sarkar D, Gupta P, James CD, Randolph A, Valerie K, Walter MR, Dent P and Fisher PB: Unique aspects of *mda-7/IL-24* antitumor bystander activity: establishing a role for secretion of MDA-7/IL-24 protein by normal cells. *Oncogene* 24: 7552-7566, 2005.
36. Madireddi MT, Su ZZ, Young CS, Goldstein NI and Fisher PB: *Mda-7*, a novel melanoma differentiation associated gene with promise for cancer gene therapy. *Adv Exp Med Biol* 465: 239-261, 2000.
37. Su ZZ, Lebedeva IV, Sarkar D, Gopalkrishnan RV, Sauane M, Sigmon C, Yacoub A, Valerie K, Dent P and Fisher PB: Melanoma differentiation associated gene-7, *mda-7/IL-24*, selectively induces growth suppression, apoptosis and radiosensitization in malignant gliomas in a p53-independent manner. *Oncogene* 22: 1164-1180, 2003.
38. Yacoub A, Mitchell C, Hong Y, Gopalkrishnan RV, Su ZZ, Gupta P, Sauane M, Lebedeva IV, Curiel DT, Mahareshti PJ, Rosenfeld MR, Broaddus WC, James CD, Grant S, Fisher PB and Dent P: MDA-7 regulates cell growth and radiosensitivity *in vitro* of primary (non-established) human glioma cells. *Cancer Biol Ther* 3: 739-751, 2004.
39. Yacoub A, Mitchell C, Brannon J, Rosenberg E, Qiao L, McKinstry R, Linehan WM, Su ZS, Sarkar D, Lebedeva IV, Valerie K, Gopalkrishnan RV, Grant S, Fisher PB and Dent P: MDA-7 (interleukin-24) inhibits the proliferation of renal carcinoma cells and interacts with free radicals to promote cell death and loss of reproductive capacity. *Mol Cancer Ther* 2: 623-632, 2003.
40. Su ZZ, Lebedeva IV, Sarkar D, Emdad L, Gupta P, Kitada S, Dent P, Reed JC and Fisher PB: Ionizing radiation enhances therapeutic activity of *mda-7/IL-24*: overcoming radiation- and *mda-7/IL-24*-resistance in prostate cancer cells overexpressing the antiapoptotic proteins bcl-x_L or bcl-2. *Oncogene* 25: 2339-2348, 2006.

41. Saito Y, Miyahara R, Gopalan B, Litvak A, Inoue S, Shanker M, Branch CD, Mhashilkar AM, Roth JA, Chada S and Ramesh R: Selective induction of cell cycle arrest and apoptosis in human prostate cancer cells through adenoviral transfer of the melanoma differentiation-associated-7 (*mda-7*)/interleukin-24 (IL-24) gene. *Cancer Gene Ther* 12: 238-247, 2005.
42. Lebedeva IV, Su ZZ, Sarkar D, Gopalkrishnan RV, Waxman S, Yacoub A, Dent P and Fisher PB: Induction of reactive oxygen species renders mutant and wild-type *K-ras* pancreatic carcinoma cells susceptible to Ad.*mda-7*-induced apoptosis. *Oncogene* 24: 585-596, 2005.
43. Lebedeva IV, Su ZZ, Sarkar D, Kitada S, Dent P, Waxman S, Reed JC and Fisher PB: Melanoma differentiation associated gene-7, *mda-7*/interleukin-24, induces apoptosis in prostate cancer cells by promoting mitochondrial dysfunction and inducing reactive oxygen species. *Cancer Res* 63: 8138-8144, 2003.
44. Su Z, Lebedeva IV, Gopalkrishnan RV, Goldstein NI, Stein CA, Reed JC, Dent P and Fisher PB: A combinatorial approach for selectively inducing programmed cell death in human pancreatic cancer cells. *Proc Natl Acad Sci USA* 98: 10332-10337, 2001.
45. Sauane M, Lebedeva IV, Su ZZ, Choo HT, Randolph A, Valerie K, Dent P, Gopalkrishnan RV and Fisher PB: Melanoma differentiation associated gene-7/interleukin-24 promotes tumor cell-specific apoptosis through both secretory and nonsecretory pathways. *Cancer Res* 64: 2988-2993, 2004.
46. Gupta P, Walter MR, Su ZZ, Lebedeva IV, Emdad L, Randolph A, Valerie K, Sarkar D and Fisher PB: BiP/GRP78 is an intracellular target for MDA-7/IL-24 induction of cancer-specific apoptosis. *Cancer Res* 66: 8182-8191, 2006.
47. Sauane M, Gupta P, Lebedeva IV, Su Z-Z, Sarkar D, Randolph A, Valerie K, Gopalkrishnan R and Fisher PB: N-glycosylation of MDA-7/IL-24 is dispensable for tumor cell-specific apoptosis and bystander anti-tumor activity. *Cancer Res* 66: 11869-11877, 2006.
48. Berridge MJ: The endoplasmic reticulum: a multifunctional signaling organelle. *Cell Calcium* 32: 235-249, 2002.
49. Berridge MJ, Lipp P and Bootman MD: The versatility and universality of calcium signalling. *Nat Rev Mol Cell Biol* 1: 11-21, 2000.
50. Sarkar D, Su ZZ, Lebedeva IV, Sauane M, Gopalkrishnan RV, Valerie K, Dent P and Fisher PB: *Mda-7* (IL-24) mediates selective apoptosis in human melanoma cells by inducing the coordinated overexpression of the GADD family of genes by means of p38 MAPK. *Proc Natl Acad Sci USA* 99: 10054-10059, 2002.
51. Lingappa VR and Blobel G: Early events in the biosynthesis of secretory and membrane proteins: the signal hypothesis. *Recent Prog Horm Res* 36: 451-475, 1980.
52. Sieger KA, Mhashilkar AM, Stewart A, Sutton RB, Strube RW, Chen SY, Pataer A, Swisher SG, Grimm EA, Ramesh R and Chada S: The tumor suppressor activity of MDA-7/IL-24 is mediated by intracellular protein expression in NSCLC cells. *Mol Ther* 9: 355-367, 2004.
53. Sauane M, Gopalkrishnan RV, Choo HT, Gupta P, Lebedeva IV, Yacoub A, Dent P and Fisher PB: Mechanistic aspects of *mda-7*/IL-24 cancer cell selectivity analysed via a bacterial fusion protein. *Oncogene* 23: 7679-7690, 2004.
54. Zhan Q, Lord KA, Alamo I Jr, Hollander MC, Carrier F, Ron D, Kohn KW, Hoffman B, Liebermann DA and Fornace AJ Jr: The *gadd* and *MyD* genes define a novel set of mammalian genes encoding acidic proteins that synergistically suppress cell growth. *Mol Cell Biol* 14: 2361-2371, 1994.
55. Sainz-Perez A, Gary-Gouy H, Portier A, Davi F, Merle-Beral H, Galanaud P and Dalloul A: High *Mda-7* expression promotes malignant cell survival and p38 MAP kinase activation in chronic lymphocytic leukemia. *Leukemia* 20: 498-504, 2006.
56. Pataer A, Vorburger SA, Barber GN, Chada S, Mhashilkar AM, Zou-Yang H, Stewart AL, Balachandran S, Roth JA, Hunt KK and Swisher SG: Adenoviral transfer of the melanoma differentiation-associated gene 7 (*mda7*) induces apoptosis of lung cancer cells via up-regulation of the double-stranded RNA-dependent protein kinase (PKR). *Cancer Res* 62: 2239-2243, 2002.
57. Emdad L, Lebedeva IV, Su Z-Z, Gupta P, Sarkar D, Settleman J and Fisher PB: Combinatorial treatment of non-small-cell lung cancers with gefitinib and Ad.*mda-7* enhances apoptosis-induction and reverses resistance to a single therapy. *J Cell Physiol* 210: 549-559, 2006.
58. Chada S, Mhashilkar AM, Liu Y, Nishikawa T, Bocangel D, Zheng M, Vorburger SA, Pataer A, Swisher SG, Ramesh R, Kawase K, Meyn RE and Hunt KK: *mda-7* gene transfer sensitizes breast carcinoma cells to chemotherapy, biologic therapies and radiotherapy: correlation with expression of bcl-2 family members. *Cancer Gene Ther* 13: 490-502, 2006.
59. Pataer A, Vorburger SA, Chada S, Balachandran S, Barber GN, Roth JA, Hunt KK and Swisher SG: Melanoma differentiation-associated gene-7 protein physically associates with the double-stranded RNA-activated protein kinase PKR. *Mol Ther* 11: 717-723, 2005.
60. Fawcett TW, Martindale JL, Guyton KZ, Hai T and Holbrook NJ: Complexes containing activating transcription factor (ATF)/cAMP-responsive-element-binding protein (CREB) interact with the CCAAT/enhancer-binding protein (C/EBP)-ATF composite site to regulate Gadd153 expression during the stress response. *Biochem J* 339: 135-141, 1999.
61. Yacoub A, Mitchell C, Lebedeva IV, Sarkar D, Su ZZ, McKinstry R, Gopalkrishnan RV, Grant S, Fisher PB and Dent P: *Mda-7* (IL-24) inhibits growth and enhances radiosensitivity of glioma cells *in vitro* via JNK signaling. *Cancer Biol Ther* 2: 347-353, 2003.
62. Kawabe S, Nishikawa T, Munshi A, Roth JA, Chada S and Meyn RE: Adenovirus-mediated *mda-7* gene expression radiosensitizes non-small cell lung cancer cells via TP53-independent mechanisms. *Mol Ther* 6: 637-644, 2002.
63. Mhashilkar AM, Stewart AL, Sieger K, Yang HY, Khimani AH, Ito I, Saito Y, Hunt KK, Grimm EA, Roth JA, Meyn RE, Ramesh R and Chada S: MDA-7 negatively regulates the beta-catenin and PI3K signaling pathways in breast and lung tumor cells. *Mol Ther* 8: 207-219, 2003.
64. Gopalan B, Litvak A, Sharma S, Mhashilkar AM, Chada S and Ramesh R: Activation of the Fas-FasL signaling pathway by MDA-7/IL-24 kills human ovarian cancer cells. *Cancer Res* 65: 3017-3024, 2005.
65. Emdad L, Sarkar D, Lebedeva IV, Su ZZ, Gupta P, Mahasreshthi PJ, Dent P, Curiel DT and Fisher PB: Ionizing radiation enhances adenoviral vector expressing *mda-7*/IL-24-mediated apoptosis in human ovarian cancer. *J Cell Physiol* 208: 298-306, 2006.
66. Leach JK, van Tuyle G, Lin PS, Schmidt-Ullrich R and Mikkelsen RB: Ionizing radiation-induced, mitochondria-dependent generation of reactive oxygen/nitrogen. *Cancer Res* 61: 3894-3901, 2001.
67. Leach JK, Black SM, Schmidt-Ullrich RK and Mikkelsen RB: Activation of constitutive nitric-oxide synthase activity is an early signaling event induced by ionizing radiation. *J Biol Chem* 277: 15400-15406, 2002.
68. Nishikawa T, Munshi A, Story MD, Ismail S, Stevens C, Chada S and Meyn RE: Adenoviral-mediated *mda-7* expression suppresses DNA repair capacity and radiosensitizes non-small-cell lung cancer cells. *Oncogene* 23: 7125-7131, 2004.
69. Su Z, Shi Y and Fisher PB: Cooperation between API and PEA3 sites within the progression elevated gene-3 (PEG-3) promoter regulate basal and differential expression of PEG-3 during progression of the oncogenic phenotype in transformed rat embryo cells. *Oncogene* 19: 3411-3421, 2000.
70. Su Z, Shi Y, Friedman R, Qiao L, McKinstry R, Hinman D, Dent P and Fisher PB: PEA3 sites within the progression elevated gene-3 (PEG-3) promoter and mitogen-activated protein kinase contribute to differential PEG-3 expression in Ha-*ras* and *v-raf* oncogene transformed rat embryo cells. *Nucleic Acids Res* 29: 1661-1671, 2001.
71. Su ZZ, Goldstein NI, Jiang H, Wang MN, Duigou GJ, Young CS and Fisher PB: *PEG-3*, a nontransforming cancer progression gene, is a positive regulator of cancer aggressiveness and angiogenesis. *Proc Natl Acad Sci USA* 96: 15115-15120, 1999.
72. Emdad L, Sarkar D, Su ZZ, Boukerche H, Bar-Eli M and Fisher PB: Progression elevated gene-3 (*PEG-3*) induces pleiotropic effects on tumor progression: modulation of genomic stability and invasion. *J Cell Physiol* 202: 135-146, 2005.
73. Sarkar D, Su ZZ, Vozhilla N, Park ES, Gupta P and Fisher PB: Dual cancer-specific targeting strategy cures primary and distant breast carcinomas in nude mice. *Proc Natl Acad Sci USA* 102: 14034-14039, 2005.
74. Sarkar D, Su ZZ, Vozhilla N, Park ES, Randolph A, Valerie K and Fisher PB: Targeted virus replication plus immunotherapy eradicates primary and distant pancreatic tumors in nude mice. *Cancer Res* 65: 9056-9063, 2005.

75. Chen J, Chada S, Mhashilkar A and Miano JM: Tumor suppressor MDA-7/IL-24 selectively inhibits vascular smooth muscle cell growth and migration. *Mol Ther* 8: 220-229, 2003.
76. Ramesh R, Ito I, Saito Y, Wu Z, Mhashikar AM, Wilson DR, Branch CD, Roth JA and Chada S: Local and systemic inhibition of lung tumor growth after nanoparticle-mediated *mda-7/IL-24* gene delivery. *DNA Cell Biol* 23: 850-857, 2004.
77. Welch DR, Bisi JE, Miller BE, Conaway D, Seftor EA, Yohem KH, Gilmore LB, Seftor RE, Nakajima M and Hendrix MJ: Characterization of a highly invasive and spontaneously metastatic human malignant melanoma cell line. *Int J Cancer* 47: 227-237, 1991.
78. Folkman J: The role of angiogenesis in tumor growth. *Semin Cancer Biol* 3: 65-71, 1992.
79. Folkman J: Angiogenesis in cancer, vascular, rheumatoid and other disease. *Nat Med* 1: 27-31, 1995.
80. Fidler IJ and Ellis LM: The implications of angiogenesis for the biology and therapy of cancer metastasis. *Cell* 79: 185-188, 1994.
81. Cristofanilli M, Charnsangavej C and Hortobagyi GN: Angiogenesis modulation in cancer research: novel clinical approaches. *Nat Rev Drug Discov* 1: 415-426, 2002.
82. Kerr DJ: Targeting angiogenesis in cancer: clinical development of bevacizumab. *Nat Clin Pract Oncol* 1: 39-43, 2004.
83. O'Reilly MS, Holmgren L, Shing Y, Chen C, Rosenthal RA, Moses M, Lane WS, Cao Y, Sage EH and Folkman J: Angiostatin: a novel angiogenesis inhibitor that mediates the suppression of metastases by a Lewis lung carcinoma. *Cell* 79: 315-328, 1994.
84. O'Reilly MS, Boehm T, Shing Y, Fukai N, Vasios G, Lane WS, Flynn E, Birkhead JR, Olsen BR and Folkman J: Endostatin: an endogenous inhibitor of angiogenesis and tumor growth. *Cell* 88: 277-285, 1997.
85. Shaheen RM, Davis DW, Liu W, Zebrowski BK, Wilson MR, Bucana CD, McConkey DJ, McMahon G and Ellis LM: Anti-angiogenic therapy targeting the tyrosine kinase receptor for vascular endothelial growth factor receptor inhibits the growth of colon cancer liver metastasis and induces tumor and endothelial cell apoptosis. *Cancer Res* 59: 5412-5416, 1999.
86. Moses MA, Sudhalter J and Langer R: Identification of an inhibitor of neovascularization from cartilage. *Science* 248: 1408-1410, 1990.
87. Laird AD, Vajkoczy P, Shawver LK, Thurnher A, Liang C, Mohammadi M, Schlessinger J, Ullrich A, Hubbard SR, Blake RA, Fong TA, Strawn LM, Sun L, Tang C, Hawtin R, Tang F, Shenoy N, Hirth KP, McMahon G and Cherrington S: U6668 is a potent antiangiogenic and antitumor agent that induces regression of established tumors. *Cancer Res* 60: 4152-4160, 2000.
88. Kumar CC, Malkowski M, Yin Z, Tanghetti E, Yaremko B, Nechuta T, Varner J, Liu M, Smith EM, Neustadt B, Presta M and Armstrong L: Inhibition of angiogenesis and tumor growth by SCH221153, a dual $\alpha(v)\beta_3$ and $\alpha(v)\beta_5$ integrin receptor antagonist. *Cancer Res* 61: 2232-2238, 2001.
89. Lingen MW, Polverini PJ and Bouck NP: Inhibition of squamous cell carcinoma angiogenesis by direct interaction of retinoic acid with endothelial cells. *Lab Invest* 74: 476-483, 1996.
90. Voest EE, Kenyon BM, O'Reilly MS, Truitt G, D'Amato RJ and Folkman J: Inhibition of angiogenesis *in vivo* by interleukin 12. *J Natl Cancer Inst* 87: 581-586, 1995.
91. Singh RK, Gutman M, Bucana CD, Sanchez R, Llansa N and Fidler IJ: Interferons alpha and beta down-regulate the expression of basic fibroblast growth factor in human carcinomas. *Proc Natl Acad Sci USA* 92: 4562-4566, 1995.
92. Ramesh R, Mhashilkar AM, Tanaka F, Saito Y, Branch CD, Sieger K, Mumm JB, Stewart AL, Boquoi A, Dumoutier L, Grimm EA, Renauld JC, Kotenko S, Chada S and Boquoi A: Melanoma differentiation-associated gene 7/interleukin (IL)-24 is a novel ligand that regulates angiogenesis via the IL-22 receptor. *Cancer Res* 63: 5105-5113, 2003.
93. Yu H and Jove R: The STATs of cancer - new molecular targets come of age. *Nat Rev Cancer* 4: 97-105, 2004.
94. Ramesh R, Ito I, Gopalan B, Saito Y, Mhashilkar AM and Chada S: Ectopic production of MDA-7/IL-24 inhibits invasion and migration of human lung cancer cells. *Mol Ther* 9: 510-518, 2004.
95. Tedjarati S, Baker CH, Apte S, Huang S, Wolf JK, Killion JJ and Fidler IJ: Synergistic therapy of human ovarian carcinoma implanted orthotopically in nude mice by optimal biological dose of pegylated interferon alpha combined with paclitaxel. *Clin Cancer Res* 8: 2413-2422, 2002.
96. Inoue K, Chikazawa M, Fukata S, Yoshikawa C and Shuin T: Frequent administration of angiogenesis inhibitor TNP-470 (AGM-1470) at an optimal biological dose inhibits tumor growth and metastasis of metastatic human transitional cell carcinoma in the urinary bladder. *Clin Cancer Res* 8: 2389-2398, 2002.
97. Hirata A, Ogawa S, Kometani T, Kuwano T, Naito S, Kuwano M and Ono M: ZD1839 (Iressa) induces antiangiogenic effects through inhibition of epidermal growth factor receptor tyrosine kinase. *Cancer Res* 62: 2554-2560, 2002.
98. Inoue S, Branch CD, Gallick GE, Chada S and Ramesh R: Inhibition of Src kinase activity by Ad-*mda7* suppresses vascular endothelial growth factor expression in prostate carcinoma cells. *Mol Ther* 12: 707-715, 2005.
99. Lebedeva IV, Sarkar D, Su ZZ, Gopalkrishnan RV, Athar M, Randolph A, Valerie K, Dent P and Fisher PB: Molecular target-based therapy of pancreatic cancer. *Cancer Res* 66: 2403-2413, 2006.
100. Madireddi MT, Dent P and Fisher PB: Regulation of *mda-7* gene expression during human melanoma differentiation. *Oncogene* 19: 1362-1368, 2000.
101. Madireddi MT, Dent P and Fisher PB: AP-1 and C/EBP transcription factors contribute to *mda-7* gene promoter activity during human melanoma differentiation. *J Cell Physiol* 185: 36-46, 2000.
102. Ekmekcioglu S, Ellerhorst JA, Mumm JB, Zheng M, Broemeling L, Prieto VG, Stewart AL, Mhashilkar AM, Chada S and Grimm EA: Negative association of melanoma differentiation-associated gene (*mda-7*) and inducible nitric oxide synthase (iNOS) in human melanoma: MDA-7 regulates iNOS expression in melanoma cells. *Mol Cancer Ther* 2: 9-17, 2003.
103. Schaefer G, Venkataraman C and Schindler U: Cutting edge: FISP (IL-4-induced secreted protein), a novel cytokine-like molecule secreted by Th2 cells. *J Immunol* 166: 5859-5863, 2001.
104. Ekmekcioglu S, Okcu MF, Colome-Grimmer MI, Owen-Schaub L, Buzaid AC and Grimm EA: Differential increase of Fas ligand expression on metastatic and thin or thick primary melanoma cells compared with interleukin-10. *Melanoma Res* 9: 261-272, 1999.
105. Garn H, Schmidt A, Grau V, Stumpf S, Kaufmann A, Becker M, Gemsa D and Siese A: IL-24 is expressed by rat and human macrophages. *Immunobiology* 205: 321-334, 2002.
106. Poindexter NJ, Walch ET, Chada S and Grimm EA: Cytokine induction of interleukin-24 in human peripheral blood mononuclear cells. *J Leukoc Biol* 78: 745-752, 2005.
107. Aggarwal S, Takada Y, Mhashilkar AM, Sieger K, Chada S and Aggarwal BB: Melanoma differentiation-associated gene-7/IL-24 gene enhances NF-kappa B activation and suppresses apoptosis induced by TNF. *J Immunol* 173: 4368-4376, 2004.
108. Kumar A, Haque J, Lacoste J, Hiscott J and Williams BR: Double-stranded RNA-dependent protein kinase activates transcription factor NF-kappa B by phosphorylating I kappa B. *Proc Natl Acad Sci USA* 91: 6288-6292, 1994.
109. Bonnet MC, Weil R, Dam E, Hovanessian AG and Meurs EF: PKR stimulates NF-kappaB irrespective of its kinase function by interacting with the I kappa B kinase complex. *Mol Cell Biol* 20: 4532-4542, 2000.
110. Ishii T, Kwon H, Hiscott J, Mosialos G and Koromilas AE: Activation of the I kappa B alpha kinase (IKK) complex by double-stranded RNA-binding defective and catalytic inactive mutants of the interferon-inducible protein kinase PKR. *Oncogene* 20: 1900-1912, 2001.
111. Miyahara R, Banerjee S, Kawano K, Efferson C, Tsuda N, Miyahara Y, Ioannides CG, Chada S and Ramesh R: Melanoma differentiation-associated gene-7 (*mda-7*)/interleukin (IL)-24 induces anticancer immunity in a syngeneic murine model. *Cancer Gene Ther* 13: 753-761, 2006.
112. Bardeesy N and De Pinho RA: Pancreatic cancer biology and genetics. *Nat Rev Cancer* 2: 897-909, 2002.
113. Su ZZ, Shi Y and Fisher PB: Subtraction hybridization identifies a transformation progression-associated gene *PEG-3* with sequence homology to a growth arrest and DNA damage-inducible gene. *Proc Natl Acad Sci USA* 94: 9125-9130, 1997.
114. Smyth MJ, Hayakawa Y, Takeda K and Yagita H: New aspects of natural-killer-cell surveillance and therapy of cancer. *Nat Rev Cancer* 2: 850-861, 2002.
115. Zheng M, Bocangel D, Doneske B, Mhashilkar A, Ramesh R, Hunt KK, Ekmekcioglu S, Sutton RB, Poindexter N, Grimm EA and Chada S: Human interleukin 24 (MDA-7/IL-24) protein kills breast cancer cells via the IL-20 receptor and is antagonized by IL-10. *Cancer Immunol Immunother* 56: 205-215, 2007.

116. Saeki T, Mhashilkar A, Chada S, Branch C, Roth JA and Ramesh R: Tumor-suppressive effects by adenovirus-mediated *mda-7* gene transfer in non-small cell lung cancer cell *in vitro*. *Gene Ther* 7: 2051-2057, 2000.
117. McKenzie T, Liu Y, Fanale M, Swisher SG, Chada S and Hunt KK: Combination therapy of Ad-*mda7* and trastuzumab increases cell death in *Her-2/neu*-overexpressing breast cancer cells. *Surgery* 136: 437-442, 2004.
118. Oida Y, Gopalan B, Miyahara R, Inoue S, Branch CD, Mhashilkar AM, Lin E, Bekele BN, Roth JA, Chada S and Ramesh R: Sulindac enhances adenoviral vector expressing *mda-7/IL-24*-mediated apoptosis in human lung cancer. *Mol Cancer Ther* 4: 291-304, 2005.
119. Swisher SG, Roth JA, Nemunaitis J, *et al*: Adenovirus-mediated p53 gene transfer in advanced non-small-cell lung cancer. *J Natl Cancer Inst* 91: 763-771, 1999.
120. Lang FF, Bruner JM, Fuller GN, Aldape K, Prados MD, Chang S, Berger MS, McDermott MW, Kunwar SM, Junck LR, Chandler W, Zwiebel JA, Kaplan RS and Yung WK: Phase I trial of adenovirus-mediated p53 gene therapy for recurrent glioma: biological and clinical results. *J Clin Oncol* 21: 2508-2518, 2003.
121. Bocangel D, Zheng M, Mhashilkar A, Liu Y, Ramesh R, Hunt KK and Chada S: Combinatorial synergy induced by adenoviral-mediated *mda-7* and Herceptin in *Her-2+* breast cancer cells. *Cancer Gene Ther* 13: 958-968, 2006.
122. Zhao L, Gu J, Dong A, Zhang Y, Zhong L, He L, Wang Y, Zhang J, Zhang Z, Huiwang J, Qian Q, Qian C and Liu X: Potent antitumor activity of oncolytic adenovirus expressing *mda-7/IL-24* for colorectal cancer. *Hum Gene Ther* 16: 845-858, 2005.
123. Sarkar D, Lebedeva IV, Su ZZ, Park ES, Chatman L, Vozhilla N, Dent P, Curiel DT and Fisher PB: Eradication of therapy resistant human prostate tumors using a cancer terminator virus. *Cancer Res* (In press).
124. Zerbini LF, Czibere A, Wang Y, *et al*: A novel pathway involving *mda-7/IL-24* mediates NSAID induced apoptosis and growth arrest of cancer cells. *Cancer Res* 66: 11922-11931, 2006.